

Instituto Juan March de Estudios e Investigaciones

153

CENTRO DE REUNIONES INTERNACIONALES SOBRE BIOLOGÍA

Workshop on

The Dynamics of Morphogenesis: Regulation of Cell and Tissue Movements in Development

Organized by

C. D. Stern and M. A. Nieto

C. Birchmeier

M. Bronner-Fraser

J. Casanova

S. Eaton

S. E. Fraser

R. Keller

M. A. Krasnow

R. Lehmann

O. Marín

P. Martin

D. McClay

D. J. Montell

M. A. Nieto

L. Solnica-Krezel

C. D. Stern

G. Tear

S. W. Wilson

L. Wolpert

M. Zernicka-Goetz

JM

153

Wor



Instituto Juan March de Estudios e Investigaciones

153 CENTRO DE REUNIONES INTERNACIONALES SOBRE BIOLOGÍA

Workshop on
The Dynamics of Morphogenesis: Regulation
of Cell and Tissue Movements in Development

Organized by

C. D. Stern and M. A. Nieto

C. Birchmeier
M. Bronner-Fraser
J. Casanova
S. Eaton
S. E. Fraser
R. Keller
M. A. Krasnow
R. Lehmann
O. Marín
P. Martin



D. McClay
D. J. Montell
M. A. Nieto
L. Solnica-Krezel
C. D. Stern
G. Tear
S. W. Wilson
L. Wolpert
M. Zernicka-Goetz

*The lectures summarized in this publication
were presented by their authors at a workshop
held on the 12th through the 14th of May, 2003,
at the Instituto Juan March.*

Depósito legal: M- 28686 / 2003

Impresión: Ediciones Peninsular. Tomelloso, 27. 28026 Madrid

INDEX

	PAGE
Introduction: Claudio D. Stern and M. Angela Nieto.....	7
Session 1: Morphogenesis and embryo patterning	
Chair: Claudio D. Stern.....	11
Scott E. Fraser: Imaging cell and tissue migrations in the developing embryo.....	13
David McClay: Mesenchymal migration and cell fate regulation in sea urchin embryo.....	15
Magdalena Zernicka-Goetz: Morphogenetic movements that establish the anterior-posterior axis after implantation respect the bilateral symmetry of the embryo rather than the uterine axis.....	16
Ray Keller: The cell motility, biomechanics, and the self-deforming "skeleton" of the amphibian gastrula.....	18
Short talk:	
Enrique Amaya: FGF signalling and the control of gastrulation movements in <i>Xenopus</i>	19
Session 2: Integrating cell behaviours	
Chair: Scott E. Fraser.....	21
Lilianna Solnica-Krezel: Regulation of cell movements during zebrafish gastrulation and tail formation.....	23
Suzanne Eaton: Control of cortical reorganization during planar polarization in <i>Drosophila</i>	25
Short talk:	
Eduard Batlle: Beta-catenin and TCF mediate cell positioning in the intestinal epithelium by controlling the expression of EphB/EphrinB.....	26
Mark A. Krasnow: Genetic and physiological control of branching morphogenesis.....	27
Claudio D. Stern: How does the embryo coordinate cell movements and cell fate? Integrating different functions of FGF signalling.....	29

	PAGE
Session 3: Directing cell migration	
Chair: Ruth Lehmann	31
Short talks:	
Enrique Martín-Blanco: The eversion of <i>Drosophila</i> imaginal discs requires a JNK-regulated change in the adhesive and motile properties of peripodial cells.....	33
David Garrod: Regulation of cell-cell adhesion in wound edge epithelium by protein kinase C alpha.....	34
Ruth Lehmann: Early germline development in <i>Drosophila</i>	35
Jordi Casanova: Cell migration and morphogenesis of the <i>Drosophila</i> tracheal system.....	37
Denise J. Montell: Genetic control of cell migration in <i>Drosophila</i>	39
Session 4: Cell movements and the developing nervous system	
Chair: M. Angela Nieto	41
Marianne Bronner-Fraser: Molecular analysis of neural crest formation.....	43
Stephen W. Wilson: Local tissue interactions across the dorsal midline underlie morphogenesis of asymmetric forebrain nuclei.....	44
Guy Tear: Axon migration at the midline of the <i>Drosophila</i> central nervous system.....	45
Oscar Marín: Molecular mechanisms of cell migration in the telencephalon.....	47
Short talks:	
Richard J. T. Wingate: Hoxa2 overexpression in rhombomere 1 reveals rhombic lip autonomous patterning of tangential migration.....	48
Domingos Henrique: Establishment of cell polarity in the chick.....	49
Session 5: Cell movements, development and disease	
Chair: Marianne Bronner-Fraser	51
M. Angela Nieto: The migratory phenotype of Snail-expressing cells.....	53
Paul Martin: Parallels between morphogenesis and wound healing in embryos..	54

	PAGE
Carmen Birchmeier: Genes that control cell migration during the mouse embryo.....	56
Lewis Wolpert: Conclusions and future prospects.	
POSTERS.....	57
Jeroen Bakkers: Hyaluronan Synthase 2 is required for dorsal migration of lateral mesodermal cells during zebrafish gastrulation.....	59
Geert Berx: Analysis of the genetic program of epithelial cell plasticity.....	60
Paola Bovolenta: Requirement of Sfrp1 in eye anlage specification in medaka embryos.....	61
Tal Burstyn-Cohen: Association between the cell cycle and neural crest delamination through specific regulation of G1/S transition.....	62
Jérôme Collignon: New insights into the initiation of gastrulation in the mouse embryo.....	63
Miguel L. Concha: Dynamics of cell rearrangement in early zebrafish gastrulation.....	64
José Luis de la Pompa: Notch lateral activation promotes epithelial-mesenchymal transitions during heart development and neoplastic transformation	65
Peter Duchek: Guidance of border cell migration by the Drosophila EGFR and PDGF/VEGF receptor orthologs.....	66
Corinne Houart: Regulation of patterning and morphology in the zebrafish embryonic forebrain.....	67
Michel Kerszberg: Modeling neurulation: computer simulations of genetic networks, cell interactions, mechanics and motion.....	68
Derek C. Radisky: Defining the molecular determinants of epimorphin/syntaxin-2-mediated mammary epithelial morphogenesis.....	69
Masazumi Tada: Prickle1 regulates cell movements during gastrulation and neuronal migration in zebrafish.....	70
Ana Teresa Tavares: Transcriptional regulation of Cerberus during embryonic development.....	71

	PAGE
Marie-Luce Vignais: TGF β -dependent regulation of trophoblast invasion during placental implantation.....	72
Kris Vleminckx: Functional interaction of p120catenin with the Rho family of small GTPases during early <i>Xenopus</i> morphogenesis.....	73
LIST OF INVITED SPEAKERS.....	75
LIST OF PARTICIPANTS.....	77

Introduction
Claudio D. Stern and M. Angela Nieto

The cellular events that direct embryonic development have broadly been subdivided into three kinds of processes: fate allocation, pattern formation and morphogenesis. *Fates* are allocated to cells through a combination of their lineage history (through cytoplasmic and nuclear determinants of fate) and cell interactions (“inductions”). *Pattern formation* is the set of processes that shape a more or less uniform field of cells by assigning different fates to cells according to their position with respect to their neighbours. *Morphogenesis* is also a set of processes that generate form, but here the main driving forces are cell movements and tissue reorganizations rather than fate allocation. To illustrate the difference: an example of pattern formation is the generation of the periodic mosaic of ommatidia in the compound eye of *Drosophila*; an example of a morphogenetic process is gastrulation, where cells arrange themselves into three layers which will then behave differently.

In the last few decades considerable effort has been spent in understanding how cells are assigned different fates and how pattern formation occurs in systems like the *Drosophila* eye and the vertebrate limb. Morphogenetic processes, on the other hand, have received comparatively little attention. Genetic and molecular analyses have concentrated on elucidating *pathways*, but very little is known about *how these pathways are integrated in space and time*. Interest is now turning to try to understand the processes that orchestrate these dynamics of cell behaviour. This applies not only to embryonic development, but also to the adult and to pathological processes such as cancer, where invasive behaviour can be considered a form of morphogenesis that profoundly influences the morbidity of the disease.

This meeting aimed to explore the current status of this field, at a time when many laboratories are starting to acknowledge that no solid understanding of development or pathology can be gained until our view can also include an understanding of how gene expression and cell behaviour are integrated in space and time.

Some of the questions are:

- We already understand something about the *mechanics* of cell movement, but what signalling cues control the directed migration of cells?
- Is there directed migration, or just assortments of permissive, repulsive and attractive sites? How general a mechanism is chemotaxis?
- To date, several protein families have been implicated in signalling outside the nervous system (including FGFs, Wnts and HGF/SF), but different signalling proteins have been shown to guide axon navigation in the CNS and PNS (including netrins, semaphorins, collapsins, Nogo, Ephrins/Eph receptors, etc.). Is this a true difference? If so, how/why did different mechanisms evolve to regulate what seems like the same process in different parts of the same organisms?
- As cells migrate, they respond to local inductive/repressive cues from their neighbours which change their gene expression profiles and consequently alter their behaviour. How are the many signals they encounter integrated? For example, as prospective mesoderm cells approach the organizer region of the vertebrate embryo, they start to express organizer-specific genes, and then turn them off as they leave the region.

When do they receive the signals that activate/repress the expression of specific genes, and how much of the timing of this is controlled by combinations of regulatory genetic elements, and how much by the precise positioning and physical range of the inductive/repressive signals?

- How does the signalling machinery result in directional movement? Can cells really sense concentration differences between one of their sides and the other?
- Do other cellular asymmetries (such as asymmetric cell division, epithelial polarity) involve similar directional cues, which direct the migration of intracellular organelles?
- How many of the signalling systems involved in development are also implicated in wound healing and cancer invasion/metastasis? Why do embryos heal their wounds without scars, while adults always scar? Why are some tumours invasive and some not?

Of course, a two-and-a-half-day meeting could not possibly answer all of these important questions. However, by focusing on the major current “model” processes (gastrulation, neural crest cell migration, germ cell migration, tubulogenesis and branching morphogenesis, epidermal wound healing and cancer), and “model organisms” (*Drosophila*, *C. elegans*, Sea Urchin, zebrafish, *Xenopus*, chick and mouse), we can obtain an inkling of the great diversity of mechanisms that exist to coordinate complex cell behaviours in time and space. Perhaps one critical conclusion that was achieved during the meeting was that biology itself is extremely diverse and that the whole concept of “model” processes and systems can lead to oversimplifications and to false generalisations. The principles of evolution and development will be best understood by exploiting the advantages of each system, and by opportunities such as this one, to exchange views and experiences between those that have been thinking very deeply about their favourite organism and biological event.

The organisers,
May 2003

**Session 1: Morphogenesis and
embryo patterning
Chair: Claudio D. Stern**

Imaging cell and tissue migrations in the developing embryo

Scott E. Fraser

Beckman Institute, California Institute of Technology, Pasadena, CA 91125 USA

The explosion of progress in the fields of cell biology, biochemistry, and molecular biology has offered unprecedented knowledge of the components involved in embryonic development. The dramatic progress of these reductionistic approaches poses the challenge of integrating this knowledge into an understanding of the underlying mechanics. The classic publications in the field of experimental embryology illustrate the power of describing cell behavior (cf. lineages, movements) and perturbing the embryo to test hypotheses of the underlying mechanisms. Advanced imaging techniques offer an important stepping stone between these disparate approaches, permitting questions about cellular and molecular events to be posed in the most relevant setting of the intact embryo.

There are two major limitations of any approach based on fluorescence in living embryos. First, the image degrades dramatically as the microscope is focused below the surface layers of the embryo. This results from increased optical aberrations in the objective lenses and increased light scattering as the light passes through a significant pathlength of tissue. Second, the fluorochromes are bleached by irradiation with the exciting light and the by-products of this bleaching can be toxic. Two-photon laser scanning microscopy (TPLSM) offers a means to minimize these concerns. TPLSM uses an intense red or infra-red ultra-fast laser to excite fluorochromes in the UV or visible wavelength range through the concurrent absorption of two photons. Because the statistics of two photons being absorbed depends on the square of the intensity and the intensity drops by the square of distance from the focal plane, the technique selectively excites fluorochromes at the focal plane. Light scattering and optical aberrations decrease the probability of the excitation, but decrease the resolution in the image much less than in other light based microscopies. Based on its performance to date, two-photon microscopy may offer the best means for detecting fluorescent labels in intact tissues. Multispectral approaches, in which the entire spectrum is taken of the emitted light from each pixel, offers a potential solution, as the spectral data can be decomposed into its component parts by simple mathematics. Fluorochromes as similar as GFP and fluorescein can be separated unambiguously, and even small amounts of FRET can be detected by our approach (now available as the Zeiss LSM-510 Meta).

In systems in which light-based imaging is problematic, we are employing microscopic MRI. In MRI, radio frequency energy is used to excite the protons of the water, which generates no toxic by-products. Spatial resolution is created by imposing gradient magnetic fields on the specimen, thereby making it possible to encode the signals from individual volume elements (voxels) by their resonant frequency and phase. By increasing the magnitudes of the

static and gradient magnetic fields, and by improving the electronics, it has become possible to increase the resolution of MRI from the 1mm voxels of a clinical instrument to $\sim 10\mu\text{m}$. This approach has the promise of making imaging analyses possible in the systems with limited access to the embryos (e.g. mouse) or in which light scattering renders deep structures invisible (e.g. frog).

Here, MRI microscopy will be used to follow the fates and motions of the amphibian Spemann organizer, originally defined by its ability to induce and organize a secondary body axis when grafted to the future ventral side of an embryo. The Spemann organizer plays a central role in directing a subregion of the surface ectoderm to adopt a neural fate. Recent molecular and experimental embryology studies have defined a set of molecular events involved in the establishment and the function of the Spemann organizer and have shown that the lessons learned from the amphibian Spemann organizer have important parallels in a variety of species. A secondary body axis forms when the "organizer" region of the early embryo is grafted to an ectopic site in species ranging from fish to chicken and mouse. Similarly, many of the molecular correlates of organizer function in the frog have been found in a variety of species. As a result, many findings in other species are reported in the context of the amphibian organizer. Despite the frog serving as the standard by which results in other species are cast, significant open questions remain as to the nature of neural induction in the amphibian. Two opposed pathways of neural induction remain viable: vertical signaling models, in which the mesoderm and endoderm (mesendoderm) involute around the blastopore lip and then induce the overlying ectoderm; planar signaling models argue that the critical inductive interactions take place between mesendoderm and ectoderm in the plane of the embryonic surface before the involution motions of gastrulation. Microscopic magnetic resonance imaging allows us to follow cell movements and contacts in the Spemann organizer region before and during gastrulation. Key events such as a vegetal rotation and epiboly bring surface ectoderm and mesendoderm into contact long before the outward signs of gastrulation. A surprising resolution of the debate between vertical and planar signaling models is the finding that the axial mesoderm is internally localized and in vertical contact with much of the future neurectoderm throughout early development. These observations are consistent with the direction of signaling proposed by vertical models and the early timing of the signaling proposed by planar models.

References:

Harland R and Gerhart J (1997): "Formation and function of Spemann's organizer" *Annu Rev Cell Dev Biol* 13:611-667

Mesenchymal migration and cell fate regulation in the sea urchin embryo

David McClay

Department of Biology, Duke University, Durham, NC 27708. USA

Mesenchymal ingression temporally precedes invagination of the archenteron in the sea urchin embryo. Primary mesenchyme cells undergo an epithelial to mesenchymal transition as they invade the blastocoel. The immediate trigger for PMC ingression is not known but is downstream of several identified transcription factors. In the process of ingression the cells endocytose their epithelial membranes and the adhesive proteins contained therein. At the same time the PMCs insert new membranes containing new adhesion molecules onto the surface. This allows for a rapid deployment of the mesenchymal adhesive phenotype. Pre-PMCs prepare for this rapid transition for many hours: a gene regulatory network begins to specify micromeres at fourth cleavage. By 8th-10th cleavage the specified micromeres begin to store proteins in vesicles for recruitment at ingression. In a distinct specification sequence secondary mesenchyme cells begin their specification sequence, also at 4th cleavage, and are separated from general endomesoderm by Notch signaling by 7th-9th cleavage. SMCs both ingress and lead the invagination of the archenteron. The trigger for initiation of archenteron invagination appears to be through activation of Rho kinase. The invagination of endoderm into the archenteron requires a wave of *brachyury* expression. That expression is necessary for many cytoskeletal and motility molecules involved in convergent extension morphogenesis of the gut. The wave of *brachyury* expression is controlled, at least in part, by a *gatae* activator, and a *foxA* repressor.

