Instituto Juan March de Estudios e Investigaciones

149 CENTRO DE REUNIONES INTERNACIONALES SOBRE BIOLOGÍA

Workshop on

Synaptic Dysfunction and Schizophrenia

Organized by

P. Levitt, D. A. Lewis and J. DeFelipe

G. Alvarez de Toledo
Y. Ben-Ari
J. P. Bourgeois
H. T. Cline
J. DeFelipe
J. M. Fritschy
P. Gaspar
P. S. Goldman-Rakic
S. Heckers

M. Laruelle J. Lerma P. Levitt D. A. Lewis J. A. Lieberman K. Mirnics P. H. Patterson T. C. Südhof R. Yuste

IJM 149 Wor



13M-149-Wor

Instituto Juan March de Estudios e Investigaciones

149 CENTRO DE REUNIONES INTERNACIONALES SOBRE BIOLOGÍA

Workshop on Synaptic Dysfunction and Schizophrenia



Organized by

P. Levitt, D. A. Lewis and J. DeFelipe

G. Alvarez de Toledo
Y. Ben-Ari
J. P. Bourgeois
H. T. Cline
J. DeFelipe
J. M. Fritschy
P. Gaspar
P. S. Goldman-Rakic
S. Heckers

M. Laruelle J. Lerma P. Levitt D. A. Lewis J. A. Lieberman K. Mirnics P. H. Patterson T. C. Südhof R. Yuste

The lectures summarized in this publication were presented by their authors at a workshop held on the 10th through the 12th of February, 2003, at the Instituto Juan March.

Depósito legal: M-14004 / 2003 Impresión: Ediciones Peninsular. Tomelloso, 27. 28026 Madrid

INDEX

Introduction: P. Levitt, D. A. Lewis, J. DeFelipe	7
Session 1: Cortical inhibitory circuitry and its dysfunction in schizophrenia Chair: Patricia S. Goldman-Rakic	11
Javier DeFelipe: Human GABAergic cortical microcircuitry: catecholaminergic interneurons	13
Jean-Marc Fritschy: Significance of GABA _A receptor heterogeneity in cortical circuits	15
David A. Lewis: Selective alterations in cortical GABA circuits in schizophrenia.	18
Short talk: Farid Hamzei-Sichani: Factor analysis classification and outlier detection of cortical neurons	20
Session 2: Neuromodulators and their alterations in schizophrenia Chair: David A. Lewis	21
Patricia S. Goldman-Rakic: Prefrontal cortex: Structural and functional aspects of cognitive dysfunction in schizophrenia.	23
Marc Laurelle: Imaging dopamine transmission with Positron Emission Tomography in functional subdivision of the human striatum	25
Patricia Gaspar: Keeping the balance of 5-HT in the developing brain: a tightly controlled process.	26
Jeffrey A. Lieberman: Antipsychotic treatment effects on the clinical and pathophysiological course of brain pathomorphology in schizophrenia	27
Short talk: Vicente Molina: NAA levels in the dorsolateral prefrontal region in the first years of schizophrenia are inversely related to disease duration	28
Session 3: Synaptic function: Understanding the pathophysiology of schizophrenia Chair: Patricia Gaspar	29
Guillermo Alvarez de Toledo: New approaches to synaptic vesicle exocytosis and endocytosis	(Madrid)

PAGE

Juan Lerma: Glutamate signalling and synaptic transmission	32
Yehezkel Ben-Ari: Basic developmental rules for the construction of cortical networks.	34
Károly Mirnics: Schizophrenia as a disease of the synapse: Gene microarray studies.	36
Short talk: Maria Toledo-Rodríguez: Molecular profiles of anatomically and electrophysiologycally characterized neocortical neurons	38
Session 4: Development and plasticity: Role in the pathogenesis of schizophrenia Chair: Pat Levitt	39
Jean P. Bourgeois: Synaptogenesis in the mammalian neocortex	41
Hollis T. Cline: Determinants of pre- and postsynaptic interactions during development.	43
Rafael Yuste: Imaging the structure and function of dendritic spines	44
Stefan Heckers: Hippocampal structure and function in schizophrenia	45
Session 5: Models systems to investigate the underlying of etiology of schizophrenia Chair: Guillermo Alvarez de Toledo	47
Thomas C. Südhof: Molecular determinants of neurotransmitter release	47
	49
Short talk: Fiorenzo Conti: Increased GLT-1 expression in the prefrontal cortex of schizophrenics	50
Paul H. Patterson: A mouse model of mental illness based on a known risk factor.	51
Pat Levitt: Determinants of developmental trajectory: Models of early insult leading to delayed or long-lasting psychopathology	52
POSTERS	55
Eva Alés: Alantamine induces neurotransmitter release likely by blocking SK channels. Instituto Juan March (N	(ladrid)

PAGE

Ruth Benavides-Piccione: Catecholamine innervation of identified pyramidal neurons in the human temporal cortex.	58
Monica Beneyto: Cortical AMPA and kainate receptor subunit expression in schizophrenia, bipolar disorder, and major depressive disorder	59
Rafael Castro: Neurobiological and behavioural abnormalities induced by prenatal stimulation of dopamine D1 receptors	60
Philip Ebert: Altered gene expression of a cell adhesion molecule subfamily in schizophrenia	61
W. Gordon Frankle: The effect of antipsychotic medication on the occupancy of the D2 receptor by endogenous dopamine	62
Francisco G. Scholl: Lateral association of Neuroligin mediate synaptic adhesion with Neurexin and induce the formation of presynaptic terminals	64
Takanori Hashimoto: Altered gene expression for BDNF signaling in the prefrontal cortex of subjects with schizophrenia	65
Greta A. Herin: Evidence for an interaction between N-terminal redox sensitive cysteines and polyamine modulatory site of the NR1/NR2A NMDA receptor	66
Jean B. Manent: Neurotransmitters action before synaptogenesis: GABA and glutamate modulate the migration of hippocampal pyramidal cells	67
Marcello Melone: Effects of the antipsychotic drug clozapine on the expression and function of the glutamate transporters GLT-1, GLAST and EAAC1 in the frontal cortex of rats.	68
Patricia Murtra: C-fos expression after cocaine or morphine sensitization in CB1 knockout mice.	69
William A. Phillips: Reduced flow through NMDA-receptor channels as a basis for cognitive disorganisation in schizophrenia.	70
Margarita Rodríguez-Moral: Dopamine D1 receptors in the macaque monkey thalamus: selectivity for thalamic projection neurons	71
Lucia Tabares: Electrophysiological study of motor function in a-Synuclein null- mice	72

PAGE

Keith A. Young: Antigliosis: Evidence for glial effects on synaptic plasticity in schizophrenia	73
LIST OF INVITED SPEAKERS	75
LIST OF PARTICIPANTS	77

Introduction P. Levitt, D. A. Lewis and J. DeFelipe

The biological substrate of disorders of cognition, emotion and thought remains a central problem of clinical and basic neuroscience. Of the diseases with the greatest negative economic and health impact on the human population, 8 of the top 10 are neuropsychiatric in nature. Indeed, improvements in the treatment and prevention of these disorders are likely to have the greatest impact on reducing disability in the populations of developed countries in the future.

Converging evidence from studies in both humans and animals indicates that dysfunctional communication between specific populations of neurons may be the principal alteration in schizophrenia, a disease that afflicts approximately 1% of the world's population. Our basic understanding of the molecular and cellular mechanisms that mediate synaptic function has never been greater, providing an opportunity to integrate basic biological concepts of synaptic chemistry and physiology with disease-related alterations in the genes and proteins that mediate these functions.

We gathered investigators, who are experts in the fields of neuropsychiatry, synaptic function and neuronal communication, to exchange information that will facilitate a more sophisticated understanding of the biological bases of schizophrenia. While these participants had not before been assembled, we believe their interactions will promote the development of novel, integrative approaches to the study of the etiopathogenesis of schizophrenia and to the identification of new targets for therapeutic interventions.

The meeting was organized around five scientific themes to address the difficult challenge of translating our understanding of the clinical disorder into fundamental biological mechanisms that underlie the etiology and pathogenesis of schizophrenia: 1) Functional Architecture of Cortical Circuitry and Its Dysfunction in Schizophrenia. Schizophrenia involves the dysfunction of cortical microcircuits, vet we are still at a very early stage in a detailed understanding of the very cellular and physiological components that altered by the disease. Investigators who focus their studies on the functional attributes of circuitry of the cerebral cortex and thalamus are using technically sophisticated approaches to provide remarkable details of both macro- and microcircuitry. This framework allowed the group to translate the newest concepts towards an understanding of the disrupted circuitry in schizophrenia. 2) Neuromodulators and Their Alterations in Schizophrenia. Both basic and clinical research has implicated abnormal functioning of neuromodulatory systems, principally the monoamines, in schizophrenia. To date, the most prevalent and effective pharmacological treatments alter the activity of these neuromodulators. Investigators reported on the use of a variety of technical strategies, to elucidate the roles for these essential neurochemical components in mediating the functions that are abnormal in schizophrenia. Participants were provided with an opportunity to probe the relationship between observations of behavioral and physiological dysfunction in animal models and the functional features of these cortical systems in the context of schizophrenia. 3) Synaptic Function: Understanding the Pathophysiology of Schizophrenia. Accumulating evidence suggests that the dysfunction of synapses, the major element through which neurons interact, underlies the pathophysiology that characterizes schizophrenia. Sophisticated molecular and physiological approaches by scientists at the meeting facilitated remarkable progress Madrid)