Morphogenetic movements that establish the anterior-posterior axis after implantation respect the bilateral symmetry of the embryo rather than the uterine axis

Magdalena Zernicka-Goetz and Daniel Mesnard

Wellcome Trust/Cancer Research Institute
Tennis Court Road
Cambridge CB2 1QR, UK

Although the anterior-posterior axis of the mouse embryo becomes explicit morphologically at embryonic day (E) 6.5, the first molecular signs of the anterior-posterior polarity after implantation are the asymmetric expression of several genes along the proximo-distal axis of the egg cylinder (for review see Robertson and Beddington, 1999). Lineage tracing has shown that this proximo-distal polarity translates back to the axis of bilateral symmetry of the blastocyst, which in turn corresponds to the polarity of the fertilised egg (Weber et al., 1999; for review see Zernicka-Goetz, 2002). It appears that asymmetric cell movements between the blastocyst and gastrula stages are key to development of this polarity (Thomas and Beddington, 1996, Weber et al, 1999). The extent to which these movements reflect differential growth of the egg cylinder, changes in its shape or cell migration is unknown. Also unknown is the relationship of these movements to the organisation of the post-implantation embryo with reference to the axes of the uterus. To gain insight into the transformations of the post-implantation embryo, we have analysed the dynamics of its morphogenetic changes from implantation until the initiation of gastrulation with particular reference to the expression pattern of Cer-1-GFP, a marker of the future anterior (Belo et al, 1997). At implantation the bilateral symmetry of the embryo is aligned with the long axis of the uterus. This relationship is lost between E5.5-E6.25 but then re-gained in a perpendicular orientation at the time of gastrulation. The asymmetric distal to anterior movement of the visceral endoderm cells that is initiated at E5.5 relates to the embryo's axes of symmetry and not to the axes of the uterus. Unexpectedly this movement occurs predominantly along the short and not the long axis of the embryo. Subsequent alignment of the embryo's anterior-posterior axis with the axis of the uterus occurs through limiting the expression domain of anterior markers. We suggest this is facilitated by rotation of the egg cylinder within extra-embryonic tissues or changes in shape of the egg cylinder itself. An understanding of these processes has to be accommodated within this new perception of anatomical dynamics during implantation.

References:

- Beddington RS, Robertson EJ. Axis development and early asymmetry in mammals. (1999). *Cell* 22;96:195-209.
- Belo, J.A., Bouwmeester, T., Leyns, L., Kertesz, N., Gallo, M., Follettie, M. and De Robertis, E.M. (1997). *Development* 68, 45-57.
- Thomas P. Beddington R. Anterior primitive endoderm may be responsible for patterning the anterior neural plate in the mouse embryo. (1996). *Curr Biol*6:1487-96.
- Weber, R., Wianny, F., Evans, M., Pedersen, R. and Zernicka-Goetz, M. (1999). *Development* 126, 5591-5598.
- Zernicka-Goetz, M. (2002). Patterning of the embryo: the first spatial decisions in the life of a mouse. *Development* 129, 815-829.

The cell motility, biomechanics, and the self-deforming “skeleton” of the amphibian gastrula

Ray Keller

Department of Biology, University of Virginia

Involution of the mesoderm, blastopore closure, and axis elongation during gastrulation and neurulation of *Xenopus* are due largely to convergence and extension of the axial and paraxial mesoderm and posterior neural tissue. During these movements, cells actively intercalate between one another along the mediolateral axis to form a narrower, longer array, but at the same time, they collectively form a stiff beam and exert a pushing force. New evidence concerning polarized protrusive activity and extracellular matrix supports a cell-traction/cell-substrate mechanism that allows self-deforming tissues such as these to actively rearrange their cells but at the same time form a self-supporting, force-generating morphogenic machine that can also serve as a dynamic “skeletons” of the embryo. Understanding how self-deforming, force-producing tissues function is important for understanding a number of morphogenic movements. Other work shows that in addition to cell intercalation, urodele amphibian embryos use an epithelial mesenchymal transition, expressed in the form of a bilateral “primitive streak”, to generate the forces produced by cell intercalation alone in *Xenopus*. These results argue that biomechanical strategy is the key parameter in evolution of gastrulation mechanisms.

FGF signalling and the control of gastrulation movements in *Xenopus*

Jeremy M. Sivak, Stephen L. Nutt and Enrique Amaya

Signal transduction through the FGF receptor is essential for the specification of the vertebrate body plan. Blocking the FGF pathway in early *Xenopus* embryos inhibits mesoderm formation and results in truncation of the anterior-posterior axis. We have previously shown that *Xenopus sprouty2* is an intracellular antagonist of FGF-dependent calcium signalling. In addition we have shown that *Xsprouty2* inhibits convergent extension movements and have suggested that there are at least two distinct FGF-dependent signal transduction pathways: a Sprouty insensitive Ras/MAPK pathway required for the transcription of most mesodermal genes, and a Sprouty sensitive pathway required for co-ordination of cellular morphogenesis. We have recently identified additional antagonists of FGF signalling, including *Xsprouty1*, *Xspred1*, *Xspred2* and *XSef* and are currently studying their role during mesoderm formation and morphogenesis.

Unexpectedly, preliminary results from mis-expression and morpholino experiments suggest that the functions of these proteins diverge during *Xenopus* gastrulation. These differences support a model in which the various FGF signalling antagonists may cooperate to modulate cell behaviours and mesoderm formation. Our eventual aim is to understand how FGF signalling is controlled in space and time resulting in the co-ordination of mesoderm formation and cell movements during gastrulation.

Session 2: Integrating cell behaviours
Chair: Scott E. Fraser

Regulation of cell movements during zebrafish gastrulation and tail formation

Lilianna Solnica-Krezel, Florence Marlow, Chunyue Yin and Encina M. Gonzalez

Department of Biological Sciences, Vanderbilt University, Nashville, TN

During the vertebrate gastrulation movements of convergence and extension (C&E) the entire embryo and most organ rudiments narrow along the mediolateral axis while extending their anterior-posterior dimension (Solnica-Krezel and Cooper, 2002). In zebrafish C&E movements are driven by directed cell migration and intercalation of mediolaterally polarized cells and require non-canonical Wnt signaling and are regulated by ventral to dorsal gradient of Bmp activity (Myers et al., 2002a; Myers et al., 2002b). We have previously identified a glypican *Knypek* (Topczewski et al., 2001), membrane protein *Trilobite/Strabismus* (Jessen et al., 2002) and Rho kinase alpha (*Rok2*) (Marlow et al., 2002) as components of this pathway.

In other systems, Rho kinase impacts cell behaviors and cytoskeleton by phosphorylating Myosin regulatory light chain (*Mrlc*). We have cloned two zebrafish *mrlc* genes and showed that their transcripts and encoded proteins are present ubiquitously during early zebrafish embryogenesis. Overexpression of a phosphomimetic, constitutively active form of *Mrlc* disrupts C&E, phenocopying non-canonical Wnt signaling mutants. Epistatic analyses place *Mrlc* downstream of Wnt 11 in regulation of C&E. Impaired dorsal convergence movements in embryos overexpressing are associated with defective cell polarity and protrusive activity.

Tail morphogenesis entails both continuation of CE movements, as well as unique movements like subduction. We tested functional interactions between components of the non-canonical Wnt signaling pathway and a transcription factor *Notail* (Brachyury). Mutations in the *knypek* gene, encoding a glypican, and *pipetail* encoding Wnt5, impair CE movements resulting in shortened trunk and tail. *notail* mutants exhibit defective notochord and tail truncations. *kny;ntl* and *ppt;ntl* double mutants show similar, synergistic phenotypes characterized by truncation of posterior trunk and tail, but relatively normal head. Gene expression studies reveal that this synergistic phenotype is not due to defective specification of caudal tissues. Furthermore, neither decreased cell proliferation nor excess cell death can account for body shortening of double mutants. Tracing movements of cell populations in vivo revealed that defects in several types of cell movements contribute to posterior body shortening in double mutants including: extension, convergence and subduction. Hence, the non-canonical Wnt signaling cooperates with *Notail* to regulate morphogenetic movements that shape trunk and tail in vertebrate embryos. Our work initiates the genetic dissection of posterior body morphogenesis and links genes to specific tail-forming movements. Moreover, we provide genetic evidence for the notion that posterior body development entails a continuation of mechanisms that operate during gastrulation together with mechanisms unique to posterior body.

References:

- Jessen, J. R., Topczewski, J., Bingham, S., Sepich, D. S., Marlow, F., Chandrasekhar, A., and Solnica-Krezel, L. (2002). Zebrafish trilobite identifies new roles for Strabismus in gastrulation and neuronal movements. *Nat Cell Biol* 4, 610-615.
- Marlow, F., Topczewski, J., Sepich, D. S., and Solnica-Krezel, L. (2002). Zebrafish Rho Kinase 2 acts downstream of Wnt11 to mediate cell polarity and effective convergence and extension movements. *Curr Biol* 12.
- Myers, D., Sepich, D. S., and Solnica-Krezel, L. (2002a). BMP activity gradient regulates convergent extension during zebrafish gastrulation. *Developmental Biology* 243, 81-98.
- Myers, D. C., Sepich, D. S., and Solnica-Krezel, L. (2002b). Convergence and extension in vertebrate gastrulae: cell movements according to or in search of identity? *Trends Genet* 18, 447-455.
- Solnica-Krezel, L., and Cooper, M. S. (2002). Cellular and genetic mechanisms of convergence and extension. *Results Probl Cell Differ* 40, 136-165.
- Topczewski, J., Sepich, D. S., Myers, D. C., Walker, C., Amores, A., Lele, Z., Hammerschmidt, M., Postlethwait, J., and Solnica-Krezel, L. (2001). The zebrafish glypican knypek controls cell polarity during gastrulation movements of convergent extension. *Dev Cell* 1, 251-264.

Control of cortical reorganization during planar polarization in *Drosophila*

Suzanne Eaton

Max Planck Institute of Molecular Cell Biology and Genetics, Dresden

A critical step in the organization of epithelial tissues is planar polarization, that is, the development of intracellular polarity along an axis within the plane of the epithelium. The coordinate beating of cilia in the oviduct and the alignment of stereocilia bundles in sensory hair cells are two striking examples of planar polarization. The alignment of hairs and bristles in the insect cuticle is one of the best-studied examples of the process. How is this polarity generated and coordinated with the overall shape of the tissue? Over the last twenty years, genetic analysis has identified a class of 6 "tissue polarity" mutations that disrupt the planar polarity of the cuticle secreting epithelial cells and produce disordered arrays of hairs and bristles [Gubb, 1993 #28]. In the last three years, analysis of the proteins encoded by these genes has shown that they organize proximal-distal cortical polarity [Adler, 2001 #359]. The distribution of these proteins is initially uniform around the junctional region of each cell. Approximately 10 hours before hairs form, they become asymmetrically distributed and segregate into proximal and distal domains with different compositions. The process is highly cooperative and polarization fails if any one of the proteins is missing. The proximal-distal cortical domains have the striking ability to propagate their polarity from cell to cell [Usui, 1999 #318; Feiguin, 2001 #403; Tree, 2002 #404]. Our lab is interested in the cell biological mechanisms underlying this fascinating process. Does it involve polarized delivery of membrane proteins? Polarized endocytosis or recycling? Polarized cytoskeletal linkage?

Since these mechanisms are basic to all cells, it is likely that mutating the genes that control them would cause early lethality or even cell lethality. To find such genes, we performed an EP overexpression screen. One interesting player we identified in this screen was *widerborst* [Hannus, 2002 #390]. Widerborst activity is absolutely required for cortical polarization; in its absence, the cortical domain proteins accumulate uniformly around the cell at a high level. Interestingly, Widerborst does not localize to the cortex itself. Instead, it is found on the distal side of a planar microtubule web that lies at the level of apical junctions. Although these microtubules do not appear to have any structural polarity, the localization of Widerborst suggests that they might be functionally polarized. Widerborst is a B' regulatory subunit of Protein Phosphatase 2A and presumably acts by targeting the catalytic subunit of the enzyme to a particular substrate on the distal microtubule web. We are performing a two-hybrid screen to find the target(s) dephosphorylated by Widerborst.

What role might the planar microtubule web play in cortical polarization? Microtubules often act as tracks for the movement of intracellular membrane compartments. With this in mind, we are investigating the role of secretion, endocytosis and recycling in polarizing the distribution of cortical proteins. Our data indicate that endocytosis is essential for this process.

Beta-catenin and TCF mediate cell positioning in the intestinal epithelium by controlling the expression of EphB/EphrinB

Eduard Batlle & Hans Clevers

Cell renewal, lineage commitment and cell differentiation in the mammalian intestinal epithelium occur throughout postnatal life. These processes are intimately coupled to cell migration in a spatially organized manner. In the small intestine, the progeny of stem cells migrate in precise patterns. Absorptive, enteroendocrine, and goblet cells migrate toward the villus while Paneth cells occupy the bottom of the crypts. We have found that in the intestinal epithelium beta-catenin and TCF couple proliferation and differentiation to the sorting of cell populations by inversely controlling the expression of the EphB2/EphB3 receptors and their ligand ephrin-B1 along the crypt-villus axis. In EphB2/EphB3 null mice, the proliferative and differentiated populations intermingle. In adult EphB3^{-/-} mice, Paneth cells do not follow their downward migratory path, but scatter along crypt and villus. We provide evidence that the proliferation/differentiation switch and the sorting of cell populations represent two independent outputs on the Wnt signaling pathway in the intestinal epithelium. Moreover, we show that during the first stages of intestinal tumorigenesis, Eph and ephrin expressing cells are further compartmentalized suggesting an additional outcome of the genetic program driven by beta-catenin/TCF in colorectal cancer.

References:

1. BATLLE, E., HENDERSON, J.T., BEGHTEL, H., VAN DEN BORN, M.M., SANCIO, E., HULS, G., MEELDIJK, J., ROBERTSON, J., VAN DE WETERING, M., PAWSON, T., CLEVERS, H. beta-Catenin and TCF Mediate Cell Positioning in the Intestinal Epithelium by Controlling the Expression of EphB/EphrinB. *Cell*. 2002; 111 (2): 251-263.
2. VAN DE WETERING, M., SANCIO, E., VERWEIJ, C., DE LAU, W., Oving, I., HURLSTONE, A., VAN DER HORN, K., BATLLE, E., COUDREUSE, D., HARAMIS, A.P., TJON-PON-FONG, M., MOERER, P., VAN DEN BORN, M., SOETE, G., PALS, S., EILERS, M., MEDEMA, R., CLEVERS, H. The beta Catenin/TCF-4 Complex Imposes a Crypt Progenitor Phenotype on Colorectal Cancer Cells. *Cell*. 2002; 111 (2): 241-250.

Genetic and physiological control of branching morphogenesis

Jill Jarecki, Mark Metzstein, Amin Ghabrial, Boaz Levi, Sung Kay Chiu, Eric Johnson, and Mark A. Krasnow

Howard Hughes Medical Institute and Department of Biochemistry, Stanford University, Stanford, CA 94305-5307 USA

Many organs including the mammalian lung, vascular system, and kidney consist of branching networks of tubes. The branching pattern and the size and shape of the branches are critical for their transport functions, but the mechanisms controlling these properties are not well understood (1). Genetic and genomic approaches are being used to elucidate the developmental and physiological programs that govern branching morphogenesis of the *Drosophila* tracheal (respiratory) system, a ramifying network of ~10,000 epithelial tubes that delivers oxygen to the tissues (2). These studies have begun to reveal the cellular and molecular processes that induce sprouting and guide outgrowth of tracheal branches and dictate tube size and shape.

An FGF ligand guides the migrations of the tracheal epithelium as it grows out and assembles into primary branches (3). Cells at the ends of the primary branches are induced to express secondary and terminal genes, and go on to form secondary and terminal branches. Terminal branch outgrowth is controlled by the same FGF ligand, but this later expression of the ligand is not hard-wired but instead regulated by oxygen (4). We have identified the oxygen response pathway that is activated in oxygen-starved cells and induces expression of the FGF ligand to attract new terminal branches. This pathway also stimulates a remarkable cellular response called "hypoxipodia" formation which rearranges existing branches to improve cellular oxygen supply (5).

We have also begun to identify and characterize genes required in tracheal cells to promote outgrowth of new terminal branches in response to the FGF ligand. One of these genes encodes an oxygen-regulated transcription factor which is broadly expressed and translocates to the nucleus in response to low oxygen. Another is a downstream target gene induced by FGF signaling. It encodes a novel Ig domain protein that promotes terminal branch outgrowth and at high levels causes tortuosity of the branches. We will also present a progress report on a saturation genetic screen that has identified over a hundred new terminal branching mutants. These provide an overview of the genetic steps required to extend a terminal branch and create a tubular structure.

References:

- (1) Lubarsky, B. and Krasnow, M.A. (2003) Tube morphogenesis: Making and shaping biological tubes. *Cell* 112, 19-28.

- (2) Ghabrial, A., Luschnig, S., Metzstein, M.M., and Krasnow, M.A. (2003) Branching morphogenesis of the *Drosophila* tracheal system. *Annual Review of Cell and Developmental Biology*, in press.
- (3) Sutherland, D., Samakovlis, C., and Krasnow, M.A. (1996) *branchless* encodes a *Drosophila* FGF homolog that controls tracheal cell migration and the pattern of branching. *Cell* 87, 1091-1101.
- (4) Jarecki, J., Johnson, E. and Krasnow, M.A. (1999) Oxygen regulation of airway branching in *Drosophila* is mediated by Branchless FGF. *Cell* 99, 211-220.
- (5) Wigglesworth, V.B. (1959) The role of the epidermal cells in migration of tracheoles in *Rhodnius prolixus*. *Journal of Experimental Biology* 36, 632-640.