defining the cellular and molecular components that are necessary to assemble and maintain mature, functional synapses. The current understanding of the presynaptic elements that control neurotransmitter release, and postsynaptic components that define the responsiveness of neurons in circuits will be placed in the context of new findings that implicate problems of the synaptic machinery in schizophrenia. 4) Development and Plasticity: Role in the Pathogenesis of Schizophrenia. The concept of a developmental etiology of schizophrenia was popularized during the last decade, but the theory has been difficult to crystallize from a cellular and molecular perspective of disrupted cortical circuits. In particular, the field wrestles with the polygenic and epigenetic nature of schizophrenia that is characterized by a significant temporal delay in the expression of overt symptoms of the disorder. These issues were addressed by investigators who have made seminal contributions to our understanding of synapse and receptor assembly in cerebral cortex, and the developmental and functional plasticity exhibited by these systems with particular genetic perturbations. This group was challenged with discussing the mechanisms that drive the assembly of dysfunctional circuits and the untapped potential for regulating plasticity as a strategy for intervention and treatment. 5) Testable Models and Strategies. The adaptive processes that occur in schizophrenia are likely to be initiated by a combination of genetic susceptibility and environmental perturbation. The issues discussed in the first four sessions provided the framework for the scientists in the last session to elaborate on new concepts of schizophrenia based on disruption of synaptic function. Testable models may provide new approaches to study the pathophysiology and to help define the etiology of the disease.

Thus, these scientists had the opportunity to integrate increasingly complex data from neuroimaging and circuit analyses, which continue to provide important details of the neural networks that are dysfunctional in schizophrenia, with the fundamental molecular and cellular processes of synaptic communication that form the basis for the disease. Rather than segregate investigators according to their specific technical approaches or broader laboratory interests, we purposely organized the sessions in which data on a key topic, extracted from studies of normal and disease states, was presented and discussed in an integrated fashion. Thus, each had broad representation and expertise. This approach facilitated the interactions among scientists with different backgrounds and perspectives, enhanced the crossfertilization of ideas from different disciplines, and emphasized the importance and opportunities for translating fundamental discoveries in synaptic function and cortical circuitry into strategies for understanding neuropsychiatric disorders.

Pat Levitt, David A. Lewis and Javier DeFelipe

Session 1: Cortical inhibitory circuitry and its dysfunction in schizophrenia Chair: Patricia S. Goldman-Rakic

Human GABAergic cortical microcircuitry: catecholaminergic interneurons

Javier DeFelipe

Cajal Institute, Madrid, Spain

In the cerebral cortex, information flows through synapses arranged in a highly organized network of intricate neuronal circuits. Modifications in this circuitry can lead to profound neurological and psychiatric diseases, and a number of alterations in cortical function probably arise from the impairment of specific intracortical circuits. Thus, it is fundamental to understand both normal and altered cortical circuitry for us to gain a better insight into cortical diseases.

Most of what is known about human neuronal circuitry is based on data obtained from the cortex of non-human mammals. More specifically, the majority of studies have been carried out on the primary sensory areas of the monkey, cat, rat and mouse. Although some aspects of neocortical organization appear to be universal across all species, at the microanatomical, molecular and hodological levels, there are significant differences between the primary and non-primary areas, and between homologous cortical areas in different species. Thus, relatively little is known about human neocortical circuitry. Cortical tissue removed during the course of surgery to treat patients with intractable epilepsy or to gain access to tumors located in subcortical structures, represents an excellent opportunity to study the human neocortex. This resected tissue can be immersed immediately in fixative, eliminating the influence of postmortem factors. Furthermore, the development of certain microanatomical tools, and a variety of light and electron immunocytochemical techniques, has now made it feasible to perform detailed studies of cortical microganization directly in the human cerebral cortex. As a result, novel aspects of neurochemical and microanatomical human cortical organization are being revealed.

Processed information leaves the cortex along the axons of pyramidal cells to reach its destination in other cortical areas or subcortical nuclei. Our current understanding of the synaptic organization of the cerebral cortex, and of how this information flow occurs, depends to a large extent on our knowledge of the synaptic inputs to pyramidal cells. In general, it is considered that the basic cortical microcircuit is composed of a pyramidal cell and its inputoutput connections. Inhibitory inputs, which mostly originate from GABAergic interneurons, terminate on the dendrites, soma and initial axon segment of pyramidal cells. Amongst the cortical interneurons, one group has been identified that contain tyrosine hydroxylase (TH), the rate-limiting enzyme in catecholamine synthesis. Because catecholamines have been shown to be involved in schizophrenia, the study of these interneurons may be particularly relevant for this disease. It is particularly interesting that in the human, these TH neurons have a peculiar laminar distribution and charateristic neurochemical attributes. Notably, they are found almost exclusively in layers V-VI and only 50% of them contain GABA. We found that neurons that contain TH do not contribute to three of the major GABA-ergic interneuron populations: double bouquet cells, chandelier cells and basket cells. We also estimated that 26% of TH neurons also contain nitric oxide synthase. In addition, we observed that TH fibers preferentially form connections with dendrites, and that pyramidal cells filled with Lucifer Yellow and reconstructed from serial confocal microscope images received a remarkable constant number of contacts per neuron from TH axons (15 to 31; average of 23). As previous studies have implicated both nitrergic neurons in deep cortical layers and the cathecolaminergic system in the pathogenesis of schizophrenia, it will be of interest to study these TH neurons and their innervation of pyramidal cells to determine whether they are specifically affected in schizophrenic patients.

Significance of GABA_A receptor heterogeneity in cortical circuits

Jean-Marc Fritschy

Institute of Pharmacology and Toxicology, University of Zurich, CH – 8057 Switzerland

GABA_A receptors mediate most of the fast inhibitory neurotransmission in vertebrate CNS by gating Cl⁻ ions through an integral membrane channel. GABA_A receptors form multimeric complexes assembled from a family of at least 21 constituent subunits (α 1-6, β 1-4, γ 1-4, δ , ρ 1-3, θ , π) (Barnard et al., 1998). The subunit composition and stoichiometry of native GABA_A receptors have not been elucidated but evidence favors the existence of pentameric complexes containing $2\alpha/2\beta/1\gamma$ subunit variants. Immunochemical, pharmacological, and functional analyses of GABA_A receptors give convergent results that the majority contains a single type of α and β subunit variant, with the $\alpha 1\beta 2\gamma 2$ combination representing the largest population of GABA_A receptors, followed by $\alpha 2\beta 3\gamma 2$ and $\alpha 3\beta 3\gamma 2$. Receptors containing the $\alpha 4$, $\alpha 5$, or $\alpha 6$ subunit, as well as the $\beta 1$, $\gamma 1$, $\gamma 3$, δ , π and θ subunit, form minor receptor populations.

Pharmacological analysis of GABAA receptors immunoprecipitated with antibodies against specific α subunit variants allows differentiating between GABA receptor subtypes. The α 1-, α^2 -, α^3 -, and α^5 -GABA_A receptors correspond to diazepam-sensitive receptors, whereas α 4- and α 6-GABAA receptors are insensitive to diazepam (Benson et al., 1998). Functionally, distinct subunit-specific properties have been identified in both recombinant and native receptors, supporting the concept that $GABA_A$ receptor heterogeneity is a major facet determining the functional properties of GABAergic inhibitory circuits. In particular, the type of α subunit determines the kinetics of receptor deactivation and the presence of the δ subunit results in markedly increased agonist affinity and apparent lack of desensitization. Immunohistochemical analyses based on the visualization of α subunit variants revealed a region- and neuron-specific distribution pattern of GABAA receptor subtypes that is largely conserved across species, suggesting that individual subtypes are present in distinct neuronal circuits. The specificity of subtype expression is underscored by the remarkable selectivity of action of diazepam in knock-in mutant mice carrying "custom-made" diazepam-insensitive GABAA receptor subtypes. Thus, abolition of diazepam binding on a2-GABAA receptors in vivo by exchanging the conserved histidine 101 residue with an arginine residue results in selective suppression of the anxiolytic action of this drug, whereas anxiolytic drugs acting by other mechanisms are unaffected (Löw et al., 2000). This finding indicates that a2-GABAA receptors are strategically located in circuits mediating the anxiolytic action of diazepam. Likewise, the sedative effects of diazepam are abolished, even at high dose, in mice carrying $\alpha 1^{H101R}$ -GABA_A receptors, while its anxiolytic action is fully retained in these mice (Rudolph et al., 1999; McKernan et al., 2000). Therefore, brain circuits containing α1-GABAA

receptors mediate sedation and activation of these receptors by diazepam is ineffective for relieving anxiety.

The cellular corollary of the specificity of diazepam action is the synapse-specific distribution of GABAA receptor subtypes, in particular in neurons expressing multiple GABAA receptor subtypes, such as hippocampal pyramidal neurons. These cells express a high level of $\alpha 1$, $\alpha 2$, and $\alpha 5$ subunit, along with $\beta 1$ -3 and $\gamma 2$ subunit, suggesting that they form at least three main GABAA receptor subtypes. al-GABAA receptors are located postsynaptically in a majority of somatodendritic synapses and to a lesser extent in the axon initial segment; in contrast, a2-GABAA receptors are particularly abundant in the axon initial segment and are only few in somatodendritic synapses (Nusser et al., 1996). Finally, a5-GABAA receptors have an extrasynaptic localization, being distributed throughout the somatodendritic compartment of hippocampal pyramidal cells without being aggregated at postsynaptic sites. Strikingly, in the soma of hippocampal pyramidal cells, $\alpha 1$ - and $\alpha 2$ -GABAA receptor subtypes are segregated to distinct synapses formed by two separate populations of basket cells. Parvalbumin-positive basket cells form GABAergic synapses containing the al subunit, whereas cholecystokinin-positive basket cells form GABAergic synapses containing the a2 subunit (Nyiri et al., 2001; Klausberger et al., 2002), suggesting that presynaptic afferents modulate targeting of receptor subtypes to specific types of synapses. Additional evidence, notably the presence of CB1 cannabinoid receptors in CCKpositive terminals (Katona et al., 1999), indicates that these two types of basket cell correspond to distinct neuronal circuits.

Finally, a substantial proportion of GABA_A receptors are found extrasynaptically, diffusely distributed in somato-dendritic membranes. These receptors likely contribute to normal GABAergic inhibition in vivo and are involved in the expression of specific behaviors. This was shown by in mice with targeted mutations of the α 5 subunit gene (Collinson et al., 2002; Crestani et al., 2002). For example, a selective role of α 5-GABA_A receptors in hippocampal-dependent tasks was reported in a mouse line carrying a H105R point mutation in the α 5 subunit gene. Unexpectedly, this mutation caused a profound reduction of α 5-GABA_A receptors selectively in the hippocampal formation and was correlated with altered performance in a delay trace-conditioning task (Crestani et al., 2002). In contrast, learning of non-hippocampus-dependent behavioral tasks was not affected, pointing to a selective role of extrasynaptic GABA_A receptors for proper hippocampal function.

In the neocortex, although the details of synaptic circuits remain largely to be worked out, similar arrangements are very likely. In addition, a distinct laminar and areal segregation of GABA_A receptor subtypes is evident, suggesting the contribution of different receptor subtypes in cortico-cortical and cortico-subcortical circuits (Fritschy & Mohler, 1995). Major differences are also seen in the thalamo-recipient layers between primary and secondary sensory areas. The adult pattern is established during the period of synaptogenesis, with major alterations in the expression and cellular distribution of receptor subtypes, indicating

that a given subunit can have distinct functions at various stages of brain development (Paysan et al., 1997).

The significance of GABA_A receptor function is underscored by the multiple neurological and psychiatric diseases for which an alteration in the GABAergic system has been postulated, including epilepsy, anxiety disorders, ethanol dependence, Huntington's disease, Angelman syndrome, and schizophrenia. It is therefore of crucial importance to unravel the mechanisms underlying the selective expression of GABA_A receptor subtypes in specific neuronal circuits under normal and pathological conditions.

References:

Barnard, E. A., Skolnick, P., Olsen, R. W., Mohler, H., Sieghart, W., Biggio, G., Braestrup, C., Bateson, A. N., & Langer, S. Z. (1998). International Union of Pharmacology. XV. Subtypes of γ -aminobutyric acid_A receptors: classification on the basis of subunit structure and function. *Pharmacol Rev 50*, 291-313.

Benson, J. A., Low, K., Keist, R., Mohler, H., & Rudolph, U. (1998). Pharmacology of recombinant γ -aminobutyric acid_A receptors rendered diazepam-insensitive by point-mutated alpha-subunits. *FEBS Lett 431*, 400-404.

Collinson, N., Kuenzi, F. M., Jarolimek, W., Maubach, K. A., Cothliff, R., Sur, C., Smith, A. D., Otu, F. M., Howell, O., Atack, J. R., McKernan, R. M., Seabrook, G. R., Dawson, G. R., Whiting, P. J., & Rosahl, T. W. (2002). Enhanced learning and memory and altered GABAergic synaptic transmission in mice lacking the α 5 subunit of the GABA_A receptor. *J Neurosci 22*, 5572-5580.

Crestani, F., Keist, R., Fritschy, J. M., Benke, D., Vogt, K., Prut, L., Bluethmann, H., Mohler, H., & Rudolph, U. (2002). Trace fear conditioning involves hippocampal α 5 GABA_A receptors. *Proc Natl Acad Sci USA 99*, 8980-8985.

Fritschy, J. M., & Mohler, H. (1995). GABA_A-receptor heterogeneity in the adult rat brain: differential regional and cellular distribution of seven major subunits. *J Comp Neurol* 359, 154-194.

Katona, I., Sperlágh, B., Sík, A., Käfalvi, A., Vizi, E. S., Mackie, K., & Freund, T. F. (1999). Presynaptically located CB1 cannabinoid receptors regulate GABA release from axon terminals of specific hippocampal interneurons. *J Neurosci 19*, 4544-4558.

Klausberger, T., Roberts, J. D., & Somogyi, P. (2002). Cell type- and input-specific differences in the number and subtypes of synaptic GABA_A receptors in the hippocampus. *J Neurosci* 22, 2513-2521.

Löw, K., Crestani, F., Keist, R., Benke, D., Brünig, I., Benson, J. A., Fritschy, J. M., Rulicke, T., Bluethmann, H., Mohler, H., & Rudolph, U. (2000). Molecular and neuronal substrate for the selective attenuation of anxiety. *Science 290*, 131-134.

McKernan, R. M., Rosahl, T. W., Reynolds, D. S., Sur, C., Wafford, K. A., Atack, J. R., Farrar, S., Myers, J., Cook, G., Ferris, P., Garrett, L., Bristow, L., Marshall, G., Macaulay, A., Brown, N., Howell, O., Moore, K. W., Carling, R. W., Street, L. J., Castro, J. L., Ragan, C. I., Dawson, G. R., & Whiting, P. J. (2000). Sedative but not anxiolytic properties of benzodiazepines are mediated by the GABA_A receptor α1 subtype. *Nature Neurosci 3*, 587-592.

Nusser, Z., Sieghart, W., Benke, D., Fritschy, J. M., & Somogyi, P. (1996). Differential synaptic localization of two major γ-aminobutyric acid type A receptor alpha subunits on hippocampal pyramidal neurons. *Proc Natl Acad Sci USA 93*, 11939-11944.

Nyiri, G., Freund, T. F., & Somogyi, P. (2001). Input-dependent synaptic targeting of α 2-subunit-containing GABA_A receptors in synapses of hippocampal pyramidal cells of the rat. *Eur J Neurosci 13*, 428-442.

Paysan, J., Kossel, A., Bolz, J., & Fritschy, J. M. (1997). Area-specific regulation of γ-aminobutyric acid A receptor subtypes by thalamic afferents in developing rat neocortex. *Proc Natl Acad Sci USA 94*, 6995-7000.

Rudolph, U., Crestani, F., Benke, D., Brünig, I., Benson, J., Fritschy, J. M., Martin, J. R., Bluethmann, H., & Mohler, H. (1999). Benzodiazepine actions mediated by specific γ -aminobutyric acid_A receptor subtypes. *Nature* 401, 796-800.

Selective alterations in cortical GABA circuits in schizophrenia

David A. Lewis

Departments of Psychiatry and Neuroscience University of Pittsburgh

Critical cognitive deficits in schizophrenia are associated with dysfunction of the dorsolateral prefrontal cortex (DLPFC). In addition, markers of GABA neuron transmission are altered in the DLPFC of subjects with schizophrenia. Understanding the contribution of altered GABA neurotransmission to the cognitive deficits of schizophrenia requires knowledge of the functional circuitry of the different classes of DLPFC GABA neurons, of the changes that these neurons undergo during developmental periods of relevance to the pathophysiology of schizophrenia, and of the identification of the specific populations of GABA neurons and circuits that are altered in schizophrenia. This presentation will describe recent data from these three types of investigations that converge upon the parvalbumin (PV)-containing class of DLPFC GABA neurons as being particularly vulnerable in schizophrenia.

First, in contrast to the calretinin-containing subset of GABA neurons, PV-containing cells receive inputs from dopamine axons, the mediodorsal thalamus, and the axon collaterals of neighboring pyramidal cells, all of which appear to be involved in the working memory processes mediated by the DLPFC. In addition, electrophysiological studies in an *in vitro* slice preparation of monkey DLPFC indicate that these two classes of GABA neurons also differ in their responses to the patterns of sustained excitatory activity recorded *in vivo* in the DLPFC during working memory tasks.

Second, during both the neonatal and adolescent periods of development, substantial changes occur in markers of presynaptic neuronal activity, synaptic levels of GABA, and postsynaptic receptors for chandelier neurons and wide arbor neurons, the two principal classes of PV-containing GABA neurons.

Third, the PV-, but not the calretinin-containing, subclass of GABA neurons demonstrate decreased gene expression in subjects with schizophrenia, with at least some of these alterations evident at the level of the cognate proteins. These alterations are accompanied by an increase in the α_2 subunit of the GABA_A receptor localized to pyramidal neuron axon initial segments, the synaptic target of PV-containing chandelier neurons. In addition, the altered gene expression in PV-containing neurons is strongly associated with reductions in BDNF-trkB signaling, suggesting a possible mechanism for the GABA changes.

Thus, these data suggest that a subpopulation of GABA neurons are altered in the DLPFC of subjects with schizophrenia, that these alterations are associated with specific

circuitry changes that arise during development, and that these abnormalities may contribute to the types of working memory disturbances that are central to the cognitive deficits observed in schizophrenia. Understanding these abnormalities in the context of the affected neural circuits may reveal novel therapeutic strategies for ameliorating the cognitive dysfunction of schizophrenia.