How does the embryo coordinate cell movements and cell fate? Integrating different functions of FGF signalling

Claudio D. Stern, Guojun Sheng and Mario dos Reis

Dept. Anatomy & Developmental Biology, University College London, Gower Street,
London WC1E 6BT, U.K.

c.stern@ucl.ac.uk <http://sternlab.anat.ucl.ac.uk>

During gastrulation the early embryo establishes three initial cell layers (ectoderm, mesoderm and endoderm) by coordinating multiple signalling events with complex cell movements. Shortly afterwards, the ectoderm is subdivided into neural and non-neural (mainly epidermis) sub-regions. A large body of work over the last decade has revealed that these complex patterning processes appear to be controlled by just a handful of signalling factors, and that the molecular pathways are conserved throughout the Animal kingdom. Among the most important signals are the Fibroblast Growth Factors (FGFs), which are required for mesoderm formation, as chemorepellents for the exit of mesoderm cells from the primitive streak, for neural induction and for caudalisation of the neural plate. These processes occur in the ectoderm, very close to each other in time and space – how do the receiving cells decide on the appropriate response?

Previous studies have revealed that FGF signalling is required, but not sufficient for neural induction (Streit et al., 2000; Streit et al., 1998; Wilson and Edlund, 2001). FGF8 induces the expression of the early pre-neural genes *Sox3* and *ERNI* within 2 hours, but unless cells are exposed to other (still unknown) signals, this expression is lost and cells revert to an epidermal fate. We have also found that 5 hours' exposure to either organiser (Hensen's node) signals or to FGF8 are required to sensitise cells to BMP antagonists, which then stabilise the expression of *Sox3*. It therefore became important to define the differences between cells that have or have not been exposed to an organiser for 5 hours.

To answer this, we conducted a differential screen to identify genes whose expression is regulated after 5 hours' exposure to the organiser. We will report the isolation and functions of one of these genes, which we have named *Churchill* (*ChCh*). It encodes a novel C4-type zinc finger that acts as a transcriptional activator, yet it represses the induction of mesodermal markers (*Brachyury* and *Tbx6L*) by FGF, suggesting that at least one of its targets is a transcriptional repressor. We identify one such target, *Smad-interacting-protein-1* (*Sip1*) and show that *ChCh* is required for normal expression of *Sip1* as well as for neural plate development. *ChCh* also sensitises cells to neural inducing signals from the organiser and regulates the cell movements of mesoderm formation. Together with the expression patterns of these components, these results suggest that *ChCh* functions as an important switch between gastrulation (mesoderm/endoderm formation) and neurulation. It also appears to act

as a gate separating two different functions of FGF signalling: in mesoderm formation and in neural induction. Finally, it provides a simple explanation for why cells exposed to FGF for 5 hours become sensitive to BMP signalling.

References:

- Streit, A., Berliner, A., Papanayotou, C., Sirulnik, A., and Stern, C. D. (2000). Initiation of neural induction by FGF signalling before gastrulation. *Nature* *406*, 74-78.
- Streit, A., Lee, K. J., Woo, I., Roberts, C., Jessell, T. M., and Stern, C. D. (1998). Chordin regulates primitive streak development and the stability of induced neural cells, but is not sufficient for neural induction in the chick embryo. *Development* *125*, 507-519.
- Wilson, S. I., and Edlund, T. (2001). Neural induction: toward a unifying mechanism. *Nat Neurosci* *4 Suppl*, 1161-1168.

Session 3: Directing cell migration
Chair: Ruth Lehmann

The eversion of *Drosophila* imaginal discs requires a JNK-regulated change in the adhesive and motile properties of peripodial cells

José Carlos Pastor-Pareja^{1,2}, Antonio García-Bellido¹ and Enrique Martín-Blanco²

1 Centro de Biología Molecular "Severo Ochoa", CSIC, Madrid, Spain

2 Instituto de Biología Molecular de Barcelona, CSIC, Barcelona, Spain

The imaginal discs of *Drosophila* are epithelial sac-like invaginations that will develop the external exoskeleton of the thorax, head and genitalia of the adult fly. During the larval period, the imaginal discs hang from stalks linked to the larval epidermis into the body cavity. Later on, during the first hours of metamorphosis, they evert and fuse. Imaginal discs consist of two kinds of epithelial cells: cubic pseudostratified cells, forming the imaginal disc proper, and squamous or rather squamous peripodial cells, which constitute their stalk and peripodial membrane. Despite the fact that the process of eversion of imaginal discs represents one of the major morphogenetic events during metamorphosis, it has only been roughly described. By serial sectioning and in vivo time-lapse analysis, we have found that at early stages of eversion, the peripodial epithelium of all discs adhere to the external larval layer. At this point, peripodial and larval cells undergo a pseudo epithelial-mesenchymal transition, detaching from each other and, as a consequence, leading to the breaking of epithelium integrity in several points, which soon coalesce into a single hole. This gap widens through continuous loss of adhesion between peripodial cells, resulting in intercalation of new cells at the edge of the initial hole. Finally, the whole peripodial epithelium has rearranged into a stripe of cells that proximally surrounds the already everted disc. This stripe of peripodial cells conforms the leading front of the discs in their movement over intervening larval cells to achieve the discs closure. Once the sealing is complete, these cells differentiate as normal epidermal cells. We have found that the whole eversion process depends of proper levels of JNK activity in peripodial cells, since mutations in both *hemipterous* (JNK-kinase) and *puckered* (JNK-phosphatase) affect across-layers adhesion and cell motility.

Regulation of cell-cell adhesion in wound edge epithelium by protein kinase C alpha

David Garrod and Mohamed Berika

Epidermal keratinocytes are bound tightly together by intercellular junctions, the desmosomes. Wounding promotes migration of cells at the wound edge to re-epithelialise the wound. To do this they partially down-regulate desmosomes so that they can move more easily. We present new data showing that desmosomal adhesion is modulated in epidermal wounds by a mechanism involving protein kinase C alpha signalling.

Recently we reported that the desmosomes in cultured confluent epithelial cell sheets are resistant to disruption by extracellular calcium chelation: they are calcium independent. When such a cell sheet is wounded, desmosomes of wound edge cells become calcium dependent within 1 hour. This effect is propagated to cells deep within the cell sheet. Treatment of the cells with protein kinase C (PKC) activators switches desmosomes rapidly from calcium independence to calcium dependence, and PKC inhibitors do the reverse. Wounding causes translocation of PKC alpha to the cell periphery where it is localised to desmosomes, and antisense depletion of PKC alpha from calcium dependent cells promotes calcium dependence.

We now show that entirely similar changes occur in desmosomes at the wound edge in mouse epidermis. PKC alpha becomes localised to the desmosomes, seemingly penetrating the dense plaques. Desmosomes become calcium dependent and lose their characteristic midline. The effect is propagated to at least 50 cells from the edge. We propose that PKC activation primes desmosomes for internalisation by cells, and that this mechanism down-regulates adhesion. These results have important implications for epithelial cells movement in embryonic development and cancer metastasis.

Early germline development in *Drosophila*

Ruth Lehmann, Rui Martinho, Prabhat Kunwar, Ana Santos, Andy Renault, Thomas Marty,
Hiroko Sano, Michelle Starz-Gaiano and Jennifer Stein

Developmental Genetics Program, Skirball Institute and HHMI, NYU School of Medicine,
540 First Ave, New York, NY 10016. [lehmann@saturn.med.nyu.edu]

In many organisms primordial germ cells (PGCs) form in a specialized germ plasm. In *Drosophila* PGCs form by budding at nuclear cycle 10 while the somatic cells form by polarized cellularization at nuclear cycle 14 in the early embryo (1). We have identified a membrane-associated protein, Slow as molasses as well as other maternal effect mutations whose function is only required for polarized cell growth but not for germ cell formation (2). This suggests that somatic and germ cells already differ by their modes of cell formation. The first molecular manifestation of germ cell specification is the lack of transcriptional activity in PGCs (3, 4). We have identified transcripts expressed in PGCs and have used these to study transcriptional repression during PGC development. Our analysis suggests that relief of transcriptional repression in PGC is mediated by a series of steps. A non-protein coding RNA is needed for the initial repression of transcription by possibly interfering with the activity of RNA polymerase II. Subsequently, members of the Brahma chromatin-remodeling complex also affect early germ line transcriptional quiescence, while the translational repressor protein Nanos plays a role during later stages.

While the PGCs originate at the posterior pole of the embryo, the cells, which will contribute the somatic portion of the gonad, derive from the mesoderm. To reach the gonadal mesoderm PGCs navigate through and along different embryonic tissues. Here, these two cell populations coalesce to form the embryonic gonad, which during the larval and pupal stages differentiates into the ovary and testis. At stage 10 of embryogenesis, germ cells move through the posterior midgut, they then move on the midgut towards the extending germ band. The germ cells then transfer from the gut to the mesoderm. In the mesoderm PGCs associate with three clusters of gonadal mesoderm cells on each side of the embryos. Arrangement of germ cells into two lateral lines and coalescence into the embryonic gonads, is regulated by the reorganization and presumably changing properties of the gonadal mesoderm (5).

In a series of genetic screens we have shown that attractive and repellant signals guide germ cells (6). The repellant signal depends on the activity of the enzyme phosphatidic acid phosphohydrolase, encoded by two redundantly acting genes *wunen* (*wun*) and *wunen-2* (*wun-2*) (7-9). An independently acting, attractive signal is produced by the gonadal mesoderm. We showed that *hmgcr*, the gene encoding the *Drosophila* homolog of 3-hydroxy-3-methylglutaryl Coenzyme A reductase (HMG-CoAR), is required for PGCs to leave the midgut and to associate with the gonadal mesoderm precursors (10). Within the germ cells,

G-protein coupled receptor signaling mediates directional response to migratory cues. Final coalescence of the gonad requires the activity of the transmembrane protein FOI and E-cadherin (11).

References:

- (1) Starz-Gaiano M. and Lehmann R (2001) Moving toward the next generation. *Mechanisms of Development* 105(1-2):5-18.
- (2) Stein JA, Broihier HT, Moore LA, Lehmann R. (2002) Slow as Molasses is required for polarized membrane growth and germ cell migration in *Drosophila*. *Development*.129(16):3925-3934.
- (3) Van Doren M., Williamson, A.L. and Lehmann R. (1998): Regulation of zygotic gene expression in *Drosophila* primordial germ cells. *Current Biol.* 8, 243-246.
- (4) Seydoux G, Dunn MA. Transcriptionally repressed germ cells lack a subpopulation of phosphorylated RNA polymerase II in early embryos of *Caenorhabditis elegans* and *Drosophila melanogaster*. *Development*. 1997 124(11):2191-201.
- (5) Moore L.A., Broihier T.H., Van Doren M. and Lehmann R. (1998b) Gonadal mesoderm and fat body initially follow a common developmental path in *Drosophila*. *Development* 125, 837-844.
- (6) Moore L.A., Broihier T.H., Van Doren M., Lunsford, L. and Lehmann R. (1998a): Identification of genes controlling germ cell migration and embryonic gonad formation in *Drosophila*. *Development* 125, 667-678.
- (7) Zhang N, Zhang J, Purcell KJ, Cheng Y, Howard K. The *Drosophila* protein Wunen repels migrating germ cells. *Nature*. 1997 Jan 2;385(6611):64-7.
- (8) Starz-Gaiano M., Cho N.K., Forbes A. and Lehmann R. (2001) Spatially restricted activity of Lipid Phosphatase guides migrating germ cells. *Development* 128, 983-991
- (9) Renault A.D., Starz-Gaiano M. and Lehmann R. Metabolism of sphingosine-1-phosphate and lysophosphatidic acid: a genome wide analysis of gene expression in *Drosophila* MOD/Gene Expression Patterns 2002, 2:337-45.
- (10) Van Doren M., Broihier T.H., Moore L.A., and Lehmann R. (1998): HMG-CoA reductase guides migrating primordial germ cells. *Nature* 396(6710), 466-469.
- (11) Van Doren M., Mathews W.R., Samuels M., Broihier T.H., Moore L.A., and Lehmann R. *fear of intimacy* encodes a novel transmembrane protein required for gonad morphogenesis in *Drosophila*. *Development* in press.

Cell migration and morphogenesis of the *Drosophila* tracheal system

Marta Llimargas and Jordi Casanova

Institut de Biologia Molecular de Barcelona (CSIC)

Cell migration is a widespread phenomenon in many biological processes, both in development and in the adult organism. In particular, cells move in an ordered pattern, following defined paths of migration, probably by recognition of distinct cues and substrates. Thus, the establishment of specific interactions between cells and their substrates is a crucial step in migration, a process ultimately determined by molecules expressed at their surface.

The tracheal system of *Drosophila* is an especially appropriate model for the study of the genetic control of morphogenesis and, in particular, for the study of the mechanisms that guide cells to migrate in specific directions. The larval tracheal system of *Drosophila* is a complex tubular network that conducts oxygen from the exterior to the internal tissues. It arises from the tracheal placodes, clusters of ectodermal cells that appear at each side of ten embryonic segments. The cells of each cluster invaginate and migrate in different and stereotyped directions to form each of the primary tracheal branches (for review, see Hogan and Yingling 1998; Metzger and Krasnow 1999). The general conclusion from many studies is that the direction of migration of the tracheal cells relies on a set of positional cues provided by nearby cells. On the one hand, *branchless* (*bnl*), a gene encoding an FGF homologue, is expressed around the developing tracheal system in clusters of cells at each position in which a new branch will form and grow. Activation of the *Breathless* (*Btl*) receptor in the tracheal cells by *Bnl* is thought to stimulate and guide tracheal migration toward these positions (Sutherland et al. 1996). On the other hand, *Dpp*, *EGF*, and *Wnt* signalling have a role in the choice between the alternative directions of migration (Llimargas and Casanova 1997; Vincent et al. 1997; Wappner et al. 1997; Chihara and Hayashi 2000; Llimargas 2000). However, it is not known how the signals from *Dpp*, *EGF*, and *Wnt* specify a particular migratory path or what cell surface proteins are used by the tracheal cells to interact with their specific substrates.

To understand the mechanisms that are responsible for the distinct migratory pathways of the cells of the different branches we have first analysed how the tracheal cells migrate in the context of the whole organism. We have found that the alternative migratory pathways of the tracheal cells are associated with distinct subsets of mesodermal cells (Franch-Marro and Casanova 2000). We have then addressed which are the cell surface proteins involved in the recognition of these migratory pathway. In particular, we have found that migration of the cells of the visceral branches, the ramifications of the tracheal tree that transport oxygen to the gut, is mediated by the restricted expression of the *PS1* integrin in their cells, which is matched by the complementary expression of the *PS2* in the cells of their

migratory substrate. Moreover, we have found that the signalling pathways that mediate the choice between alternative paths of migration regulate the appropriate expression of the α PS1 subunit in the subset of the tracheal cells of the visceral branches (Boube et al., 2001). These results support a model in which signalling by transduction pathways specifies the particular migratory pathways of tracheal cells by regulating a precise array of adhesion proteins such as integrins at their surface. We think that this can be a very general mechanism to regulate integrin expression in distinct subpopulations of cells within a wider field and to confer them different substrate recognition properties. At present, we are focusing our study on the identification of new genes, specially coding for additional cell surface proteins involved in tracheal cell migration and morphogenesis of the tracheal system.

References:

- Boube, M., Martin-Bermudo, M.D., Brown N.H. and Casanova, J. (2001). *Genes and Dev.* 15, 1554-1562.
- Chihara, T. and Hayashi, S. 2000. *Development* 127: 4433-4442.
- Franch-Marro, X. and Casanova, J. 2000. *Dev. Biol.* 270: 80-90.
- Hogan, B.L.M. and Yingling, J.M. 1998. *Curr. Opin. Genet. Dev.* 8: 481-486.
- Llimargas, M. 2000. *Development* 127: 4407-4417.
- Llimargas, M. and Casanova, J. 1997. *Development* 124:3273-3281.
- Manning, G. and Krasnow, M.A. 1993. Development of the *Drosophila* tracheal system. In *The development of Drosophila melanogaster*. (ed. M. Bate and A. Martínez-Arias), Vol. I, pp. 609-685. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.
- Metzger, R.J. and Krasnow, M.A. 1999. *Science* 284: 1635-1639.
- Sutherland, D., Samakovlis, C., and Krasnow, M.A. 1996. *Cell* 87: 1091-1101.
- Vincent, S., Ruberte, E., Grieder, N.C., Cha, C., Haerry, T., Schuh, R., and Affolter, M. 1997. *Development* 124: 2741-2750.
- Wappner, P., Gabay, L., and Shilo, B-Z. 1997. *Development* 124: 4707-4716.