References:

Woo T-U, Whitehead RE, Melchitzky DS, Lewis DA: A subclass of prefrontal -aminobutyric acid axon terminals are selectively altered in schizophrenia. *Proc Natl Acad Sci* USA 95:5341-5346, 1998.

Volk DW, Austin MC, Pierri JN, Sampson AR, Lewis DA: Decreased GAD₆₇ mRNA expression in a subset of prefrontal cortical GABA neurons in subjects with schizophrenia. *Arch Gen Psychiatry* 57:237-245, 2000.

Volk DW, Austin MC, Pierri JN, Sampson AR, Lewis DA: GABA transporter-1 mRNA in the prefrontal cortex in schizophrenia: Decreased expression in a subset of neurons. *Am J Psychiatry* 158:256-265, 2001.

Volk DW, Pierri JN, Fritschy J-N, Auh S, Sampson AR, Lewis DA: Reciprocal alterations in pre- and postsynaptic inhibitory markers at chandelier cell inputs to pyramidal neurons in schizophrenia. Cereb Cortex 12:1063-1070, 2002.

Erickson SL, Lewis DA: Postnatal development of parvalbumin- and GABA transporter-immunoreactive axon terminals in monkey prefrontal cortex. *J Comp Neurol* 448:186-202, 2002.

Factor analysis classification and outlier detection of cortical neurons

Hamzei-Sichani, F., Kozloski, J., Yuste, R.

Classification of neurons is of central importance to Neuroscience, yet this task has traditionally been done using qualitative criteria, which are not universally accepted. We have explored the use of quantitative methods to classify neocortical interneurons, using a morphological and electrophysiological database from 30 randomly selected interneurons from layer 5 of mouse primary visual cortex. The data was examined using a combination of multivariate techniques including factor analysis (FA) and principal component & classification analysis (PCCA). These techniques allowed us to study the underlying structure of the data (FA) and the characterization of the resultant clusters (PCCA). For the 30 neurons, 34 measured morphological and physiological variables produced 3 principal orthogonal factors in descending order of their contribution to the total variance of the data. The first factor detected variables related to action potential characteristics and spike train. The second factor selected morphological variables, whereas the third factor identified differences in after-hyperpolarizations. Using these three factors, application of PCCA resulted in the neurons being grouped in 3 unique clusters, composed of 26 of the 30 neurons. The three clusters were characterized by a combination of electrophysiological and morphological characteristics. One cluster was composed of fast spiking cells, another by low threshold spiking cells and the third was a mix. These techniques allowed us to use a quantitative unbiased analysis to define neuronal classes as an important first step in understanding neuronal circuits.

Session 2: Neuromodulators and their alterations in schizophrenia Chair: David A. Lewis

Prefrontal cortex: Structural and functional aspects of cognitive dysfunction in schizophrenia

P.S. Goldman-Rakic

Department of Neurobiology, Yale University School of Medicine, New Haven, CT 06510, USA

The stimulus-independent sustained activation of prefrontal neurons and their contentspecific coding of information constitute the fundamental cellular basis of the brain's working memory functions. We have proposed that a breakdown in the circuitry mediating these functions may be responsible for the cognitive deficits which are increasingly considered core problems in schizophrenia (Goldman-Rakic, 1987; 1994). The two properties – persistent neuronal firing – that outlasts a stimulus by seconds, and preferential coding of stimulus parameters derive both from biophysical properties and network dynamics. My laboratory is focused on elucidating these mechanisms as they pertain to and underpin human mental capacity and its dissolution in numerous disorders.

A first order hypothesis for understanding persistent activity in prefrontal cortex is that it is mediated, in part, by recurrent excitation among pyramidal neurons. We have used two methods to examine these interactions. Simultaneous recording of multiple neurons in monkeys trained on working memory tasks to address questions of functional interactions between pairs of pyramidal cells. Our findings from diverse studies have established that microcircuits exist for memory phenomena and that they are constrained both by local circuit organization and by content (Constantinidis et al., 2001). We have also studied the modulation of elemental neuronal interactions by dopamine both in cortical slices (Gao et al., 2001) and in the behaving animal (Williams et al., in preparation). Our findings indicate that dopamine depresses excitatory transmission between pairs of pyramidal neurons by actions at presynaptic D1 receptors both in vivo and in vitro. A presynaptic mechanism is supported by electronmicroscopic evidence that D1 receptors are located on axon terminals of excitatory afferents. These findings indicate, therefore, that a functional state of hypoglutamatergia can result from the actions of dopamine D1 receptors. Other equally important actions of dopamine in the prefrontal cortex arise from its modulation of inhibitory circuits that are engaged both in the spatial and temporal properties of prefrontal cognitive operations. (Rao et al., 1999; 2000; Constantinidis et al., 2002; Gao and Goldman-Rakic, in press).

The importance of the dopamine hypothesis of schizophrenia has been well known for half a century and will be further discussed in the context of recent studies on the existence of both D1 and D2 dopamine-receptor interacting proteins. As an example, the functional significance of D1 signaling mechanisms in schizophrenia has been highlighted by the recent finding that calcyon, an intracellular protein that interacts with D1 family receptors, is abnormally high in schizophrenic patients (Koh et al., in press). Further evidence from a recent genetic analysis has found that the region of chromosome 10 where the calcyon gene is located is at risk for schizophrenia or bipolar disorder (Jorgenson et al., 2002). Abnormalities in the prefrontal cortex of patients have also been observed in two recent PET studies of D1 receptor binding potential (Okubo et al., 1997; Abi-Dargham et al., 2002). Both studies found significant correlations between D1 binding potential and cognitive performance.

The studies to be reported are building a highly differentiated portrait of prefrontal functional circuitry and its dopamine-mediated signaling mechanisms which constitute the brain's information processing system.

References:

Abi-Dargham, A., Mawlawi, O., Lombardo, I., Gil, R., Martinez, D., Huang, Y., Hwang, D.R., Keilp, J., Kochan, L., Van Heertum, R., Gorman, J.M. and Laruelle, M. (2002) Prefrontal dopamine D1 receptors and working memory in schizophrenia. J. Neurosci. 22(9):3708-3719.

Constantinidis, C., Franowicz, M.N. and Goldman-Rakic, P.S. (2001) Coding specificity in cortical microcircuits: a multiple electrode analysis of primate prefrontal cortex. J. Neurosci. 21:3646-3655.

Constantinidis, C., Williams, G.V. and Goldman-Rakic, P.S. (2002) A role for inhibition in shaping the temporal flow of information in prefrontal cortex. Nat. Neurosci. 5:175-180.

Gao, W.-J., Krimer, L.S. and Goldman-Rakic, P.S. (2001) Presynaptic regulation of recurrent excitation by D1 receptors in prefrontal circuits. Proc. Natl. Acad. Sci. U.S.A. 98:295-300.

Gao. W.-J. and Goldman-Rakic, P.S. Differential dopamine modulation of peridendritic and perisonatic inhibition in prefrontal cortex. J. Neurosci., in press.

Goldman-Rakic, P.S. (1987) Circuitry of primate prefrontal cortex and regulation of behavior by representational memory. In Handbook of Physiology, The Nervous System, Higher Functions of the Brain (ed. Plum, F.), pp. 373-417. Am. Physiol. Soc., Bethesda, Maryland.

Goldman-Rakic, P.S. (1994) Working memory dysfunction in schizophrenia. J. Neuropsychiatry 6(4):348-357.

Jorgensen, T.H., Borglum, A.D., Mors, O., Wang, A.G., Pinaud, M., Flint, T.J., Dahl, H.A., Vang, M., Kruse, T.A., Ewald, H. (2002) Search for common haplotypes on chromosome 22q in patients with schizophrenia or bipolar disorder from the Faroe Islands. Am. J. Med. Genet. 114(2):245-252.

Koh, P.O., Bergson, C., Undie, A.S., Goldman-Rakic, P.S. and Lidow, M.S. Receptor-interacting proteins as sites of dopamine system dysfunction in schizophrenia: Upregulation of the D1-receptor-interacting protein, calcyon, in schizophrenic patients. Arch. Gen. Psychiat., in press.

Okubo, Y., Suhara, T., Suzuki, K., Kobayashi, K., Inoue, O., Terasaki, O., Someya, Y., Sassa, T., Sudo, Y., Matsushima, E., Iyo, M., Tateno, Y. and Toru, M. (1997) Decreased prefrontal dopamine D1 receptors in schizophrenia revealed by PET [see comments]. Nature 385(6617):634-636.

Rao, S.G., Williams, G.V. and Goldman-Rakic, P.S. (1999) Isodirectional tuning of adjacent interneurons and pyramidal cells during working memory: Evidence for microcolumnar organization in PFC. J. Neurophysiol. 81:1903-16.

Rao, S.G., Williams, G.V. and Goldman-Rakic, P.S. (2000) Destruction and creation of spatial tuning by disinhibition: GABA(A) blockade of prefrontal cortical neurons engaged by working memory. J. Neurosci. 20:485-94.

Imaging dopamine transmission with Positron Emission Tomography in functional subdivision of the human striatum

Marc Laruelle

Columbia University College of Physicians and Surgeons New York State Psychiatric Institute, Unit 31 1051 Riverside Drive New York, NY 10032 Phone: 212-5435388 Fax: 212-5686171 Email: ml393@columbia.edu

This presentation will illustrate how PET imaging, with state-of-the-art resolution and analytical techniques, provides a unique tool to study neurochemical pathways in the living human brain. The human striatum can be organized into limbic, associative and sensorimotor subdivisions, which process information related to emotional, cognitive and motor function. Dopamine (DA) projections ascending from the midbrain provide important modulatory input to these striatal subregions. The aim of this study was to compare dopamine transmission and stimulation of dopamine D2 receptors following the administration of amphetamine in these functional subdivisions of the human striatum. D2 receptor availability (V3") was measured with Positron Emission Tomography (PET) and [11C]raclopride in fourteen healthy volunteers under control conditions and following the intravenous administration of amphetamine (0.3 mg/kg). Amphetamine induced a significantly larger reduction in D2 receptor availability (ΔV_3) in limbic (ventral striatum, -15.3± 11.8%) and motor (postcommisural putamen, -16.2 \pm 9.6%) regions compared to associative regions (caudate and precommisural putamen, -8.1 \pm This region of interest analysis was then confirmed by a voxel-based analysis. 7.2%). Subjects were also asked to rate their subjective experience following amphetamine, and reported an increase in euphoria which was associated with a greater ΔV_3 " in limbic and motor regions, but not in the associative regions.

These results demonstrate significant differences in the DA response to amphetamine between the functional subdivisions of the human striatum. The mechanism behind these regional differences in amphetamine-induced DA release remain unclear, but could be related to the pattern of asymmetrical feed forward mediating the integration of limbic, cognitive, and motor function via the DA cell bodies in the ventral midbrain. The implication of these findings for the development of addictions and psychoses will be discussed.

Keeping the balance of 5-HT in the developing brain : a tightly controlled process

Patricia Gaspar, Tania Vitalis and Olivier Cases

INSERM U 106, Hôpital Salpêtrière, Paris, France

The developmental role of serotonin (5-HT) has been stressed since the early studies of Buznikov on sea urchin embryos. Subsequent studies in the mammalian central nervous reported that 5-HT had a wide variety of effects, on proliferation, migration, neurite outgrowth and synaptogenesis. These studies led however to a somewhat blurred picture of what 5-HT is actually doing because of the wide variety of 5HT receptors and the difficulty in obtaining complete and selective inactivation or stimulation of these receptors with classical pharmacological compounds. Molecular genetic approaches in the mouse have recently contributed to sharpen the picture, and have allowed to clarify some of the targets of 5-HT during development. These studies also demonstrated the functional consequences of disrupting normal 5-HT metabolism *in vivo* during critical periods of development, both as concerns the wiring of the brain and the behaviour of adult animals.

One important concept that emerged from these studies is that 5-HT levels are tightly controlled, during development, by 3 major genes of the clearance pathway of 5-HT : the serotonin transporter (SERT), the vesicular monoamine transporter (VMAT2) and the monoamine oxidase A (MAOA). All 3 genes are broadly expressed at early time points in embryonic life and during late postnatal life. These genes are expressed in a wide population of non-monoaminergic neurones, particularly in the major sensory pathways. This transient developmental expression is shut down fairly sharply as the neurones mature in a cell autonomous manner. Disrupting the function of any of these 3 genes alters the level of brain 5-HT, either augmenting it, in the MAOA and SERT-knockout mice or decreasing it, in the VMAT2 knock outs. The consequences of these genetic changes on brain development are generally subtle : the basic structure of the brain is not modified but a number of alterations found in the late process of maturation of the brain, such as the refinement of are topographic axonal maps, the trophic-dependent cell survival, and the dendritic differenciation of several neuronal populations. These changes are important for normal adult brain function, since a number of behavioural changes are observed in these mouse strains such as altered aggressive behaviour, learning deficits, vulnerability to drug intake, or increased anxiety-related behaviour.

We will discuss in greater detail the effects of 5-HT excess or lack of 5-HT in the maturation of the barrel cortex in the primary somatosensory area and in the cingulate cortex. Different 5-HT receptor subtypes appear to be implicated in the maturation process of the various cellular components of this cortical circuitry - the 5-HT1B receptor are important for the maturation of the thalamocortical neurones and the 5-HT2 receptors in the maturation of the cortico-cortical neurones. This shows that only a thorough genetic molecular analysis will allow to completely decipher and tease out the once global "developmental effects" of 5-HT-id

Antipsychotic treatment effects on the clinical and pathophysiological course of brain pathomorphology in schizophrenia

Jeffrey A. Lieberman

University of North Carolina School of Medicine

Evidence suggests that morphologic abnormalities reflecting cellular and circuit based disturbances which underlie major mental illnesses like schizophrenia are either inborn and developmental,or progressive and possibly degenerative in nature or both. Despite the substantial evidence and intellectual appeal of the "Neurodevelopmental Theory" of schizophrenia, the progressive and deteriorating course of the illness is undeniable. Moreover, an increasing number of methodologically rigorous longitudinal neuroimaging studies which examined patients prospectively from their early stages of illness have demonstrated that regionally specific morphologic changes occur over, and in relation to, the course of their disorder. Among the questions that these findings have raised is what are the effects of treatment on brain morphology and in the progressive changes that occur.

Preclinical studies in rodents and subhuman primates have reported psychotropic drug induced neuroplastic alterations occurring in cell function and ultrastructural development that can produce changes that are morphometrically detectable. Human studies have to an extent replicated these findings. One problem in studying treatment effects on brain structure and function in animals is that they do not simulate the potential interaction between pharmacologic effects and disease pathophysiology. Even when animal models are used one can't be certain that the model (be it pharmacolgic, lesion-based or targeted gene modification) fully simulates in-vivo disease pathophysiology.

This presentation will present findings on treatment effects on brain morphology and examine them in terms of whether they are drug effects on normative function and structure (iatrogenic) or on disease pathophysiology (therapeutic), and then consider the mechanisms by which these occur (i.e. palliative, neuroprotective, neurogenetic).

NAA levels in the dorsolateral prefrontal region in the first years of schizophrenia are inversely related to disease duration

Vicente Molina(1); Javier Sánchez (2); Santiago Reig(2); Javier Sanz(1); Carlos Benito(3); Javier Pascau(2); Fernando Sarramea(4); Juan D. Gispert(2); José M. Misiego(1); Tomás Palomo(1); Manuel Desco(2)

(1) Dept. of Psychiatry, Hospital Doce de Octubre, Madrid, Spain
 (2) Dept. of Experimental Medicine, Hospital "Gregorio Marañón", Madrid, Spain
 (3) Dept. of Neuroradiology, Hospital "Gregorio Marañón", Madrid, Spain
 (4) Dept. of Psychiatry, Hospital Reina Sofia, Córdoba, Spain

Background: Magnetic resonance spectroscopy studies of schizophrenia have revealed consistently diminished N-acetyl aspartate (NAA) levels in chronic schizophrenic patients, but not in cases of recent onset. Studies on the relationship between this marker and disease duration have generally been negative, although it is also true that they have been performed on patients with long-standing illness.

Method: We have compared NAA levels in the dorsolateral prefrontal area in 16 recent-onset patients (RO; duration, 1.8 ± 0.6 years), 19 chronic (CR; duration, 9.7 ± 6.1 years), and 20 healthy controls (CO). We have studied the NAA/ creatine and choline/ creatine ratios in the dorsolateral prefrontal region in both hemispheres, controlling for the effect of age.

Results: The chronic patients had significantly lower NAA/ Cr ratios in the left hemisphere compared to the recent-onset patients and healthy controls, with no difference in the Cho/ Cr ratio. There were no differences observed between controls and RO. There was a significant, inverse relationship between left side NAA/ Cr and disease duration.

Conclusions: These data support the hypothesis that prefrontal NAA levels may progressively decrease in schizophrenia. Taken within the context of the existing literature, these results indicate that this process may be limited to the first few years following onset of illness. Finding diminished prefrontal levels of NAA in schizophrenia may therefore be limited to chronic patients.

Key words: N-acetyl aspartate, schizophrenia, prefrontal, MRS, neurodegeneration, duration

Session 3: Synaptic function: Understanding the pathophysiology of schizophrenia Chair: Patricia Gaspar

New approaches to synaptic vesicle exocytosis and endocytosis

Alvarez de Toledo, G. & Tabares L.

Department of Physiology & Biophysics. University of Seville. Spain

Kiss-and-run exocytosis is a rapid and efficient method of releasing secretory products to the extracellular medium through the transient formation of a fusion pore. During kiss and run, the vesicle membrane does not collapse into the plasma membrane, maintaining the secretory vesicle its position, identity and potentiality to perform a second round of exocytosis. In a previous report, we found that the incidence of kiss-and-run increases as the extracellular calcium concentration rises in the mM range (Ales et al. 1999, Tabares et al. 2001). These kiss-and-run events lasted only for milliseconds (average of 50 ms, called "Fast kiss-and-run") and released the same amount of transmitter than a conventional exocytosis. However, we did not demonstrate whether calcium mediated this effect extracellularly or by an indirect cytosolic calcium increase. To answer this question, we have done whole cell membrane capacitance measurements in peritoneal rat mast cells in which we could experimentally control both the extracellular and the intracellular calcium concentrations. This preparation also allows resolving individual fusion events. To monitor release, we have done simultaneous measurements of serotonin release with amperometry by positioning a carbon fiber electrode over the cell surface.