Genetic control of cell migration in *Drosophila*

Tina Bridges, Erika Geisbrecht, Jocelyn McDonald, Debby Silver, Xuejiao Wang,
Denise J. Montell

Johns Hopkins School of Medicine

Border cell migration in the *Drosophila* ovary has emerged as a useful genetic model for studying the conversion of stationary epithelial cells to invasive migratory cells in vivo. Three signaling pathways have been defined, which control different aspects of the migration (reviewed in Montell, 2003). A steroid hormone signal regulates when the cells become migratory and may regulate adhesion dynamics. Signaling through the JAK/STAT pathway defines which epithelial cells become migratory. Finally a growth factor signal contributes to guiding the cells to their destination. Each of these types of signaling pathways is known to be upregulated in a variety of human cancers, suggesting that the same signaling pathways that are known for promoting cell proliferation and survival may also contribute to metastasis by regulating cell motility and invasiveness. To test this hypothesis we have studied the effects of inhibiting JAK/STAT signaling on the motility of human ovarian cancer cell lines.

The small GTPase Rac is also required for border cells to migrate, and we have conducted a genetic screen in vivo for genes that, when over-expressed, suppress the migration defects due to dominant-negative Rac. Over-expression of wild-type Rac, actin, profilin, or a Rac exchange factor suppresses the migration defect. In addition we identified a new mediator of Rac-dependent actin polymerization in this screen.

References:

Montell, D. J. (2003). Border-cell migration: the race is on, *Nat Rev Mol Cell Biol* 4, 13-24.

**Session 4: Cell movements and the developing
nervous system**
Chair: M. Angela Nieto

Molecular analysis of neural crest formation

Marianne Bronner-Fraser

Division of Biology, California Institute of Technology, Pasadena, CA 91125 USA
(mbronner@caltech.edu)

Neural crest cells arise within the ectoderm during neurulation and give rise to most of the peripheral nervous system. Following neural tube closure, they come to lie within the dorsal neural tube from which they emerge and subsequently migrate extensively to numerous and characteristic sites. There, they differentiate into neurons and glia of the peripheral nervous system, cartilage and bone of the face, melanocytes and various other cell types. Fate mapping experiments have demonstrated that the neural crest arises at the juncture between presumptive epidermis and neural plate. However, injection of lineage tracer into individual cells reveals that single neural fold cells are not committed to a neural crest fate; rather these cells can form all ectodermal derivatives (epidermis, neural tube, neural crest).

Inductive interactions between the neural and non-neural ectoderm can generate neural crest cells, suggesting that signals travel through the epidermis to generate neural crest cells prior to neural tube closure. Induction of the neural crest appears to be a multiphasic process and involves a combination of an early Wnt signal, likely mediated by Wnt6, together with later functions for BMP signaling pathways. We show that Wnt is both necessary and sufficient for this stage of neural crest induction. Recent experiments have focused on the initiation of neural crest induction in the gastrulating embryo. Our results suggest that cells in the gastrula are already conditionally specified to form neural crest cells. Finally, we have been taking a genomics approach to identify the array of molecules expressed as a result of neural crest induction and will discuss the results of these screens.

Local tissue interactions across the dorsal midline underlie morphogenesis of asymmetric forebrain nuclei

Miguel L. Concha, Claire Russell, Jennifer C. Regan, Marcel Tawk, Jon Clarke and Stephen W. Wilson

Animals show behavioural, cognitive and neuroanatomical asymmetries but the mechanisms that establish these asymmetries are not well understood. The dorsal diencephalon (epithalamus) is one of the most evolutionarily conserved sites of asymmetry in the vertebrate brain (Concha and Wilson, 2001). In this region, the habenular nuclei are usually asymmetric and the photoreceptive parapineal, when present, is frequently positioned or projects axons asymmetrically. Our previous studies in collaboration with Alex Schier's group showed that the Nodal signalling pathway is necessary for specifying laterality in the epithalamus, but is not essential for the development of asymmetry per se (Concha et al. 2000). More recently we have been analysing the morphogenetic events and tissue interactions that regulate the development of asymmetric nuclei in the epithalamic region of the dorsal forebrain. We show that the unilateral parapineal organ has a bilateral origin and that some parapineal precursors migrate across the midline to form this left-sided nucleus. The parapineal subsequently innervates the left habenular nucleus, which itself derives from ventral epithalamic cells directly adjacent to the parapineal precursors. We show that ablation of cells in the left ventral epithalamus of wild type embryos can lead to reversal of epithalamic laterality and that such ablations can impose the direction of CNS asymmetry in embryos in which laterality is normally randomised. These data lead us to propose that laterality is determined by a competitive interaction between the left and the right epithalamus. We are currently testing the notion that the role of Nodal signalling is to bias the outcome of this competition.

Our collaborators on this and related projects include Becky Burdine, Samuel Sidi, Darren Gilmour, Enrique Amaya, David Kimelman, Lauro Sumoy, Teresa Nicolson, Stefan Gründer, Miranda Gomperts and Alex Schier.

References:

- Concha, M. and Wilson, S.W. (2001). Asymmetry in the epithalmus of vertebrates. **J.Anatomy** 199, 63-84.
 Concha, M., Burdine, R., Russell, C., Schier, A.S. and Wilson, S.W. (2000). A Nodal signalling pathway regulates the laterality of CNS asymmetries in the zebrafish forebrain. **Neuron** 28 399-409.

Axon migration at the midline of the *Drosophila* central nervous system

Marios Georgiou¹, Anna Myat¹, Pauline Henry², Daniela Rotin² and Guy Tear¹

¹Molecular Neurobiology Group, MRC Centre for Developmental Neurobiology, New Hunts House, Guy's Hospital Campus, King's College, London SE1 1UL UK and ²Program in Cell Biology, The Hospital for Sick Children, Toronto, Canada M5G 1X8

Neurons within the central nervous system (CNS) extend axons that migrate towards, near to or away from the midline. The specialised cells that lie at the midline produce a range of signals with a key role in regulating the guidance decisions of these neurons. Those axons that do migrate towards the midline also switch their behaviour so that they do not remain at the midline but extend across to the contralateral side. These axons rarely recross but ignore previously attractive midline cues and extend within a longitudinal tract or exit the CNS. The changes in axonal behaviour are produced in part by the modification of cell surface receptor protein activity. The full extent of the mechanisms used and how they switch the response of axons is not yet fully understood. It is clear that there is tight spatial and temporal regulation of the surface expression of guidance receptors as well as the activation of novel molecular interactions that modify axonal responses to midline signals as the axons cross the midline cells. Commissureless (Comm) is one of the few molecules known to have an important role to regulate the levels of a receptor protein necessary for axon guidance at the midline. Comm is able to down-regulate levels of the Roundabout (Robo) protein that acts as a receptor for the midline derived axon outgrowth inhibitor Slit. Down-regulation of Robo is necessary for axons to cross the midline.

Comm is a novel transmembrane molecule that is expressed on midline cells and is seen to accumulate on commissural axons. Comm protein is required in the embryo for normal axon guidance. We have discovered that Comm is required and expressed both in commissural neurons and in midline cells. Comm localisation within these cells is dynamic where it is found within intracellular vesicles and at the cell surface. Comm can bind Robo and is able to sequester Robo within the intracellular vesicles within commissural neurons prior to crossing the midline. This allows the commissural axons to extend toward the midline cells that express the outgrowth inhibitory Robo ligand Slit. The intracellular portion of Comm is essential for its function since a truncated form of the protein lacking this region is inactive. Using a yeast two-hybrid screen we identified a *Drosophila* protein homologous to the known vertebrate protein Nedd4 which specifically binds the intracellular portion of Comm. Nedd4 is a modular protein containing three or four WW domains, a C2 Ca²⁺ /lipid binding domain and an E3 ubiquitin ligase domain. We have identified the amino acid sequence within Comm that binds to two of the Nedd4 WW domains. We have identified that Nedd4 catalyses the ubiquitination of Comm, is necessary for Comm localisation to the

vesicles and for Comm to sequester Robo away from the cell surface. We have further identified that an interaction between Nedd4 and Comm is also necessary for overexpressed Comm to downregulate Robo in the embryo.

The Comm protein expressed by the commissural axons accumulates at the midline and is thus prevented from acting on Robo once the axons have crossed the midline. Specific domains within the extracellular and transmembrane regions of Comm appear to be necessary for this targeting of the Comm protein. Comm can also bind itself through its extracellular domains and a possible trans-interaction between Comm at the midline and on the commissural axons may confine high Comm activity to the commissures.

Molecular mechanisms of cell migration in the telencephalon

Oscar Marín

Neurons are most frequently born at a distance from the place where they finally become integrated in a specific neuronal circuit, so that they have to migrate to reach their final destination. The process of cell migration requires young neurons to perfectly synchronize multiple actions, including the timing for initiation and cessation of the movement as well as the appropriate responses to multiple guidance cues encountered through their trajectory.

The telencephalon is undoubtedly one of the most intricate regions of the mammalian brain, and its extraordinary degree of organization reflects the complexity of the migratory movements required to generate it. Two general modes of migration are distinguished in the telencephalon: radial migration, which established the general cytoarchitectonical framework of the different telencephalic subdivisions, and tangential migration, which increases the cellular complexity of its circuits by allowing the dispersion of multiple neuronal types. This later type of migration appears to be governed by mechanisms similar to those that control the guidance of growing axons, i.e. contact guidance (permissive and non-permissive substrates for migration) and diffusible gradients (attractive and repulsive cues).

The movement of interneurons from the basal telencephalon to the embryonic cortex is one of the most prominent examples of tangential migration in the telencephalon. Despite being a highly directional process, the molecular mechanisms controlling this migration are still poorly understood. Here, I will review the cellular and molecular mechanisms underlying the migration of interneurons to the embryonic cortex and discuss how emerging concepts in neuronal migration are reshaping our understanding of brain development in normal and pathological situations.

Referencias:

- Corbin, J. G., Nery, S. and Fishell, G. (2001). Telencephalic cells take a tangent: non-radial migration in the mammalian forebrain. *Nature Neuroscience* 4, 1177-82.
- Marín, O. and Rubenstein, J. L. R. (2001). A long, remarkable journey: tangential migration in the telencephalon. *Nature Reviews Neuroscience* 2, 780-790.
- Marín, O. and Rubenstein, J. L. R. (2003). Cell migration in the forebrain. *Annual Review of Neuroscience* in press.
- Tessier-Lavigne, M. and Goodman, C. S. (1996). The molecular biology of axon guidance. *Science* 274, 1123-33.

Hoxa2 overexpression in rhombomere 1 reveals rhombic lip autonomous patterning of tangential migration

Mark Eddison, Leah Toole, Esther Bell, Andrew Lumsden and Richard J. T. Wingate

The rhombic lip of rhombomere 1 gives rise to a unique population of tangentially migrating precursor cells, which condense as the external granule cell layer of the cerebellum. As the most rostral hindbrain segment, rhombomere 1 is defined by an absence of Hox gene expression. To assess the role of segmental identity in specifying rhombic lip development, we have used both infection by RCASBP(B)Hoxa2 virus and electroporation of an RCASBP(B)Hoxa2 construct to drive ectopic Hoxa2 expression in chick. Overexpression of Hoxa2 within r1 results in the downregulation of granule cell markers (Pax6, ErbB4), while the expression of Phox2a, which normally characterises the trochlear nucleus and locus coeruleus in ventral rhombomere 1, is upregulated throughout the cerebellum. To follow the fate of migratory cells, Hoxa2 misexpression was targeted to rhombic lip precursors by combining electroporation with the microsurgical construction of quail-chick chimaeras. This reveals that the loss of granule cell markers in cerebellum expressing ectopic Hoxa2 can be attributed to a respecification of the migration path of cerebellar rhombic lip derivatives to a single ventrolateral, extra-cerebellar target. A complementary grafting strategy confirms that specification of migration path is autonomous to the rhombic lip.

Establishment of cell polarity in the chick

Cristina Afonso, Domingos Henrique

We are particularly interested in understanding the mechanisms of polarized membrane growth during neural tube morphogenesis. While much emphasis has been given to the contribution of the actin cytoskeleton per se, the role of protein and membrane trafficking in remodeling the cell surface and establishing polarized structures within the neuroepithelial cell has received little attention.

We have isolated several chick homologues of the *C. elegans* par-3 and par-6 genes, which encode PDZ proteins, and studied their expression during neural development. Using antibodies against one of the PAR-3 proteins and one of the PAR-6 proteins, we have recently found that dividing neuroepithelial cells in the chick embryo assemble a basally-located molecular complex during mitosis, including the PAR3 and PAR6 proteins, the NUMB protein and some proteins known to be involved in vesicular trafficking.

Given the known role of these molecules, our current hypothesis is that the PAR3/PAR6 proteins nucleate the building of a macromolecular complex, which includes probably other proteins like the GTPases CDC42 and Rac. This complex would orchestrate various steps during creation of neuroepithelial polarity, like the reorganization of the cytoskeleton and directed trafficking of proteins and membranes that leads to the polarized growth of neuroepithelial cells, after exit of mitosis.

To test this hypothesis, we are following the localization of PAR3 and PAR6-GFP fusion proteins in vivo, using time-lapse recording in chick neural tube slices. We also use other markers to study the organization and dynamics of organelles, as well as the intracellular transport of integral membrane proteins in these live slices of chick neural tube. We also try to interfere with the normal function of the PAR3/PAR6/... basal complex, by overexpressing different forms of its constituent proteins in single neuroepithelial cells, using electroporation of embryonic neural tube. We are following subsequent events inside the cell, dynamically, by timelapse microscopy of cultured neural tube slices, and evaluate the effects on neuronal division, migration and differentiation.

**Session 5: Cell movements, development
and disease**

Chair: Marianne Bronner-Fraser

The migratory phenotype of Snail-expressing cells

Sonia Vega, Oscar Ocaña, Agnès Boutet, Marta García del Barrio and M. Angela Nieto

Instituto Cajal, CSIC. Doctor Arce, 37, 28002 Madrid. E-mail: anieto@cajal.csic.es

The Snail family of zinc-finger transcription factors is involved in processes that imply profound cell movements both during embryonic development and tumour progression. We and others have previously shown that they are crucial for the formation of the mesoderm and the neural crest and for the acquisition of migratory and invasive properties in epithelial tumours through the triggering of the epithelial-mesenchymal transition (EMT) (reviewed in 1).

The function of Snail in the triggering of EMT is in part mediated by the direct repression of E-cadherin transcription (2,3). However, the EMT implies a dramatic phenotypic change concomitant not only with the loss of epithelial markers but also with the gain of mesenchymal markers and changes in cell shape. Since Snail is able to induce a full EMT in epithelial cells, it must have additional targets. Indeed, there are other direct targets already identified such as the epithelial Mucin-1 (4) or components of the tight junctions such as claudin and occludin (5). Through the analysis of Snail transfected cells and overexpression experiments in embryos we found that in addition to cell adhesion molecules, Snail is upstream of molecules involved in cytoskeletal changes (6). We are currently further characterizing the phenotype of the Snail-expressing cells and have found that this transcription factor is also involved in cell cycle control and cell survival. I will discuss these results in the frame of the properties that both embryonic and tumour cells share when they become migratory and invasive.

References:

- 1.- Nieto, M. A. (2002). *Nature Rev. Mol. Cell Biol.* 3, 155-166.
- 2.- Cano, A., Pérez, M. A., Rodrigo, I., Locascio, A., Blanco, M. J., Del Barrio, M. G., Portillo, F. and Nieto, M. A. (2000). The transcription factor Snail controls epithelial-mesenchymal transitions by repressing E-cadherin expression. *Nature Cell Biol.* 2, 76-83
- 3.- Batlle, E., Sancho, E., Franci, C., Dominguez, D., Monfar, M., Baulidam J. and Garcia De Herreros, A. *Nature Cell Biol.*, 2, 84-89 (2000).
- 4.- Guaita, S., Puig, I., Franci, C., Garrido, M., Dominguez, D., Batlle, E., Sancho, E., Dedhar, S., De Herreros, A.G. and Baulida, J. (2002). Snail induction of epithelial to mesenchymal transition in tumor cells is accompanied by MUC1 repression and ZEB1 expression. *J Biol Chem.* 277, 39209-16.
- 5.- Ikenouchi, J., Matsuda, M., Furuse, M. and Tsukita, M. (2003). Regulation of tight junctions during epithelium-mesenchyme transition: direct repression of the gene expression of claudins/ occludin by Snail. *J. Cell Science* 116, 1959-1967.
- 6.- Del Barrio, M.G. and Nieto M.A. (2002). *Development* 129, 1583-1594.

Parallels between morphogenesis and wound healing in embryos

Paul Martin

Department of Anatomy and Development Biology
University College London
Gower Street
GB – London WC1E 6BT
Phone: +44 207 679 6577
Fax: +44 207 679 7349
E-mail: paul.martin@ucl.ac.uk

Embryos heal wounds very rapidly and efficiently and without leaving a scar. Studying how they do this can tell us much about the natural morphogenetic movements of embryogenesis as well as suggesting ways in which we might make adult tissues repair more efficiently. Using live confocal imaging of transgenic *Drosophila* embryos expressing gfp-actin in epithelial tissues we have revealed the key actin machineries that drive the paradigm morphogenetic process of dorsal closure which appears to bear striking analogy with re-epithelialisation of a vertebrate skin wound. Using embryos expressing mutant forms of the various small GTPases, we have tested the function of each of these actin-based elements - the actin cable and dynamic filopodia and lamellipodia - in both dorsal closure and the repair of laser-generated wound holes in the fly embryo. Our experiments in embryonic chicks and mice and in the neonatal PU.1 null mouse, which is genetically macrophageless, suggest that an inflammatory response is not essential for healing and may indeed be causal of fibrosis in post-embryonic animals. Consequently, we have used a microarray approach with this mouse in order to identify a portfolio of candidate inflammation/fibrosis genes. Finally, by taking advantage of the translucency of the zebrafish larval tail we have begun to make DIC movies of the inflammatory response and to dissect the genetics of this process by screening for mutants that fail to recruit leukocytes to the wound site and by morpholino knockdown of candidate “inflammation” genes. Our hope is that these basic cell and molecular studies in genetically tractable organisms will supply us with the clues we need to design the new repair and regeneration medicines of the future.

References:

- Jacinto, A., Wood, W., Turmaine, M., Martinez-Arias, A. & Martin, P. (2000). Dynamic actin based adhesion and cell matching during *Drosophila* dorsal closure. *Curr. Biol.* **10**, 1420-1426.
- Jacinto, A., Martinez-Arias, A. & Martin, P. (2001) Mechanisms of epithelial fusion and repair. *Nature Cell Biol.* **3**, E117-123.

- Harsum, S., Clarke, J. and Martin, P. (2001). A reciprocal relationship between cutaneous nerves and repairing skin wounds in the developing chick embryo. *Dev. Biol.* **238**, 27-39.
- Grose, R., Harris, B., Cooper, L., Topilko, P. and Martin, P. (2002). The immediate early genes *krox-24* and *krox-20* are rapidly upregulated following wounding in the embryonic and adult mouse. *Dev Dynamics* **223**, 371-378.
- Jacinto, A., Wood, W., Woolner, S., Hiley, C., Turner, L., Wilson, C., Martinez-Arias, A. & Martin, P. (2002) Dynamic analysis of actin cable function during *Drosophila* dorsal closure. *Curr Biol.* **12**, 1245-1250.
- Jacinto, A., Woolner, S. and Martin, P. (2002). Dynamic analysis of dorsal closure in *Drosophila* – from genetics to cell biology. *Dev. Cell* **3**, 9-19.
- Wood, W., Jacinto, A., Gale, J., Grose, R., Woolner, S., Wilson, C. and Martin P. (2002). Wound healing recapitulates morphogenesis in *Drosophila* embryos. *Nature Cell Biol.* **4**, 907-912.



Genes that control cell migration during mouse embryo

Carmen Birchmeier

During development, cell migration is an important process and it is frequently observed that cells are born at one position and subsequently move to their final locations. In development, migration events are tightly controlled. Cells are released at defined stages and positions, and they move along defined routes to their particular target sites. Molecules that control such migration events have received much attention. Of particular interest is the molecular nature of signals responsible for the release of the cells, for the maintenance of cellular motility, or for directed migration and target finding. I will summarize here data from my laboratory that contributed to an understanding of the molecular control of cell migration in the embryo. Over the years, we analysed genes that control migration of two developing cell types, neural crest cells and muscle progenitor cells. Both cell types are released from epithelial structures, and in order to detach and become motile they first undergo an epithelial-mesenchymal conversion. Mechanistically, the release of the cells and their subsequent motility resembles the process that is observed late during the progression of malignant carcinomas. There, cells detach from the primary tumor by epithelial-mesenchymal conversion and migrate in an uncontrolled manner to form metastases at sites distant of the primary tumor. Because of the mechanistic similarities, it is not so astonishing that genes implicated in tumor progression play also important roles in cell migration events during development. In particular, work from my laboratory demonstrated that tyrosine kinase receptors implicated in tumorigenesis, like the c-ErbB or c-Met tyrosine kinase receptors, turn out to control decisive steps in migration of embryonic cells.

P O S T E R S

Hyaluronan Synthase 2 is required for dorsal migration of lateral mesodermal cells during zebrafish gastrulation

Jeroen Bakkers, Carina Kramer, Joris Pothof, Nicolette Quaedvlieg, Herman Spaink and Matthias Hammerschmidt

The large polysaccharide Hyaluronan (HA) and its synthesizing enzymes (Has) have been implicated in regulating the migratory potential of metastatic cancer cells. Here, we analyze the roles of zebrafish Has2 in normal development. Antisense morpholino oligonucleotide (MO)-mediated knockdown of zebrafish Has2 leads to severe migratory defects during gastrulation, somite morphogenesis, and germ cell development. During gastrulation, ventrolateral cells of *has2* morphant embryos fail to develop lamellipodia and to migrate dorsally, resulting in a blockage of dorsal convergence, while extension of the dorsal axis is normal. The effect is cell autonomous, suggesting that HA acts as an autocrine or local signal. Upon ectopic expression in axial cells, *has2* causes the formation of supernumerary lamellipodia and a blockage of mediolateral cell-cell intercalations and axis extension. Epistasis analyses indicate that these effects of Has2 are mediated by activating the Rho GTPase Rac1. Together, the data suggest that Has2 and Rac1 are necessary and sufficient components of a linear pathway inducing the migration of lateral cells, while blocking extension movements. This provides first genetic evidence that convergence and extension are separate morphogenetic movements of gastrulation.

Analysis of the genetic program of epithelial cell plasticity

Geert Berx, Joke Comijn, Bram De Craene, Kristin Strumane, Frans Van Roy

Epithelial-mesenchymal transitions (EMTs) are a manifestation of epithelial cell plasticity during morphogenesis, wound healing, and tumour progression. Loss of the cell-cell adhesion molecule E-cadherin seems to be heavily involved in EMT appointing E-cadherin as one of the key caretakers of the epithelial phenotype. In cancer the amount of E-cadherin is decreased through mutations, chromosomal loss or active repression of the E-cadherin promoter. Different transcription factors like SIP1 (smad interacting protein 1), Snail and Slug repress E-cadherin transcription in vitro and in vivo by binding to E-box sequences of the E-cadherin promoter. The transcription factor SIP1 belongs to the dEF-1/Zfh-1 family of two-handed zinc finger/homeodomain proteins, which can efficiently repress cell-cell adhesion and induce invasion. Conditional expression in an epithelial cell modelsystems of SIP1 modulates epithelial plasticity by repressing E-cadherin and inducing EMT. Transfer of E-cadherin in an E-cadherin negative model cellsystem showing high SIP1 expression resulted in restoration of cell-cell adhesion and epithelial differentiation. We performed a comparative transcriptome screen from these model cellsystems using cDNA arrays of 17.268 cDNAs to delineate signaling pathways and transcriptional events that determine epithelial cell plasticity controlled by E-cadherin, and SIP1. Analysis of the identified pathways and genes will advance the understanding of the molecular mechanisms that underlie tumor invasiveness and metastasis.

Requirement of Sfrp1 in eye anlage specification in medaka embryos

Pilar Esteve, Javier Lopez-Rios, Josana Rodriguez- Sanchez and Paola Bovolenta

It is currently believed that the levels of Wnt activity may specify posterior to anterior fates in the neural plate and that the most rostral fates will develop only in a Wnt-signalling free zone. In line with this idea Hourt et al. (2002, *Neuron* 35 ; 255-265) have demonstrated that in zebrafish, telecephalic specification require the activity of a Wnt inhibitor of the Secreted Frizzled Related family (SFRP), known as tlc. We had previously isolated in chick (Esteve et al, 2001; Esteve et al, 2003) and medaka fish a different member of this family, Sfrp1. In both species, Sfrp1 is strongly expressed in the anterior neural plate including the eye anlage. To test whether this molecule could also contribute to the specification of anterior neural territories, we have attempted to interfere with the expression of this gene by injecting specific morpholinos into medaka fish (*Oryzias latipes*) embryos at the two cell stage. Morphological and molecular analysis, using anterior or retinal specific markers (Rx, Six3, Emx1, Pax6, BF1), indicates that morpholino treated-embryos have a severely reduced eye field. On the other hand, over-expression of Sfrp1 in medaka embryos leads to an expansion of the anterior neural fates, particularly of the eye anlage, as judged by molecular analysis. This “anteriorization” of the embryos is associated either to A-P axis truncation or axis duplication. Though over-expression experiment might be complicated by a general interference with Wnts activities, affecting both the canonical and the PCP pathway and we believe that altogether our result point to a specific role of Sfrp1 in eye anlage specification.

Association between the cell cycle and neural crest delamination through specific regulation of G1/S transition

Tal Burstyn-Cohen and Chaya Kalcheim

The neural crest (NC) is a group of transient progenitors which rapidly disperses in the embryo and differentiates into a rich variety of derivatives (Le Douarin and Kalcheim, 1999). Successful migration is essential for cells to reach their homing sites where differentiation occurs. To engage in migration, epithelial pre-migratory cells must convert into mesenchyme. Epithelio-mesenchymal transitions are, therefore, an essential prerequisite for the development of many embryonic tissues and organs. When converting into mesenchyme, they become loosely associated or fully individualize, acquire motile properties and invade initially the extracellular matrix surrounding the dorsolateral neural tube (Le Douarin and Kalcheim, 1999, Kalcheim, 2000, Nieto, 2001). The identity of signals triggering NC cell delamination remained elusive until recently. Sela-Donenfeld and Kalcheim (1999) have reported that a gradient of BMP4 activity created along the dorsal neural tube triggers emigration of NC progenitors.

Here we show that NC cells emigrate in the S-phase of the cell cycle. Moreover, we establish that this feature is part of the mechanism of cell delamination as treatment of explanted neural primordia with the G1-S transition inhibitors olomoucine, AG555 or mimosine prevented initial delamination of NC cells, that could be rescued upon removal of the inhibitors. In contrast, similar treatments with aphidicolin or VM-26, which inhibit the cycle at S and G2 phases, respectively, had no effect. To further examine the significance of G1-S transition on cell delamination in the living embryo, we electroporated hemi-neural tubes with several constructs leading to a cell cycle arrest at the G1 phase, resulting in an almost total prevention of NC delamination in the treated side.

Taken together, we demonstrate for the first time that the transition between G1 to S is necessary for the epithelial-to-mesenchymal conversion of premigratory NC. Molecular events occurring during the progression of G1-S transition may therefore be translated into downstream signals that generate cell movement and/or be required for the cells to respond to the environmental factors that trigger delamination.

References:

- Kalcheim C.(2000). Mechanisms of early neural crest development: From cell specification to migration. *Int Rev Cytol* 200, 143-196.
- Le Douarin NM, and Kalcheim C. (1999). *The Neural Crest*. 2nd ed. New York: Cambridge University Press.
- Nieto AM.(2001). The early steps in neural crest development. *Mech Dev* 105, 27-35.
- Sela-Donenfeld D, and Kalcheim C.(1999). Regulation of the onset of neural crest migration by coordinated activity of BMP4 and Noggin in the dorsal neural tube. *Development* 126, 4749-4762.

New insights into the initiation of gastrulation in the mouse embryo

Aitana Perea-Gomez, Anne Moreau, Anne Camus, Christian Cibert, Jérôme Collignon

Institut Jacques Monod, Paris

In the mouse embryo, gastrulation starts at 6.5 days of development with the appearance of the primitive streak at the posterior pole of the embryo. Fate mapping studies and gene expression analysis have hinted at the existence of a complex cellular choreography between the time of implantation and the initiation of gastrulation. However, current data offer only a fragmented view of these processes and we still lack a full understanding of their dynamics.

Using histology, expression studies, optical coherence tomography and cell-lineage, we documented the cellular and molecular events that precede the initiation of gastrulation at 6.5 dpc. Our marker studies show that the antero-posterior (AP) axis of the embryo at 6.0 dpc, shortly before gastrulation, is perpendicular to the direction which would have been inferred from earlier studies. However, through a mechanism which can involve asymmetrical regulation of gene expression and associated changes in the behaviour of a subset of epiblast cells, its orientation, given by the formation of the primitive streak (PS) at 6.5 dpc, can change rapidly and ends up being particular to each embryo. Our results therefore suggest additional steps in the establishment of the AP axis in the mouse embryo. We are currently investigating whether early positional clues in the preimplantation embryo have a predictive value regarding the position at which the primitive streak forms at later stages.

Dynamics of cell rearrangement in early zebrafish gastrulation

Miguel L. Concha, Benjamin Feldman, Daisy Faruque, Derek L. Stemple, Stephen W. Wilson, and Richard J. Adams

Gastrulation defines the process by which the three germ layers, ectoderm, mesoderm and endoderm are formed. We have been studying the cellular mechanisms that underlie this crucial event by using *in vivo* time-lapse imaging and cell movement analyses. In zebrafish, gastrulation involves extensive cell internalisation along the blastoderm margin to form the prospective mesendoderm, a process that is intimately co-ordinated with the vegetally directed epibolic movements of marginal cells, enveloping layer (EVL), and yolk syncytial layer (YSL). As gastrulation begins, marginal cells accumulate into a sheet that moves coherently vegetal-ward in close association with epiboly of the EVL and YSL. Strikingly, this movement of the blastoderm persists and a velocity gradient develops along the animal-vegetal axis of this tissue -- the fastest moving cells being those closest to the margin -- even when epiboly of the YSL itself pauses.

This strongly suggests that the generating force for internalisation resides at the margin. During this time the YSL changes shape and marginal blastomeres within two rows of the margin start to internalise. Although internalisation occurs concurrently in neighbouring marginal cells the patterns of cell rearrangement, motility and shape changes indicate that cells are moving autonomously giving the impression of active ingression. Hence, the process of cell internalisation in zebrafish represents a model in which individual cell ingressions are constrained to the narrow edge of a cell sheet. Examination of mutant and morphant embryos reveals a key role of Nodal signalling in regulating the dynamics of internalisation. Enhancement of Nodal signalling by depletion of *lefty1/2* proteins induces an extended period of active internalisation that leads to an excessive accumulation of hypoblast cells, seen as an exaggerated thickening of the germ ring and shield. This phenotype contrasts with the complete failure in cell internalisation and absence of germ ring and shield in embryos lacking Nodal signalling.

Notch lateral activation promotes epithelial-mesenchymal transitions during heart development and neoplastic transformation

Luika A. Timmerman, Joaquin Grego, Esther Bertrán, José María Pérez-Pomares, Juan Diez, Sergi Aranda, Sergio Palomo, Angel Raya, Frank McCormick, Juan Carlos Izpisua-Belmonte
& José Luis de la Pompa

The Notch pathway regulates cell-fate choices in embryonic and adult tissues via mechanisms termed lateral inhibition and lateral activation, the latter being poorly understood in vertebrates. Disruption of Notch has severe developmental consequences and ectopic expression of its intracellular domain (Notch IC) is oncogenic in mammals. Here we show that Notch ligands and receptors are co-expressed at high levels in cells of the embryonic endocardium, including cells which overlie the heart valve primordia (endocardial cushions). Ablation of the Notch1 receptor or the effector RBPJK curtails expression of Notch ligands, receptors and the target gene *HRT1*, demonstrating the existence of a positive feed-back loop consistent with lateral activation. Notch-expressing endocardial cells undergo a TGF β -mediated endothelial-mesenchymal transition (EMT), to cellularize developing cardiac valves. Loss of Notch activity attenuates expression of TGF β 2 and its receptors, prevents local expression of the snail transcriptional repressor, and stabilizes expression of the endothelial cellular adhesion molecule VE-cadherin, resulting in concomitant loss of endocardial EMT. Conversely, transient ectopic expression of Notch1IC in zebrafish embryos results in abnormally enlarged and hypercellular cardiac cushions. Overexpression of Notch1IC in endothelial cells *in vitro* also induces severe attenuation of VE-cadherin expression and loss of contact inhibition. These transformed cells undergo an apparent EMT, as evidenced by independent cell migration on plastic, through collagen, and in xenografts. We conclude that Notch plays an unexpected, novel role in the promotion of EMT in both developmental and tumor formation settings, in part via regulation of the cellular adhesion system.

Guidance of border cell migration by the *Drosophila* EGFR and PDGF/VEGF receptor orthologs

Peter Duchek and Pernille Rorth

Border cells are a group of about 8 specialized follicle cells that perform a stereotypic migration during *Drosophila* oogenesis. This developmentally controlled cell migration serves as an *in vivo* model system that can be genetically modified, allowing both gain-of-function as well as loss-of-function analyses. It was not known what guides border cell migration, but the graded expression of guidance cues might serve this purpose. We reasoned that uniform overexpression of such guidance factors should destroy the gradient and lead to inefficient migration. In a genetic screen we overexpressed random endogenous genes in the tissue through which the border cells migrate. We found that uniform activation of the EGFR or the newly identified PDGF/VEGF receptor ortholog PVR inhibits border cell migration, as is expected for guidance receptors. The EGFR ligand Gurken as well as the PDGF/VEGF-like protein PVF1 are expressed in the oocyte, which is the target for border cell migration. Although border cells mutant for either EGFR or PVR are still able to migrate properly in the majority of cases, co-expression of dominant negative forms of both EGFR and PVR completely inhibits border cell migration, indicating a requirement *in vivo* and a redundancy of both RTKs for the guidance function. Surprisingly, we found that this guidance function is independent of the PI3K, PLCgamma, and ERK pathways.