We analyzed well-defined single exocytotic fusion events from capacitance and amperometric measurements. The incidence of kiss-and-run increased when the rate of degranulation was decreased either by lowering the amount of secretagogue (GTP γ S) or by dialyzing the cells with lower calcium concentrations. At similar amounts of GTP γ S, the number of kiss-and-run events decreases as the cytosolic calcium concentration was raised from 100 nM to 1 μ M. Simultaneous amperometric measurements revealed that these kiss-and-run events were slow (lasting on average 500 ms), and release only a small fraction of the vesicle contents. By contrast, when the extracellular calcium concentration was raised from 2 mM up to 90 mM, the incidence of kiss-and-run increased and the amount of serotonin was similar to the quantal contents of a rat mast cell granule, indicating the occurrence of fast kiss-and-run events.

These data reveal that fast kiss-and-run events are evoked by calcium acting from the extracellular medium and that these events represent a different kinetic process from the slow kiss-and-run events. We propose a kinetic model of exocytosis that account for slow and fast kiss-and-run exocytosis.

References:

Ales, E., Tabares, L., Poyato, J., Valero, Lindau, M & Alvarez de Toledo G. High calcium concentrations shift the mode of exocytosis to the kiss-and run mechanism. *Nat Cell Biol* 1, 40-4. (1999).

Tabares, L., Ales, E, Lindau, M. & Alvarez de Toledo, G. 2001. Exocytosis of Catecholamine (CA)-containing and CA-free granules in chromaffin cells. J. Biol. Chem. 276, 39974-39979

Glutamate signalling and synaptic transmission

Juan Lerma

Instituto Cajal, CSIC. Av. Doctor Arce 37, 28002-Madrid

Alterations of glutamatergic neurotransmission have been related to the neuronal damage observed after episodes of ischemia and hypoglycemia, as well as to the etiology of a series of neurological conditions including epilepsy, Alzheimer's disease, Huntington's chorea and amyotrophic lateral sclerosis. The cloning of a large number of glutamate receptor proteins and the discovery of their structural relationships have paved the way to most of our current understanding of the biophysical properties and the physiological role of each subtype in the mammalian brain. Of the glutamate receptor subtypes, NMDA, AMPA and kainate receptors, the latter is by far the less well understood, despite of their wide distribution in the brain.

Whilst the physiological role of these receptors is still not absolutely clear, recent advances have highlighted different aspects of their behavior (see ref 2 for a review). Progress in this field has been hampered by the lack of specific pharmacological tools. However, the discovery of a specific AMPA receptor antagonist, GYKI53655 (3,5), has made functional studies feasible. As a result, it has been proposed that kainate receptors play a role in both synaptic transmission and synaptic plasticity. In addition, it has been shown that the activation of kainate receptors modulates neurotransmitter release from a number of synapses. Indeed, in keeping with their presumptive role in epilepsy (e.g. ref. 1), kainate has been found to depress inhibitory GABA transmission in the rat hippocampus.

Kainate receptors share the membrane topology of ionotropic glutamate receptors (i.e. AMPA and NMDA receptors), they contain 3 transmembrane domains as well as a membrane domain that contributes to the ion channel wall. As a result, it was somewhat surprising to find, that inhibition of GABA release by kainate receptors in the hippocampus involved a second messenger cascade mediated by a Pertussis toxin sensitive G-protein and activation of Protein Kinase C (i.e. a rather classical metabotropic cascade; ref. 4). Since this discovery, further examples of such metabotropic activity have been reported, as well as other mechanisms through which kainate can act. However, in none of these cases has the receptor subunit involved in this activity been specified.

Hypothetically, the ionotropic and metabotropic activities of kainate receptors could be mediated through the same receptor complex. Alternatively, since the membrane topology of kainate receptors is in principle incongruous with the structure of G protein-coupled receptors, they may involve distinct or unrelated receptors. We have explored these possibilities in a simple cell model, that of cultured dorsal root ganglion (DRG) neurons. These neurons express a rather homogeneous population of kainate receptors, predominantly containing GluR5 subunits. In a population of DRG cells, we found that kainate was able to elicit a G-protein dependent increase in [Ca²⁺]_{int.} By using confocal microscopy and after application of kainate in the absence of extracellular Ca²⁺, the levels of [Ca²⁺]_{int} were seen to increase in localized areas of the cell body, as well as in discrete spots along the neurites and close to bifurcation points. Measurements from these points indicated that the larger increases in [Ca²⁺]_{int} were in the neurites rather than in the soma. Furthermore, a brief exposure to kainate inhibited voltage-dependent Ca²⁺-channels, a process that was sensitive to inhibitors of G-proteins and Protein Kinase C (PKC). This metabotropic effect of kainate does not seem to require ion channel receptor activity. However, it was not observed in neurons prepared from mice deficient for the ion channel forming subunit GluR5. Therefore, our results not only provide evidence for the existence of a metabotropic kainate receptor that inhibits Ca²⁺ channels, but also indicate that an ion channel forming subunit is involved independently in both ionotropic and metabotropic activities. Given that the nature of the signal transduction pathway triggered by the receptor determines the cellular response, our results indicate that kainate receptor activation may serve a number of novel functions.

References:

- Ben-Ari, Y. (1985). Limbic seizure and brain damage produced by kainic acid: mechanisms and relevance to human temporal lobe epilepsy. Neuroscience 14, 375-403.
- Lerma, J., Paternain, A.V., Rodriguez-Moreno, A., and Lopez-Garcia, J.C. (2001). Molecular physiology of kainate receptors. Physiol. Rev. 81, 971-98.
- Paternain, A.V., Morales, M., and Lerma, J. (1995). Selective antagonism of AMPA receptors unmasks kainate receptor-mediated responses in hippocampal neurons. Neuron 14, 185-9.
- Rodriguez-Moreno, A., and Lerma, J. (1998). Kainate receptor modulation of GABA release involves a metabotropic function. Neuron 20, 1211-8
- Wilding, T.J., and Huettner, J.E. (1995) Differential antagonism of alpha-amino-3-hydroxy-5-methyl-4isoxazolepropionic acid-preferring and kainate-preferring receptors by 2,3-benzodiazepines. Mol Pharmacol 47, 582-587.

Basic developmental rules for the construction of cortical networks

Yehezkel Ben-Ari

Institut de Neurobiologie de la Méditerranée (INMED). INSERM U29 Parc Scientifique de Luminy. 13273 Marseille Cedex 09 Tel.: 04 91 82 81 24- Fax: 04 91 82 81 01

In the course of studies initiated in the late nineties, we have shown three basic developmental rules for the maturation of synaptic activity in a cortical circuit:

I) GABAergic synapses are initially excitatory because of a higher intracellular concentration of chloride in immature neurons. Consequently, the activation of GABAergic receptors and synapses lead to the generation of sodium and calcium potentials and an increase of (Ca)I by the removal of the voltage dependent Mg++ block from NMDA channels.

II) GABAergic synapses are formed before glutamatergic ones in both principal cells and interneurons and this has a morphological substrate: neurons are silent (no synapses) when they have no dendrites, they have only GABAergic synapses when they have a small apical dendrite (less than 50 uM) and they have also glutamate synapses when they have a large dendritic arbor that reaches the superficial layer. This sequence is observed earlier in interneurones (in utero in rodents at a time when pyramidal neurons are all silent); therefore, GABAergic interneurons that divide and arborize prior to the principal cells are the source and the target of the first synapses formed in the circuit and GABA is the principal excitatory agent at a time when glutamatergic synapses are absent.

III) There is in the developing network a unique pattern of network driven activity- generated by the excitatory actions of GABA in association with developing glutamatergic synapsesthat provides most of the synaptic activity at an early developmental stage. We have now shown this dominating rhytm in vivo (patch recordings in the neonatal rat in situ) and in our intact hippocampi in vitro in which it is possible to study the propagation of these activities within the hippocampus and between hippocampi.

These rules appear to have been kept throughout evolution, as excit atory actions of GABA in immature neurons have been described in a large number of species and brain structures. We have suggested elsewhere (Ben-Ari, 2002) that this sequence enables to solve a dilemma that developing structures have, namely how to construct a structure without having a potentially toxic imbalance between excitation and inhibition. It is suggested that these patterns play an important role in the strategy followed by the brain to shift from a silent structure with no electrical activity and no synapses to an active one that possesses a highly diversified range of electrical signals and billion of selective synapses. Furthermore, the earliest formed GABAergic synapses are liable to alterations by the Hebbian principles and activity can also shift the type of neurons from one to the other (silent to GABA only for instance). We have recently extended this model to the primate by making the first recording of central neurons in utero in primates. Thus, these rules apply also to non-human primates and most likely to humans as well (Khazipov et al, 2001).

In the presentation, I shall also briefly review data on the role of activity in the formation of the first synapses and report that prior to synapses formation, very immature neurons have already a paracrine mode of communication by means of a non-vesicular non Calcium and non SNARE dependent release of GABA and glutamate that generates in target neurons, a slow current present only at early developmental stages. It is suggested that all these early mechanisms interfere in certain conditions with the normal construction of the brain leading to permanent deleterious conditions. This is exemplified by migration disorders that can lead to the formation of heterotopias and other disorders.