References:

- Duchek, P., and Rorth, P. (2001). Guidance of cell migration by EGF receptor signaling during *Drosophila* oogenesis. *Science* 291: 131
- Duchek, P., et al. (2001). Guidance of cell migration by the *Drosophila* PDGF/VEGF receptor. *Cell* 107: 17

Regulation of patterning and morphology in the zebrafish embryonic forebrain

Luca Caneparo, Isabelle Foucher, Karima Kissa and Corinne Houart

In the past decade, tremendous progress has been made in understanding the molecular and cellular events leading to the establishment of the body plan in vertebrate. The most amazing finding was the great conservation in the genetic pathways used for any given process. The making of a limb, an eye or a brain is depending upon very similar cascades of molecular events from fish to human. However, the morphology of each of those structures differs drastically amongst vertebrates. We chose the zebrafish (*danio rerio*) as a model organism to study the early events defining the, size and shape of the forebrain compartments (telencephalon, eye field and diencephalons). Our study let us to identify signalling events, inside the neural plate, responsible for the early regionalisation of the forebrain territory (Houart et al, 1998; Houart et al. 2002). In the course of our study, we were surprised to observe that modifications in patterning events have often a very weak impact on morphogenesis. Reciprocally, changes in morphology can be found without altering the patterning of the brain. We will present a set of data showing striking cases where morphology and patterning seem to be controlled independently and will discuss the possibility of such an uncoupling as a general mechanism for evolution of shapes.

References:

- Houart C, Caneparo L, Heisenberg C, Barth K, Take-Uchi M, Wilson S. (2002) Establishment of the telencephalon during gastrulation by local antagonism of Wnt signaling. *Neuron*. 35, 255-65.
- Houart C, Westerfield M, Wilson SW. (1998) A small population of anterior cells patterns the forebrain during zebrafish gastrulation. ; *Nature*.; 391, 788-92.

Modeling neurulation: computer simulations of genetic networks, cell interactions, mechanics and motion

Michel Kerszberg

I introduce ctrl-Dev, an experimental computer system adapted to the description and simulation of genetical, molecular and biomechanical processes in cells and groups of cells, i.e. tissues. The system is one of the first to integrate within a single framework cell motility and genetic interactions. I shall present a study using ctrl-Dev as applied to the early events in neurogenesis (neurulation). The genetic networks responsible for neurulation in Insects and Vertebrates (Xenopus, and chick to a lesser extent) exhibit remarkable homologies. Yet the morphogenesis is very different: the Insects have a (ventral) neural cord consisting of discrete ganglia arising from delaminating neuroblasts; Vertebrates have a (dorsal) neural tube. Here I use ctrl-Dev to test the workability of an hypothesis which might account for these diverse morphologies (though not for the dorso/ventral inversion itself). The genetic networks included in the simulations are (using the Vertebrate "language"): the neural induction system (messages such as chordin and Sonic hedgehog secreted by the notochord, and BMP protein); "memory" networks for differentiation of neural plate (e.g. Nkx2, Pax3) and of neurons [AS-C, E(spl)]; and a cytoskeletal network responsible for apical/basal differentiation and cell motions. I demonstrate how a simple switch in upstream control of this last network can explain alternatively the formation of a neural tube or of ganglia, thus proposing a testable mechanism for their evolutionary divergence. A film of tube formation will be shown. The program is now being applied to Insect segmentation and to a simulation of cell movements during gastrulation and primitive streak formation.

Defining the molecular determinants of epimorphin/syntaxin-2-mediated mammary epithelial morphogenesis

D. C. Radisky, S. Bennett, Y. Hirai, and M. J. Bissell

The fully differentiated mammary gland is highly optimized for secretion of milk into the lumen of a highly branched ductal epithelial structure, and tubulogenesis is an essential component of the development and ramification of the mammary ductal system. We have identified a particular role in this process for epimorphin/syntaxin-2 (EPM), a mesenchymal protein that directs tubular morphogenesis in mammary gland development. EPM is a transmembrane protein localized to the cell surface that has been shown to exist in both intracellular and extracellular topological orientations. When presented to the extracellular surface of mammary epithelial cells, EPM acts as a morphogen and produces distinct morphogenic outcomes depending upon the mode of presentation. When presented in a polar, basal fashion, EPM produces branching morphogenesis, whereas presented in an apolar fashion, EPM leads to luminal cyst morphogenesis. We have used a three-dimensional (3D) collagen gels to define EPM function, and have found that the activity of EPM is contained within an N-terminal three-helix bundle. We have used the well-studied and highly-homologous syntaxin-1A (which shows no activity as a mammary morphogen) as a basis for homology modeling of EPM, to predict the active site of EPM action. We performed site-specific mutagenesis of syntaxin-1A to alter the six residues that are different in EPM at this predicted active site, and we found that this hybrid product was functional; in doing so, we created an active morphogen from an inactive template. This information can be used to more completely define the role of EPM in mammary morphogenesis.

Prickle1 regulates cell movements during gastrulation and neuronal migration in zebrafish

Filipa Carreira-Barbosa, Miguel L. Concha, Masaki Takeuchi, Naoto Ueno, Stephen W. Wilson and Masazumi Tada

During vertebrate gastrulation, mesodermal and ectodermal cells undergo convergent extension, a process characterised by prominent cellular rearrangements in which polarised cells intercalate along the medio-lateral axis leading to elongation of the antero-posterior axis. Recently, it has become evident that a non-canonical Wnt/Frizzled (Fz)/Dishevelled (Dsh) signalling pathway, related to the planar cell polarity (PCP) pathway in flies, regulates convergent extension during vertebrate gastrulation. Here we isolate and functionally characterise a zebrafish homologue of *Drosophila prickle* (*pk*), a gene implicated in the regulation of PCP. Zebrafish *pk1* is expressed maternally and in moving mesodermal precursors. Abrogation of Pk1 function by morpholino oligonucleotides leads to defective convergent extension movements, enhances the *silberblick* (*slb*)/*wnt11* and *pipetail/wnt5* phenotypes and suppresses the ability of Wnt11 to rescue the *slb* phenotype. Gain-of-function of Pk1 also inhibits convergent extension movements and enhances the *slb* phenotype, most likely due to the ability of Pk1 to block the Fz7-dependent membrane localisation of Dsh by down-regulating levels of Dsh protein. Furthermore, we show that *pk1* genetically interacts with *trilobite* (*tri*)/*strabismus* to mediate the caudally directed migration of cranial motor neurons as well as convergent extension. These results suggest that during zebrafish gastrulation, Pk1 acts in part through interaction with the non-canonical Wnt11/Wnt5 pathway to regulate convergent extension cell movements, but is unlikely to simply be a linear component of this pathway. In addition, Pk1 interacts with Tri to mediate posterior migration of branchiomotor neurons, probably independent of the non-canonical Wnt pathway.

Transcriptional regulation of Cerberus during embryonic development

Ana Teresa Tavares, Ana Cristina Borges, Jose Antonio Belo

In the post genome-sequencing era, one of the main challenges will be to identify the sequence information that specifies when and where a given gene is expressed, that is, its cis-regulatory program. These programs are essential in the control of developmental processes, and changes in their sequence and organisation are one of the major causes of animal evolution (Davidson, 2001). Cerberus-like genes code for cystine-knot secreted proteins and exhibit both common and specific expression patterns during *Xenopus*, chick and mouse embryonic development (Bouwmeester et al., 1996; Rodriguez-Esteban et al., 1999; Belo et al., 1997). In order to study the regulation of chick Cerberus, we have cloned and analysed portions of its cis-regulatory regions. In brief, 5'-genomic sequences were subcloned in EGFP expression vectors and introduced into early chick embryos by electroporation (in *New culture*). Deletion and site-directed mutagenesis analyses led to the identification of distinct candidate regulatory regions that direct expression in the anterior mesendoderm and in the left side mesoderm. Since the stability of the EGFP RNA and protein is higher than that of Cerberus RNA we were able to trace the fates of the Cerberus-expressing cells. Interestingly, our observations indicate that the anterior mesendodermal cells that express Cerberus constitute a population of precursors precursor cells common to the foregut endoderm, heart and anterior blood islands. Additionally, the ongoing cross-species analysis will help to understand the evolutionary divergence of Cerberus-like gene regulation. Preliminary observations of chick Cerberus-EGFP transgenic mice suggest that the upstream regulators of chick Cerberus expression are also present in the mouse anterior mesendoderm and left side mesoderm.

References:

- Belo, J.A. et al. (1997). *Mech Dev.* 68:45-57.
- Bouwmeester, T. et al. (1996) *Nature.* 382:595-601.
- Davidson, E.H. (2001) *Genomic regulatory systems: Development and evolution.* Academic Press, SanDiego, USA.
- Rodriguez-Esteban, C. et al. (1999) *Nature.* 401(6750):243-51.

TGF β -dependent regulation of trophoblast invasion during placental implantation

Patrick Fafet, Carolina Segura-Morales, Jean Marie Blanchard, Thierry Maudelonde and Marie-Luce Vignais

The placenta is the first organ to form during mammalian embryogenesis. This organ is vital as it provides the interface between fetal and maternal tissues and enables nutrient exchange between the mother and the fetus. Within the placenta, the trophoblast lineage constitutes the most important cell type, with the syncytiotrophoblast and the extravillous cytotrophoblast playing important structural and functional roles for the exchanges between the maternal blood system and the fetus vasculature. Implantation of the placenta requires migration of the trophoblast cells within the maternal endometrium in an invasion process which is controlled both in time and space.

We are studying the molecular mechanisms of trophoblast invasion in humans and their regulation by the TGF β signaling pathway. We have designed a culture system based on the coculture of human primary endometrial fibroblasts and of trophoblastic villi obtained from elective abortions. Using placenta from early pregnancies (3 to 5 weeks), we can reconstitute the invasion process which occurs in utero with both active syncytiotrophoblast and extravillous cytotrophoblast. These cocultures are analyzed by phase contrast microscopy in time lapse experiments, under controlled CO₂ and at 37°C, for periods of 24 to 48 hours. We observe that trophoblast cells invade the endometrial fibroblast layer and form lacuna.

This invasion process is reversed by TGF β . We are currently investigating the role of metalloproteases in this invasion and its regulation using immunofluorescence and RT-PCR on cells isolated by laser microdissection. These studies are carried out in parallel in primary human dermal and endometrial fibroblasts.

Functional interaction of p120catenin with the Rho family of small GTPases during early *Xenopus* morphogenesis

Malgorzata Ciesiolka, Mieke Delvaeye, Frans Van Roy and Kris Vleminckx

We are studying early morphogenetic processes in the early *Xenopus* embryo.

We are especially interested in the functional regulation of cadherin mediated cell-cell adhesion during cellular migration and tissue rearrangements. A good candidate regulator of cadherin functionality is p120ctn, a protein that is situated at the interplay between the cadherin adhesion complex and the cytoskeleton. We investigated the expression of Xp120ctn during early embryogenesis. In order to understand the function of Xp120ctn in early development we either overexpressed Xp120ctn or mutants of E-cadherin that can not bind Xp120ctn. In both cases the head structures were affected as reflected by malformations of the eyes and the craniofacial skeleton. Interestingly, as has also been documented in vitro, we observed that overexpressed Xp120ctn modulates the activity of endogenous RhoA. Moreover, Xp120ctn overexpression was phenocopied by dominant-negative RhoA and the morphological defects obtained by Xp120ctn overexpression could be fully rescued by coinjection of wild type or constitutive active RhoA. In addition, coinjection of XE-cadherin with Xp120ctn could rescue the embryos and relieved the RhoA inhibition, possibly by sequestering the excess of cytosolic p120ctn. These results indicate that Xp120ctn is critically involved in regulating morphogenesis in the early *Xenopus* embryo both through action on the cytoskeleton and by regulating the activity of the cadherins.

LIST OF INVITED SPEAKERS

- Carmen Birchmeier** Max-Delbrück-Centrum für Molekulare Medizin. Robert-Roessle-Strasse 10, 13125 Berlin (Germany). Tel.: 49 30 94 06 2403. Fax: 49 30 94 06 2656. E-mail: cbirch@mdc-berlin.de
- Marianne Bronner-Fraser** Division of Biology, California Institute of Technology, Pasadena, CA. 91125 (USA). Tel.: 1 626 3953 355. Fax: 1 626 395 7717. E-mail: mbronner@caltech.edu
- Jordi Casanova** Institut de Biologia Molecular de Barcelona (CSIC). Jordi Girona, 18-26, 08034 Barcelona (Spain). Tel.: 34 93 204 06 00. Fax: 34 93 204 5904. E-mail: jcrbmc@cid.csic.es
- Suzanne Eaton** Max Planck Institute of Molecular Cell Biology and Genetics. Pfotenhauerstrasse 108, 01307 Dresden (Germany). Tel.: 49 351 210 2526. Fax: 49 351 210 2000. E-mail: eaton@mpi-cbg.de
- Scott E. Fraser** Beckman Institute, California Institute of Technology, Pasadena, CA. 91125 (USA). Tel.: 1 626 395 2790. Fax: 1 626 449 5163. E-mail: sefraser@caltech.edu
- Ray Keller** Department of Biology, University of Virginia. Gilmer Hall, Charlottesville, VA. 22904-4328 (USA). Tel.: 1 434 982 5769. Fax: 1 434 982 5626. E-mail: rek3k@Virginia.edu
- Mark A. Krasnow** Howard Hughes Medical Institute and Department of Biochemistry, Stanford University, Stanford, CA. 94305-5307 (USA). Tel.: 1 650 723 7191. Fax: 1 650 723 6783. E-mail: krasnow@pmgm2.stanford.edu
- Ruth Lehmann** Developmental Genetics Program, Skirball Institute and HHMI, NYU School of Medicine. 540 First Ave, New York, NY. 10016 (USA). Tel.: 1 212 263 8071. Fax: 1 212 263 7760. E-mail: lehmann@saturn.med.nyu.edu
- Oscar Marín** Instituto de Neurociencias. Unidad de Neurobiología del Desarrollo. Universidad Miguel Hernández. Campus de San Juan, 03550 San Juan, Alicante (Spain). Tel.: 34 965 91 94 87. Fax: 34 965 91 94 15. E-mail: o.marin@umh.es
- Paul Martin** Department of Anatomy and Developmental Biology. University College London. Gower Street, London WC1E 6BT (UK). Tel.: 44 207 679 6577. Fax: 44 207 679 7349. E-mail: paul.martin@ucl.ac.uk
-

-
- | | |
|---------------------------------|---|
| David McClay | Department of Biology, Duke University, Durham, NC. 27708-1000 (USA). Tel.: 1 919 613 8188. Fax: 1 919 613 8177. E-mail: dmcclay@duke.edu |
| Denise J. Montell | Johns Hopkins School of Medicine. 725 N. Wolfe St., 414 WBSB, Baltimore, MD. 21205 (USA). Tel.: 1 410 614 2016. Fax: 1 410 955 5759. E-mail: dmontell@jhmi.edu |
| M. Angela Nieto | Instituto Cajal, CSIC. Doctor Arce, 37, 28002 Madrid (Spain). Tel.: 34 91 585 47 23. Fax: 34 91 585 47 54. E-mail: anieto@cajal.csic.es |
| Lilianna Solnica-Krezel | Department of Biological Sciences, Vanderbilt University. VU Station B 351634, Nashville, TN. 37235-1634 (USA). Tel.: 1 615 343 9413. Fax: 1 615 343 6707. E-mail: lilianna.solnica-krezel@vanderbilt.edu |
| Claudio D. Stern | Dept. of Anatomy & Developmental Biology, University College London. Gower Street, London WC1E 6BT (UK). Tel.: 44 20 7679 3346. Fax: 44 20 7679 2091. E-mail: c.stern@ucl.ac.uk |
| Guy Tear | Molecular Neurobiology Group, MRC Centre for Developmental Neurobiology, New Hunts House, Guy's Hospital Campus, King's College, London SE1 1UL (UK). Tel.: 44 207 848 6539. Fax: 44 207 848 6816. E-mail: guy.tear@kcl.ac.uk |
| Stephen W. Wilson | Dept. of Anatomy and Developmental Biology. University College London. Gower Street, London WC1E 6BT (UK). Tel.: 44 20 7679 3348. Fax: 44 20 7679 7349. E-mail: s.wilson@ucl.ac.uk |
| Lewis Wolpert | Anatomy & Developmental Biology University College London. Gower Street, London WC1E 6BT (UK). Tel.: 44 207 679 13 20. Fax: 44 207 813 28 13. E-mail: l.wolpert@ucl.ac.uk |
| Magdalena Zernicka-Goetz | Wellcome Trust/Cancer Research Institute. Tennis Court Road, Cambridge CB2 1QR (UK). Tel.: 44 1223 76 3291. Fax: 44 1223 33 4089. E-mail: mzg@mole.bio.cam.ac.uk |
-