References:

BEN-ARI Y., CHERUBINI E., CORRADETTI R., GAIARSA J.L. Giant synaptic potentials in immature rat CA3 hippocampal neurones Journal of Physiology, London, 1989, 416, 303-325

CHERUBINI E., GAIARSA J.-L., BEN-ARI Y. GABA: an excitatory transmitter in early postnatal life. Trends in Neuroscience, 1991, 14,, 515-519

LEINEKUGEL X, TSEEB V, BEN-ARI Y, BREGESTOVSKI P: Synaptic GABAA activation induces Ca2+ rise in pyramidal cells and interneurons from neonatal hippocampal slices. Journal of Physiology, 1995, 487(2):319-329.

MCLEAN H, CAILLARD O, BEN-ARI Y, GAIARSA J: Bidirectional plasticity expressed by GABAergic synapses in neonatal rat hippocampus. Journal of Physiology (London), 1996,496(2):471-477

BEN-ARI, Y., KHAZIPOV, R.; CAILLARD.O., LEINEKUGEL, X.; GAIARSA, J. L. GABAA, NMDA and AMPA receptors: A developmentally regulated "ménage à trois". Trends In Neurosciences : 20 (11) : 499-547, 1997

KHALILOV J., ESCLAPEZ, M., KHAZIPOV, R., AGGOUN, D., LAMSA, K., LEINEKUGEL, X., MEDINA, I., BEN-ARI, Y. A novel In Vitro Preparation: The Intact Hippocampal Formation. Neuron: Vol. 19, 743-749, 1997.

LEINEKUGEL, X., MEDINA, I., KHALILOV, I., BEN-ARI, Y., KHAZIPOV, R. Ca2+ oscillations mediated by the synergistic excitatory actions of GABAA and NMDA receptors in the neonatal hippocampus. Neuron 18:243-255, 1997.

KHAZIPOV, R., LEINEKUGEL, X., KHALILOV, I., GAIARSA, J. L., BEN-ARI, Y. Synchronization of GABAergic Interneuron network in CA3 subfield of neonatal rat hippocampal slice. Journal of Physiology (Lond.) 498:763-772, 1997

LEINEKUGEL X, KHALILOV I, BEN-ARI Y AND KHAZIPOV R. Giant depolarizing potentials : the septal pole of the hippocampus paces the activity of the developing intact septo- hippocampal complex in vitro. Journal of Neuroscience, 18(16), 6349-6357,1998.

CAILLARD O, BEN-ARI Y, GAIARSA JL. Mechanisms of induction and expression of long-term depression at GABAergic synapses in the neonatal rat hippocampus. J Neuroscience., 1999, 19(17):7568-77. <u>KHAZIPOV R, DESFRERES L, KHALILOV I, BEN-ARI Y</u>. A three-independent-compartment chamber to study in vitro commissural synapses Journal of Neurophysiology, 1999, 81(2):921-

TYZIO R, REPRESA A, JORQUERA I, BEN-ARI Y, GOZLAN H, ANIKSZTEJN L. The establishment of GABAergic and glutamatergic synapses on CA1 pyramidal neurons is sequential and correlates with the development of the apical dendrite. J Neuroscience., 1999, 19(23):10372-82

BEN-ARI Y, Developing networks play the same melody. TINS. 2001, 24 (6); pp353-360. Khazipov et al, Early development of neuronal activity in the primate hippocampus in utero. J Neuroscience, 21(9770-9781)

Ben-Ari Y. Excitatory actions of GABA: the nature of the nuture. Nature Reviews Neuroscience. 2002(vol3). 728-739

Leinekugel et al., Correlated bursts of activity in the neonatal rat hippocampus in vivo. Science, 296-(2049-2052).

Demarque et al. Paracrine mode of communication between neuron prior to synapses formation. Neuron (in press) 2002

Schizophrenia as a disease of the synapse: Gene microarray studies

Károly Mirnics

Depts. of Psychiatry and Neurobiology, U. of Pittsburgh

DNA microarray technologies have enabled researchers to analyze complex gene expression changes for tens of thousands of genes. This method is ideally suited for studying complex brain disorders like schizophrenia, where interactions of genetic and environmental factors hold the key to the disease. The revealed transcriptome changes often define expression signatures disease states, identifying patterns that suggest previously unknown molecular interactions.

Simultaneous assessment of the transcriptome for the majority of the human genome will lead to the individualization of diagnosis and will eventually link molecular phenotypes to the precise clinical profile of the disorder. In our recent DNA microarray studies we uncovered novel and consistent gene expression patterns in the prefrontal cortex of individuals with schizophrenia when compared to matched controls. We observed robust changes in the expression of genes related to presynaptic secretory release. Transcripts for Synapsin 2, N-ethylmaleimide-sensitive factor, SNAP, vesicular proton pumps and 14-3-3 levels were decreased in the majority of comparisons, albeit in a subject-specific manner. Furthermore, we also uncovered consistent transcripts related to GABAergic and glutamatergic transmission, which may be secondary to altered presynaptic drive. In addition, subjects with schizophrenia also reported consistent transcript decreases of multiple genes participating in various energy metabolism pathways and ubiquination, which may be also related to the presynaptic gene expression alterations.

Importantly, the presynaptic transcriptome alterations observed in schizophrenia showed a subject-specific pattern. That is, although all subjects showed decreased expression of multiple presynaptic gene products, the specific combination of affected transcripts varied from subject to subject. This finding strongly argues for a continuity of molecular phenotypes in schizophrenia that is likely to be related to the variability in the clinical manifestations of the disease.

The same microarray data set also revealed multiple changes that occur at the postsynaptic cell. These, amongst others, expression of AMPA2, regulator of G-protein signaling 4 (RGS4) and multiple genes involved in GPCR signaling were altered. Our microarray findings also seem to implicate multiple genes involved in both phosphatidylinositol and cAMP signaling pathways, which are likely consequences of more specific upstream changes in GPCR signaling, possibly involving dopamine signaling alterations.

In contrast, in a study of chronic haloperidol-treated monkeys we did not detect transcript alterations for the same presynaptic and postsynaptic genes. This suggests that the uncovered expression changes in schizophrenia may not be result of chronic antipsychotic treatment.

Undoubtedly, microarray studies and other complex transcriptome analysis methods will guide future attempts for deciphering complex molecular phenotypes associated with mental disorders. Defining disease-specific transcriptome changes will also facilitate development of animal models that closely mimic common molecular events associated with psychiatric conditions and holds a great promise for individualization of diagnosis and treatment.

References:

Mirnics, K. et al. (2000) Molecular characterization of schizophrenia viewed by microarray analysis of gene expression in prefrontal cortex. *Neuron* 28 (1), 53-67

Mirnics, K. and Lewis, D.A. (2001) Genes and subtypes of schizophrenia. Trends Mol Med 7 (7), 281-283

Mirnics, K. et al. (2001) Analysis of complex brain disorders with gene expression microarrays: schizophrenia as a disease of the synapse. *Trends Neurosci* 24 (8), 479-486

Mirnics, K. et al. (2001) Disease-specific changes in regulator of G-protein signaling 4 (RGS4) expression in schizophrenia. *Mol Psychiatry* 6 (3), 293-301

Lewis, D.A. and Levitt, P. (2002) Schizophrenia as a disorder of neurodevelopment. Annu Rev Neurosci 25, 409-432

Middleton, F.A. et al. (2002) Gene expression profiling reveals alterations of specific metabolic pathways in schizophrenia. J Neurosci 22 (7), 2718-2729

Pongrac, J. et al. (2002) Gene expression profiling with DNA microarrays: advancing our understanding of psychiatric disorders. *Neurochem Res* 27 (10), 1049-1063

Molecular profiles of anatomically and electrophysiologycally characterized neocortical neurons

Toledo-Rodriguez M., Blumenfeld B., Wu C.Z. & Markram, H

Multiple neurobilogical pathologies are result of neuronal malfunction. The understanding of these pathologies is based on the full knowledge of neuronal behavior in non -pathological conditions. Neurons influence the activity of other neurons and the microcircuit in general via neuromodulators. Up till now studies investigating neuromodulator expression and function have been limited due to methodological constrains. In order to study this diversity, we have undertaken a large-scale multiplex single cell RT-PCR study in which we examined the expression of 3 calcium-binding proteins, 10 neuropeptides, 5 synthesizing enzymes in anatomically, physiologically and molecularly characterized neurons at the single cell level. The study of more than 300 neurons using this technique lead us to the conclusion that the biochemical coexpression profiles are more complex that though till now, but nevertheless they follow three rules.

* SELECTIVITY->There is selectivity in the coexpression patterns, as only few of all the possible gene expression combinations are found.

* SPECIFICITY->Each co-expression pattern is specific for one or few morphological and electrophysiological groups.

* HOMOGENEITY->In some exceptional cases one neuropeptide is expressed homogeneously by all the members of a single anatomical group, as in the case of MCs that all express Somatostatin or SBCs that all express VIP.

Session 4: Development and plasticity: Role in the pathogenesis of schizophrenia Chair: Pat Levitt

Synaptogenesis in the mammalian neocortex

J. P. Bourgeois

Laboratoire «Récepteurs et Cognition». Département des Neurosciences. Institut Pasteur. 75724. Paris Cedex 15. France

After neurogenesis and hodogenesis, synaptogenesis represents a crucial final step in cortical development. The cerebral cortex of the rhesus monkey provides an excellent animal model for the study of the development and plasticity of the human cerebral cortex. Description of the kinetics of synaptogenesis in several cortical areas of the macaque monkey from conception to death reveals several properties:

1) Onset of synaptogenesis is a very early developmental event, occuring in the first half of gestation, i.e. long before interactions with the environment.