LIST OF PARTICIPANTS

- Jane Alfred** Development. The Company of Biologists Ltd. Bidder Building. 140 Cowley Road, Cambridge CB4 0DL (UK). Tel.: 44 1223 420 007. Fax: 44 1223 423 353. E-mail: jane@biologists.com
- Enrique Amaya** Wellcome Trust/Cancer Research UK Institute. University of Cambridge. Tennis Court Road, Cambridge CB2 1QR (UK). Tel.: 44 1223 334 195. Fax: 44 1223 334 089. E-mail: ea3@mole.bio.cam.ac.uk
- Pilar Aroca** Departamento de Anatomía Humana. Facultad de Medicina. Universidad de Murcia, 30120 Espinardo, Murcia (Spain). Tel.: 34 968 364 682. Fax: 34 968 363 955. E-mail: pilarroca@um.es
- Jeroen Bakkers** Max-Planck Institut für Immunbiologie. Stuebeweg 51, 79108 Freiburg (Germany). Tel.: 49 761 5108 493. Fax: 49 761 5108 333. E-mail: bakkers@immunbio.mpg.de
- Eduard Batlle** Hubrecht Laboratory. Netherlands Inst. for Developmental Biol. Uppsalalaan 8, 3584 CT Utrecht (The Netherlands). Tel.: 31 30 212 1849. Fax: 31 30 251 6464. E-mail: ebatlle@niob.knaw.nl
- Geert Berx** Dept. of Molecular Biomedical Research. VIB and Ghent Univ. Ledeganckstraat 35, 9000 Gent (Belgium). Tel.: 32 9 2645318. Fax: 32 9 2645348. E-mail: Geert.Berx@dmb.rug.ac.be
- María José Blanco** Departamento de Ciencias Morfológicas I. Universidad Complutense de Madrid. Avda. Complutense s/n, 28040 Madrid (Spain). Tel.: 34 91 585 4736. Fax: 34 91 585 4754. E-mail: mjblanco@cajal.csic.es
- Paola Bovolenta** Instituto Cajal, CSIC. Doctor Arce, 37, 28002 Madrid (Spain). Tel.: 34 91 585 4717. Fax: 34 91 585 4754. E-mail: bovolenta@cajal.csic.es
- Tal Burstyn-Cohen** Department of Anatomy and Cell Biology. The Hebrew University. Hadassah Medical School, Jerusalem 91120 (Israel). Tel.: 972 2 6757039. Fax: 972 2 6757451. E-mail: talic@cc.huji.ac.il
- Katrin Bussell** Nature Reviews Molecular Cell Biology. 4 Crinan Street, London N1 9XW (UK). Tel.: 44 2078 433 630. Fax: 44 2078 433 696. E-mail: K.Bussell@nature.com
-

-
- | | |
|------------------------------|---|
| Jérôme Collignon | Dept. de Biologie du Développement. Institut Jacques-Monod. CNRS, Universités Paris 6 et 7. 2 place Jussieu, 75251 Paris cedex 05 (France). Tel.: 33 144 276 108. Fax: 33 144 275 265. E-mail: collignon@ijm.jussieu.fr |
| Miguel L. Concha | Instituto de Ciencias Biomédicas, Facultad de Medicina. Universidad de Chile, Santiago 7 (Chile). Tel.: 56 2 6786364. Fax: 56 2 6786264. E-mail: mconcha@machi.med.uchile.cl |
| José Luis de la Pompa | Molecular Oncology Dept. Institut de Recerca Oncologica. Hospital Duran i Reynals. Gran Via s/n, km. 2,7, 08907 L'Hospitalet de Llobregat, Barcelona (Spain). Tel.: 34 93 2607828. Fax: 34 93 2607426. E-mail: jldelapompa@iro.es |
| Flora de Pablo | Centro de Investigaciones Biológicas, CSIC. Velázquez, 144, 28006 Madrid (Spain). Tel.: 34 91 564 8978. Fax: 34 91 564 7518. E-mail: fdelpablo@cib.csic.es |
| Peter Duchek | Ludwig Institute for Cancer Research. Courtauld Building. 91 Riding House Street, London W1W 7BS (UK). Tel.: 44 2078 784 047. Fax: 44 2078 784 040. E-mail: duchek@ludwig.ucl.ac.uk |
| David Garrod | School of Biological Sciences. University of Manchester. 2339 Stopford Building, Oxford Road, Manchester M13 9PT (UK). Tel.: 44 1612 755 243. Fax: 44 1612 753 915. E-mail: david.garrod@man.ac.uk |
| Domingos Henrique | Instituto Medicina Molecular. Faculdade de Medicina de Lisboa - IHE. Av Prof Egas Moniz, 1649-028 Lisboa (Portugal). Tel.: and. Fax: 351 21 794 0058. E-mail: henrique@fm.ul.pt |
| Corinne Houart | MRC Centre for Developmental Neurobiology. King's College London. Guy's Campus, London SE1 1UL (UK). Tel.: 44 2078 486 409. Fax: 44 2078 486 550. E-mail: corinne.houart@kcl.ac.uk |
| Michel Kerszberg | Récepteurs et Cognition. Institut Pasteur. 25 rue du Dr. Roux, 75724 Paris Cedex 15 (France). Tel.: 33 1 45688808. Fax: 33 1 45688836. E-mail: mkersz@pasteur.fr |
| Miguel Manzanares | Instituto de Investigaciones Biomédicas CSIC-UAM. Arturo Duperier 4, 28029 Madrid (Spain). Tel.: 34 91 585 4493. Fax: 34 91 585 4401. E-mail: mmanzanares@iib.uam.es |
| Enrique Martín-Blanco | Instituto de Biología Molecular de Barcelona, CSIC. Jordi Girona Salgado 18-26, 08034 Barcelona (Spain). Tel.: 34 93 400 6100. Fax: 34 93 204 5904. E-mail: embbmc@cid.csic.es |
-

Roberto Mayor	Howard Hughes Medical Institute, International Scholar Millennium Nucleus in Dev. Biology. Fac. de Ciencias. Univ. de Chile, Casilla 653, Santiago (Chile). Tel.: 562 678 7351. Fax: 562 276 3802. E-mail: rmayor@uchile.cl
Aixa V. Morales	Department of Developmental Neurobiology. Instituto Cajal (CSIC). Doctor Arce, 37, 28002 Madrid (Spain). Tel.: 34 91 585 4736. Fax: 34 91 585 4754. E-mail: aixamorales@cajal.csic.es
Beverly Purnell	Science. 1200 New York Ave., NW, Washington, DC. 20005 (USA). Tel.: 1 410 734 0307. Fax: 1 410 734 0311. E-mail: bpurnell@aaas.org
Derek C. Radisky	Lawrence Berkeley National Laboratory. 1 Cyclotron Road, MS 83-101, Berkeley, CA. 94720 (USA). Tel.: 1 510 486 4368. Fax: 1 510 486 5586. E-mail: dcradisky@lbl.gov
Deborah J. Sweet	Cell Press. 1100 Massachusetts Avenue, Cambridge, MA. 02138 (USA). Tel.: 1 617 661 7057. Fax: 1 617 397 2810. E-mail: dsweet@cell.com
Masazumi Tada	Department of Anatomy and Developmental Biology. University College London. Gower Street, London WC1E 6BT (UK). Tel.: 44 2076 793 362. Fax: 44 2076 797 349. E-mail: m.tada@ucl.ac.uk
Ana Teresa Tavares	Instituto Gulbenkian de Ciencia. Rua da Quinta Grande, 6, Ap.14, 2781-901 Oeiras (Portugal). Tel.: 351 21 4407918. Fax: 351 21 4407970. E-mail: atavares@igc.gulbenkian.pt
Miguel Torres	Centro Nacional de Biotecnología. CSIC. Campus UAM, Cantoblanco, 28049 Madrid (Spain). Tel.: 34 91 585 4849. Fax: 34 91 372 0493. E-mail: mtorres@cnb.uam.es
Marie-Luce Vignais	IGM/CNRS UMR 5535. 1919 Route de Mende, 34293 Montpellier Cedex 5 (France). Tel.: 33 467 61 36 50. Fax: 33 467 04 02 31. E-mail: vignais@igm.cnrs-mop.fr
Kris Vleminckx	Department of Molecular Biomedical Research. Unit of Developmental Biology. Ghent University-VIB. K.L. Ledeganckstraat 35, 9000 Gent (Belgium). Tel.: 32 9 264 8719. Fax: 32 9 264 5348. E-mail: krisv@dmbrug.ac.be
Richard J. T. Wingate	MRC Centre for Developmental Neurobiology. King's College London. Guy's Campus, London SE1 1UL (UK). Tel.: 44 2078 486 542. Fax: 44 2078 486 550. E-mail: richard.wingate@kcl.ac.uk

*Texts published in the
SERIE UNIVERSITARIA*

by the

FUNDACIÓN JUAN MARCH

*concerning workshops and courses organized within the
Plan for International Meetings on Biology (1989-1991)*

*: Out of stock.

***246 Workshop on Tolerance: Mechanisms and Implications.**

Organizers: P. Marrack and C. Martínez-A.

***247 Workshop on Pathogenesis-related Proteins in Plants.**

Organizers: V. Conejero and L. C. Van Loon.

***248 Course on DNA - Protein Interaction.**
M. Beato.

***249 Workshop on Molecular Diagnosis of Cancer.**

Organizers: M. Perucho and P. García Barreno.

***251 Lecture Course on Approaches to Plant Development.**

Organizers: P. Puigdomènech and T. Nelson.

***252 Curso Experimental de Electroforesis Bidimensional de Alta Resolución.**

Organizer: Juan F. Santarén.

253 Workshop on Genome Expression and Pathogenesis of Plant RNA Viruses.

Organizers: F. García-Arenal and P. Palukaitis.

254 Advanced Course on Biochemistry and Genetics of Yeast.

Organizers: C. Gancedo, J. M. Gancedo, M. A. Delgado and I. L. Calderón.

***255 Workshop on the Reference Points in Evolution.**

Organizers: P. Alberch and G. A. Dover.

***256 Workshop on Chromatin Structure and Gene Expression.**

Organizers: F. Azorín, M. Beato and A. A. Travers.

257 Lecture Course on Polyamines as Modulators of Plant Development.

Organizers: A. W. Galston and A. F. Tiburcio.

***258 Workshop on Flower Development.**

Organizers: H. Saedler, J. P. Beltrán and J. Paz-Ares.

***259 Workshop on Transcription and Replication of Negative Strand RNA Viruses.**

Organizers: D. Kolakofsky and J. Ortín.

***260 Lecture Course on Molecular Biology of the Rhizobium-Legume Symbiosis.**

Organizer: T. Ruiz-Argüeso.

261 Workshop on Regulation of Translation in Animal Virus-Infected Cells.

Organizers: N. Sonenberg and L. Carrasco.

***263 Lecture Course on the Polymerase Chain Reaction.**

Organizers: M. Perucho and E. Martínez-Salas.

***264 Workshop on Yeast Transport and Energetics.**

Organizers: A. Rodríguez-Navarro and R. Lagunas.

265 Workshop on Adhesion Receptors in the Immune System.

Organizers: T. A. Springer and F. Sánchez-Madrid.

***266 Workshop on Innovations in Proteases and Their Inhibitors: Fundamental and Applied Aspects.**

Organizer: F. X. Avilés.

- 267 **Workshop on Role of Glycosyl-Phosphatidylinositol in Cell Signalling.**
Organizers: J. M. Mato and J. Larner.
- 268 **Workshop on Salt Tolerance in Microorganisms and Plants: Physiological and Molecular Aspects.**
Organizers: R. Serrano and J. A. Pintor-Toro.
- 269 **Workshop on Neural Control of Movement in Vertebrates.**
Organizers: R. Baker and J. M. Delgado-García.

Texts published by the

CENTRE FOR INTERNATIONAL MEETINGS ON BIOLOGY

- 1 **Workshop on What do Nociceptors Tell the Brain?**
Organizers: C. Belmonte and F. Cerveró.
- *2 **Workshop on DNA Structure and Protein Recognition.**
Organizers: A. Klug and J. A. Subirana.
- *3 **Lecture Course on Palaeobiology: Preparing for the Twenty-First Century.**
Organizers: F. Álvarez and S. Conway Morris.
- *4 **Workshop on the Past and the Future of Zea Mays.**
Organizers: B. Burr, L. Herrera-Estrella and P. Puigdomènech.
- *5 **Workshop on Structure of the Major Histocompatibility Complex.**
Organizers: A. Arnaiz-Villena and P. Parham.
- *6 **Workshop on Behavioural Mechanisms in Evolutionary Perspective.**
Organizers: P. Bateson and M. Gomendio.
- *7 **Workshop on Transcription Initiation in Prokaryotes**
Organizers: M. Salas and L. B. Rothman-Denes.
- *8 **Workshop on the Diversity of the Immunoglobulin Superfamily.**
Organizers: A. N. Barclay and J. Vives.
- 9 **Workshop on Control of Gene Expression in Yeast.**
Organizers: C. Gancedo and J. M. Gancedo.
- *10 **Workshop on Engineering Plants Against Pests and Pathogens.**
Organizers: G. Bruening, F. García-Olmedo and F. Ponz.
- 11 **Lecture Course on Conservation and Use of Genetic Resources.**
Organizers: N. Jouve and M. Pérez de la Vega.
- 12 **Workshop on Reverse Genetics of Negative Stranded RNA Viruses.**
Organizers: G. W. Wertz and J. A. Melero.
- *13 **Workshop on Approaches to Plant Hormone Action**
Organizers: J. Carbonell and R. L. Jones.
- *14 **Workshop on Frontiers of Alzheimer Disease.**
Organizers: B. Frangione and J. Ávila.
- *15 **Workshop on Signal Transduction by Growth Factor Receptors with Tyrosine Kinase Activity.**
Organizers: J. M. Mato and A. Ullrich.
- 16 **Workshop on Intra- and Extra-Cellular Signalling in Hematopoiesis.**
Organizers: E. Donnall Thomas and A. Grafena.
- *17 **Workshop on Cell Recognition During Neuronal Development.**
Organizers: C. S. Goodman and F. Jiménez.

- 18 **Workshop on Molecular Mechanisms of Macrophage Activation.**
Organizers: C. Nathan and A. Celada.
- *19 **Workshop on Viral Evasion of Host Defense Mechanisms.**
Organizers: M. B. Mathews and M. Esteban.
- *20 **Workshop on Genomic Fingerprinting.**
Organizers: M. McClelland and X. Estivill.
- 21 **Workshop on DNA-Drug Interactions.**
Organizers: K. R. Fox and J. Portugal.
- *22 **Workshop on Molecular Bases of Ion Channel Function.**
Organizers: R. W. Aldrich and J. López-Barneo.
- *23 **Workshop on Molecular Biology and Ecology of Gene Transfer and Propagation Promoted by Plasmids.**
Organizers: C. M. Thomas, E. M. H. Willington, M. Espinosa and R. Díaz Orejas.
- *24 **Workshop on Deterioration, Stability and Regeneration of the Brain During Normal Aging.**
Organizers: P. D. Coleman, F. Mora and M. Nieto-Sampedro.
- 25 **Workshop on Genetic Recombination and Defective Interfering Particles in RNA Viruses.**
Organizers: J. J. Bujarski, S. Schlesinger and J. Romero.
- 26 **Workshop on Cellular Interactions in the Early Development of the Nervous System of *Drosophila*.**
Organizers: J. Modolell and P. Simpson.
- *27 **Workshop on Ras, Differentiation and Development.**
Organizers: J. Downward, E. Santos and D. Martín-Zanca.
- *28 **Workshop on Human and Experimental Skin Carcinogenesis.**
Organizers: A. J. P. Klein-Szanto and M. Quintanilla.
- *29 **Workshop on the Biochemistry and Regulation of Programmed Cell Death.**
Organizers: J. A. Cidlowski, R. H. Horvitz, A. López-Rivas and C. Martínez-A.
- *30 **Workshop on Resistance to Viral Infection.**
Organizers: L. Enjuanes and M. M. C. Lai.
- 31 **Workshop on Roles of Growth and Cell Survival Factors in Vertebrate Development.**
Organizers: M. C. Raff and F. de Pablo.
- 32 **Workshop on Chromatin Structure and Gene Expression.**
Organizers: F. Azorín, M. Beato and A. P. Wolffe.
- *33 **Workshop on Molecular Mechanisms of Synaptic Function.**
Organizers: J. Lerma and P. H. Seeburg.
- *34 **Workshop on Computational Approaches in the Analysis and Engineering of Proteins.**
Organizers: F. S. Avilés, M. Billeter and E. Querol.
- 35 **Workshop on Signal Transduction Pathways Essential for Yeast Morphogenesis and Cell Integrity.**
Organizers: M. Snyder and C. Nombela.
- 36 **Workshop on Flower Development.**
Organizers: E. Coen, Zs. Schwarz-Sommer and J. P. Beltrán.
- *37 **Workshop on Cellular and Molecular Mechanism in Behaviour.**
Organizers: M. Heisenberg and A. Ferrús.
- 38 **Workshop on Immunodeficiencies of Genetic Origin.**
Organizers: A. Fischer and A. Arnaiz-Villena.
- 39 **Workshop on Molecular Basis for Biodegradation of Pollutants.**
Organizers: K. N. Timmis and J. L. Ramos.
- *40 **Workshop on Nuclear Oncogenes and Transcription Factors in Hematopoietic Cells.**
Organizers: J. León and R. Eisenman.

- *41 **Workshop on Three-Dimensional Structure of Biological Macromolecules.**
Organizers: T. L. Blundell, M. Martínez-Ripoll, M. Rico and J. M. Mato.
- 42 **Workshop on Structure, Function and Controls in Microbial Division.**
Organizers: M. Vicente, L. Rothfield and J. A. Ayala.
- *43 **Workshop on Molecular Biology and Pathophysiology of Nitric Oxide.**
Organizers: S. Lamas and T. Michel.
- *44 **Workshop on Selective Gene Activation by Cell Type Specific Transcription Factors.**
Organizers: M. Karin, R. Di Lauro, P. Santisteban and J. L. Castrillo.
- 45 **Workshop on NK Cell Receptors and Recognition of the Major Histocompatibility Complex Antigens.**
Organizers: J. Strominger, L. Moretta and M. López-Botet.
- 46 **Workshop on Molecular Mechanisms Involved in Epithelial Cell Differentiation.**
Organizers: H. Beug, A. Zweibaum and F. X. Real.
- 47 **Workshop on Switching Transcription in Development.**
Organizers: B. Lewin, M. Beato and J. Modolell.
- 48 **Workshop on G-Proteins: Structural Features and Their Involvement in the Regulation of Cell Growth.**
Organizers: B. F. C. Clark and J. C. Lacal.
- *49 **Workshop on Transcriptional Regulation at a Distance.**
Organizers: W. Schaffner, V. de Lorenzo and J. Pérez-Martín.
- 50 **Workshop on From Transcript to Protein: mRNA Processing, Transport and Translation.**
Organizers: I. W. Mattaj, J. Ortín and J. Valcárcel.
- 51 **Workshop on Mechanisms of Expression and Function of MHC Class II Molecules.**
Organizers: B. Mach and A. Celada.
- 52 **Workshop on Enzymology of DNA-Strand Transfer Mechanisms.**
Organizers: E. Lanka and F. de la Cruz.
- 53 **Workshop on Vascular Endothelium and Regulation of Leukocyte Traffic.**
Organizers: T. A. Springer and M. O. de Landázuri.
- 54 **Workshop on Cytokines in Infectious Diseases.**
Organizers: A. Sher, M. Fresno and L. Rivas.
- 55 **Workshop on Molecular Biology of Skin and Skin Diseases.**
Organizers: D. R. Roop and J. L. Jorcano.
- 56 **Workshop on Programmed Cell Death in the Developing Nervous System.**
Organizers: R. W. Oppenheim, E. M. Johnson and J. X. Comella.
- 57 **Workshop on NF- κ B/I κ B Proteins. Their Role in Cell Growth, Differentiation and Development.**
Organizers: R. Bravo and P. S. Lazo.
- 58 **Workshop on Chromosome Behaviour: The Structure and Function of Telomeres and Centromeres.**
Organizers: B. J. Trask, C. Tyler-Smith, F. Azorín and A. Villasante.
- 59 **Workshop on RNA Viral Quasispecies.**
Organizers: S. Wain-Hobson, E. Domingo and C. López Galíndez.
- 60 **Workshop on Abscissic Acid Signal Transduction in Plants.**
Organizers: R. S. Quatrano and M. Pagès.
- 61 **Workshop on Oxygen Regulation of Ion Channels and Gene Expression.**
Organizers: E. K. Weir and J. López-Barneo.
- 62 **1996 Annual Report**
- 63 **Workshop on TGF- β Signalling in Development and Cell Cycle Control.**
Organizers: J. Massagué and C. Bernabéu.
- 64 **Workshop on Novel Biocatalysts.**
Organizers: S. J. Benkovic and A. Ballesteros.

- 65 **Workshop on Signal Transduction in Neuronal Development and Recognition.**
Organizers: M. Barbacid and D. Pulido.
- 66 **Workshop on 100th Meeting: Biology at the Edge of the Next Century.**
Organizer: Centre for International Meetings on Biology, Madrid.
- 67 **Workshop on Membrane Fusion.**
Organizers: V. Malhotra and A. Velasco.
- 68 **Workshop on DNA Repair and Genome Instability.**
Organizers: T. Lindahl and C. Pueyo.
- 69 **Advanced course on Biochemistry and Molecular Biology of Non-Conventional Yeasts.**
Organizers: C. Gancedo, J. M. Siverio and J. M. Cregg.
- 70 **Workshop on Principles of Neural Integration.**
Organizers: C. D. Gilbert, G. Gasic and C. Acuña.
- 71 **Workshop on Programmed Gene Rearrangement: Site-Specific Recombination.**
Organizers: J. C. Alonso and N. D. F. Grindley.
- 72 **Workshop on Plant Morphogenesis.**
Organizers: M. Van Montagu and J. L. Micol.
- 73 **Workshop on Development and Evolution.**
Organizers: G. Morata and W. J. Gehring.
- *74 **Workshop on Plant Viroids and Viroid-Like Satellite RNAs from Plants, Animals and Fungi.**
Organizers: R. Flores and H. L. Sänger.
- 75 **1997 Annual Report.**
- 76 **Workshop on Initiation of Replication in Prokaryotic Extrachromosomal Elements.**
Organizers: M. Espinosa, R. Díaz-Orejas, D. K. Chattoraj and E. G. H. Wagner.
- 77 **Workshop on Mechanisms Involved in Visual Perception.**
Organizers: J. Cudeiro and A. M. Sillito.
- 78 **Workshop on Notch/Lin-12 Signalling.**
Organizers: A. Martínez Arias, J. Modolell and S. Campuzano.
- 79 **Workshop on Membrane Protein Insertion, Folding and Dynamics.**
Organizers: J. L. R. Arrondo, F. M. Goñi, B. De Kruijff and B. A. Wallace.
- 80 **Workshop on Plasmodesmata and Transport of Plant Viruses and Plant Macromolecules.**
Organizers: F. García-Arenal, K. J. Oparka and P. Palukaitis.
- 81 **Workshop on Cellular Regulatory Mechanisms: Choices, Time and Space.**
Organizers: P. Nurse and S. Moreno.
- 82 **Workshop on Wiring the Brain: Mechanisms that Control the Generation of Neural Specificity.**
Organizers: C. S. Goodman and R. Gallego.
- 83 **Workshop on Bacterial Transcription Factors Involved in Global Regulation.**
Organizers: A. Ishihama, R. Kolter and M. Vicente.
- 84 **Workshop on Nitric Oxide: From Discovery to the Clinic.**
Organizers: S. Moncada and S. Lamas.
- 85 **Workshop on Chromatin and DNA Modification: Plant Gene Expression and Silencing.**
Organizers: T. C. Hall, A. P. Wolffe, R. J. Ferl and M. A. Vega-Palas.
- 86 **Workshop on Transcription Factors in Lymphocyte Development and Function.**
Organizers: J. M. Redondo, P. Matthias and S. Pettersson.
- 87 **Workshop on Novel Approaches to Study Plant Growth Factors.**
Organizers: J. Schell and A. F. Tiburcio.
- 88 **Workshop on Structure and Mechanisms of Ion Channels.**
Organizers: J. Lerma, N. Unwin and R. MacKinnon.
- 89 **Workshop on Protein Folding.**
Organizers: A. R. Fersht, M. Rico and L. Serrano.

- 90 **1998 Annual Report.**
- 91 **Workshop on Eukaryotic Antibiotic Peptides.**
Organizers: J. A. Hoffmann, F. García-Olmedo and L. Rivas.
- 92 **Workshop on Regulation of Protein Synthesis in Eukaryotes.**
Organizers: M. W. Hentze, N. Sonenberg and C. de Haro.
- 93 **Workshop on Cell Cycle Regulation and Cytoskeleton in Plants.**
Organizers: N.-H. Chua and C. Gutiérrez.
- 94 **Workshop on Mechanisms of Homologous Recombination and Genetic Rearrangements.**
Organizers: J. C. Alonso, J. Casadesús, S. Kowalczykowski and S. C. West.
- 95 **Workshop on Neutrophil Development and Function.**
Organizers: F. Mollinedo and L. A. Boxer.
- 96 **Workshop on Molecular Clocks.**
Organizers: P. Sassone-Corsi and J. R. Naranjo.
- 97 **Workshop on Molecular Nature of the Gastrula Organizing Center: 75 years after Spemann and Mangold.**
Organizers: E. M. De Robertis and J. Aréchaga.
- 98 **Workshop on Telomeres and Telomerase: Cancer, Aging and Genetic Instability.**
Organizer: M. A. Blasco.
- 99 **Workshop on Specificity in Ras and Rho-Mediated Signalling Events.**
Organizers: J. L. Bos, J. C. Lacal and A. Hall.
- 100 **Workshop on the Interface Between Transcription and DNA Repair, Recombination and Chromatin Remodelling.**
Organizers: A. Aguilera and J. H. J. Hoeijmakers.
- 101 **Workshop on Dynamics of the Plant Extracellular Matrix.**
Organizers: K. Roberts and P. Vera.
- 102 **Workshop on Helicases as Molecular Motors in Nucleic Acid Strand Separation.**
Organizers: E. Lanka and J. M. Carazo.
- 103 **Workshop on the Neural Mechanisms of Addiction.**
Organizers: R. C. Malenka, E. J. Nestler and F. Rodríguez de Fonseca.
- 104 **1999 Annual Report.**
- 105 **Workshop on the Molecules of Pain: Molecular Approaches to Pain Research.**
Organizers: F. Cervero and S. P. Hunt.
- 106 **Workshop on Control of Signalling by Protein Phosphorylation.**
Organizers: J. Schlessinger, G. Thomas, F. de Pablo and J. Moscat.
- 107 **Workshop on Biochemistry and Molecular Biology of Gibberellins.**
Organizers: P. Hedden and J. L. García-Martínez.
- 108 **Workshop on Integration of Transcriptional Regulation and Chromatin Structure.**
Organizers: J. T. Kadonaga, J. Ausió and E. Palacián.
- 109 **Workshop on Tumor Suppressor Networks.**
Organizers: J. Massagué and M. Serrano.
- 110 **Workshop on Regulated Exocytosis and the Vesicle Cycle.**
Organizers: R. D. Burgoyne and G. Álvarez de Toledo.
- 111 **Workshop on Dendrites.**
Organizers: R. Yuste and S. A. Siegelbaum.
- 112 **Workshop on the Myc Network: Regulation of Cell Proliferation, Differentiation and Death.**
Organizers: R. N. Eisenman and J. León.
- 113 **Workshop on Regulation of Messenger RNA Processing.**
Organizers: W. Keller, J. Ortín and J. Valcárcel.
- 114 **Workshop on Genetic Factors that Control Cell Birth, Cell Allocation and Migration in the Developing Forebrain.**
Organizers: P. Rakic, E. Soriano and A. Álvarez-Buylla.

- 115 **Workshop on Chaperonins: Structure and Function.**
Organizers: W. Baumeister, J. L. Carras-cosa and J. M. Valpuesta.
- 116 **Workshop on Mechanisms of Cellular Vesicle and Viral Membrane Fusion.**
Organizers: J. J. Skehel and J. A. Melero.
- 117 **Workshop on Molecular Approaches to Tuberculosis.**
Organizers: B. Gicquel and C. Martín.
- 118 **2000 Annual Report.**
- 119 **Workshop on Pumps, Channels and Transporters: Structure and Function.**
Organizers: D. R. Madden, W. Kühlbrandt and R. Serrano.
- 120 **Workshop on Common Molecules in Development and Carcinogenesis.**
Organizers: M. Takeichi and M. A. Nieto.
- 121 **Workshop on Structural Genomics and Bioinformatics.**
Organizers: B. Honig, B. Rost and A. Valencia.
- 122 **Workshop on Mechanisms of DNA-Bound Proteins in Prokaryotes.**
Organizers: R. Schleif, M. Coll and G. del Solar.
- 123 **Workshop on Regulation of Protein Function by Nitric Oxide.**
Organizers: J. S. Stamler, J. M. Mato and S. Lamas.
- 124 **Workshop on the Regulation of Chromatin Function.**
Organizers: F. Azorín, V. G. Corces, T. Kouzarides and C. L. Peterson.
- 125 **Workshop on Left-Right Asymmetry.**
Organizers: C. J. Tabin and J. C. Izpisua Belmonte.
- 126 **Workshop on Neural Patterning and Specification.**
Organizers: K. G. Storey and J. Modolell.
- 127 **Workshop on Signalling at the Growth Cone.**
Organizers: E. R. Macagno, P. Bovolenta and A. Ferrús.
- 128 **Workshop on Molecular Basis of Ionic Homeostasis and Salt Tolerance in Plants.**
Organizers: E. Blumwald and A. Rodríguez-Navarro.
- 129 **Workshop on Cross Talk Between Cell Division Cycle and Development in Plants.**
Organizers: V. Sundaresan and C. Gutiérrez.
- 130 **Workshop on Molecular Basis of Human Congenital Lymphocyte Disorders.**
Organizers: H. D. Ochs and J. R. Regueiro.
- 131 **Workshop on Genomic vs Non-Genomic Steroid Actions: Encountered or Unified Views.**
Organizers: M. G. Parker and M. A. Valverde.
- 132 **2001 Annual Report.**
- 133 **Workshop on Stress in Yeast Cell Biology... and Beyond.**
Organizer: J. Ariño.
- 134 **Workshop on Leaf Development.**
Organizers: S. Hake and J. L. Micol.
- 135 **Workshop on Molecular Mechanisms of Immune Modulation: Lessons from Viruses.**
Organizers: A. Alcami, U. H. Koszinowski and M. Del Val.
- 136 **Workshop on Channelopathies.**
Organizers: T. J. Jentsch, A. Ferrer-Montiel and J. Lerma.
- 137 **Workshop on Limb Development.**
Organizers: D. Duboule and M. A. Ros.
- 138 **Workshop on Regulation of Eukaryotic Genes in their Natural Chromatin Context.**
Organizers: K. S. Zaret and M. Beato.
- 139 **Workshop on Lipid Signalling: Cellular Events and their Biophysical Mechanisms.**
Organizers: E. A. Dennis, A. Alonso and I. Varela-Nieto.
- 140 **Workshop on Regulation and Functional Insights in Cellular Polarity.**
Organizers: A. R. Horwitz and F. Sánchez-Madrid.

- 141 **Workshop on The Structure of the Cortical Microcircuit.**
Organizers: R. Yuste, E. M. Callaway and H. Markram.
- 142 **Workshop on Control of NF- κ B Signal Transduction in Inflammation and Innate Immunity.**
Organizers: M. Karin, I. M. Verma and J. Moscat.
- 143 **Workshop on Engineering RNA Virus Genomes as Biosafe Vectors.**
Organizers: C. M. Rice, W. J. M. Spaan and L. Enjuanes.
- 144 **Workshop on Exchange Factors.**
Organizers: X. R. Bustelo, J. S. Gutkind and P. Crespo.
- 145 **Workshop on the Ubiquitin-Proteasome System.**
Organizers: A. Ciechanover, D. Finley, T. Sommer and C. Mezquita.
- 146 **Workshop on Manufacturing Bacteria: Design, Production and Assembly of Cell Division Components.**
Organizers: P. de Boer, J. Errington and M. Vicente.
- 147 **2002. Annual Report.**
- 148 **Workshop on Membranes, Trafficking and Signalling during Animal Development.**
Organizers: K. Simons, M. Zerial and M. González-Gaitán.
- 149 **Workshop on Synaptic Dysfunction and Schizophrenia.**
Organizers: P. Levitt, D. A. Lewis and J. DeFelipe.
- 150 **Workshop on Plasticity in Plant Morphogenesis.**
Organizers: G. Coupland, C. Fankhauser and M. A. Blázquez.
- 151 **Workshop on Wnt Genes and Wnt Signalling.**
Organizers: J. F. de Celis, J. C. Izpizúa Belmonte and R. Nusse.
- 152 **Workshop on Molecular and Genetic Basic of Autoimmune Diseases: SLE and RA.**
Organizers: A. Coutinho, W. Haas and C. Martínez-A.

*: Out of Stock.

The Centre for International Meetings on Biology
was created within the
Instituto Juan March de Estudios e Investigaciones,
a private foundation specialized in scientific activities
which complements the cultural work
of the *Fundación Juan March*.

The Centre endeavours to actively and
systematically promote cooperation among Spanish
and foreign scientists working in the field of Biology,
through the organization of Workshops, Lecture
Courses, Seminars and Symposia.

From 1989 through 2002,
a total of 176 meetings,
all dealing with a wide range of
subjects of biological interest,
were organized within the
scope of the Centre.



Instituto Juan March de Estudios e Investigaciones
Castelló, 77 • 28006 Madrid (España)
Tel. 34 91 435 42 40 • Fax 34 91 576 34 20
<http://www.march.es/biology>

The lectures summarized in this publication were presented by their authors at a workshop held on the 12th through the 14th of May, 2003, at the Instituto Juan March.

All published articles are exact reproduction of author's text.

There is a limited edition of 400 copies of this volume, available free of charge.