2) The kinetics of synaptogenesis is complex rather than linear. Five distinct phases were identified in the whole cortical mantle of the macaque monkey. They were also described in rodent (deFelipe et al., Cerebral Cortex, 1997; 7: 619-634) and human brains, suggesting a highly conserved process through neocortical evolution. For the human cerebral cortex, due to sampling difficulties, the data are scarce and diverse (Larroche, Anat. Embryol., 1981; 162; 301-312; Huttenlocher and Dabholkar, J. Comp. Neurol. 1997; 387: 167-178; Zecevic, Cerebral Cortex 1998; 8: 245-252; Chugani, Neuroscientist 1999; 5: 29-40). We can tentatively propose an approximate timeline for each of these phases of synaptogenesis. Phase 1 (slow synaptogenesis in cortical preplate) would start near the 7th week of gestation. Phase 2 (slow synaptogenesis in cortical plate) would end near 17-22 weeks. Phase 3 (very rapid synaptogenesis in cortical plate) would start near the 26th week of gestation and end by 8-12 postnatal months in the primary visual cortex, and 2-3 years in the prefrontal cortex. Phase 4 («en plateau» characterized by very high densities of synapses) lasts for 2-3 years in the visual cortex and a whole decade in the prefrontal cortex, until puberty. In phase 5, following puberty, the mean density of synapses drops to the density observed at birth. This density is maintained troughout adulthood, and then decreases significantly during senescence, until death.

3) Synaptogenesis comes into successive waves for diverse classes of synapses, progressively building the synaptoarchitectonic organization of the cerebral cortex. The developmental pattern of this organization is similar in the whole cortical mantle of the macaque and begins in the second half of gestation. Many anatomical (synaptoarchitectony, ocular dominance columns in primary visual cortex, columns in prefrontal cortex, etc..), physiological, and functional properties of these neocortical circuits are apparent prenatally and perinatally, and are thought to depend on innate mechanisms.

4) The early phases 1,2, and onset of phase 3 of synaptogenesis, are very robust developmental events. They take place even in challenging experimental (preterms, early bilateral enucleation, null mutations...) or pathological (double cortex, schizophrenia...) conditions.

5) Paradoxically, recent results suggest that «milder» affective perturbations such as maternal care (Liu D et al. Nature Neuroscience, 2000; $\underline{3}(8)$: 799-806), or maternal deprivation (Helmeke C. et al. Cerebral Cortex, 2001; 11: 717-727) may cause discrete modifications of the synaptoarchitectony in specific cortical territories.

The early phases 1,2, and onset of phase 3 of synaptogenesis, appear controlled by genetic and epigenetic mechanisms intrinsic to the cerebral cortex. Later on, at the end of phase 3, and during phase 4 "en plateau", these intrinsic mechanisms are also modulated by the environment. This period of synaptic plasticity allows the fine tuning of the neurono-synaptic circuits involved in the progressive and coordinated maturations of all the sensory, motor, and associational cortical functions, until puberty.

References:

Bourgeois J.P. Synaptogenesis in the neocortex of the newborn: the ultimate frontier for individuation? In: The Newborn Brain: Neuroscience and clinical applications. H. Lagercrantz, M. Hanson, P. Evrard, C. Rodeck, Editors. Cambridge University Press. pp 91-113, 2002.

Determinants of pre- and postsynaptic interactions during development

Edward S. Ruthazer, Colin J. Akerman, and Hollis T. Cline

Cold Spring Harbor Laboratory Cold Spring Harbor, NY 11724

Regulation of the development of neuronal structures, such as axons and dendrites, affect the function of brain circuits. It is important to understand the mechanisms controlling the development of neuronal circuits in order to assess how these mechanisms are affected under pathological conditions. We used in vivo time-lapse imaging to determine the role of visual stimulation and glutamatergic synaptic transmission on the development of presynaptic retinal axon arbor structure in the visual system of Xenopus tadpoles. We imaged labeled retinal axons in Xenopus tadpoles in which surgery was used to induce binocular innervation of the optic tectum. Under these conditions, retinal inputs from the two eyes were initially overlapping and gradually segregated into eye-specific zones over time. This system permitted us to correlate structural dynamics of the arbor axon branches with degree of local innervation from axons from either the same eye or the opposite eye and thereby test whether patterned activity instructs axon arbor development. Axons eliminated branches preferentially from territory dominated by the eye opposite to their eye of origin, but added branches with approximately equal probability in both eye territories. NMDA receptor blockade prevented the selective stabilization of branches. In addition, the rate of branch additions was increased in sparsely innervated territory regardless of the eye of origin. These results 1) suggest that an activity-independent mechanism promotes branch additions in sparsely innervated brain regions, and 2) provide evidence for a correlation-based mechanism for the fine-tuning of axon arbor morphology in which the locations of branch stabilizations are governed by postsynaptic NMDAR-dependent signaling.

Imaging the structure and function of dendritic spines

R. Yuste

Dept. Biological Sciences, Columbia University, New York, NY 10027

Dendritic spines are sites of most excitatory contacts in the cortex and have been practically inaccessible to functional measurements until the introduction of two-photon microscopy (1). Using a custom-built two-photon microscope (2) we have studied two questions:

A- Calcium compartmentalization of spines: Mechanisms of calcium influx and decay kinetics.

We have shown that dendritic spines can compartmentalize calcium (1) and explored the mechanisms of calcium influx into spines (3). Decay kinetics are controlled by (i) active calcium extrusion and (ii) diffusion of calcium from spine to dendrite (4). Although extrusion is likely to dominate in physiological conditions, the contribution of calcium pumps and diffusion varies from spine to spine, depending on the location of the spine along the dendrite, producing different sensitivity to synaptic depression (5). Decay kinetics change during spine motility (6).

B- Rapid motility of spines: Developmental regulation, mechanisms and possible functions.

We have characterized spine motility in acute and cultured slices (7). In all cells, dendritic protrusions (filopodia and spines) are highly dynamic, exhibiting a diversity of morphological rearrangements over short (<1 min) times (8). Spine motility declines during postnatal maturation, but changes are apparent in most spines in late postnatal neurons. Spine motility could be related to synaptogenesis. These morphological reorganizations are actin-based and can be regulated by Rho family GTPases (9). Finally, spines can move even with synaptic contacts (10).

References:

- 1. Yuste and Denk (1995). Nature 375:682.
- 2. Majewska et al. (2000). Pflügers Archiv 441:398.
- 3. Yuste et al.(1999), J. Neurosci. 19:1976.
- 4. Majewska et al. (2000). J. Neurosci. 20:1722.
- 5. Holthoff et al (2002). Neuron 33, 425-437
- 6. Majewska et al. (2000). J. Neurosci. 20: 8262.
- 7. Dunaevsky et al. (1999). PNAS 96:13438..
- 8. Parnass et al.(2000). Hippocampus 10:561.
- 9. Tashiro et al. (2000). Cerebral Cortex 10:927
- 10. Dunaevsky et al. (2001). Nature Neurosci 4:685.

Hippocampal structure and function in schizophrenia

Stephan Heckers

Department of Psychiatry, Massachusetts General Hospital, Harvard Medical School Boston, MA 02115

Subtle yet significant volume reduction of the hippocampus is one of the most replicated brain abnormalities in schizophrenia. Neuroimaging studies have shown that smaller hippocampal volume predicts the onset of psychosis in high-risk subjects and is fully developed at the first presentation of psychotic symptoms, but may progress throughout the disease process. Postmortem studies have demonstrated that total hippocampal neuron number is not different in schizophrenia, but that subsets of neurons are functionally abnormal. Etiology and mechanism of the volumetric and cellular abnormality of the hippocampus in schizophrenia remain unclear, but appear to involve a disturbance of synaptic organization.

Recent functional neuroimaging experiments have supported the hypothesis that these structural abnormalities lead to functional deficits of the hippocampus in schizophrenia. Hippocampal recruitment during conscious recollection is impaired in schizophrenia, but simple recognition remains intact, even in the context of significant hippocampal volume reduction. Taken together, studies of hippocampal structure point to selective abnormalities of hippocampal architecture, resulting in impairments of some but not all hippocampal functions in schizophrenia.

Session 5: Models systems to investigate the underlying of etiology of schizophrenia Chair: Guillermo Álvarez de Toledo

Molecular determinants of neurotransmitter release

Thomas C. Südhof

Center for Basic Neuroscience, Dpt. of Molecular Genetics, and Howard Hughes Medical Institute, UT Southwestern Medical Center, Dallas, TX 75390; Phone: 214-648 1876; Fax: 214-648 1879; e-mail: Thomas.Sudhof@UTSouthwestern.edu

Our laboratory is interested in understanding how a presynaptic terminal contacts a postsynaptic neuron, and how the terminal secretes neurotransmitters in a tightly regulated and topologically restricted manner. Neurotransmitter release occurs by synaptic vesicle exocytosis that is triggered by influx of Ca^{2+} into the presynaptic neuron. Ca^{2+} -evoked synaptic vesicle exocytosis is one of the fastest and most tightly controlled reactions in biology, and can be modulated during synaptic plasticity. The speed and plasticity of neurotransmitter release are one of the major factors that shape the the exquisite speed and precision with which synaptic networks function.

In a longstanding program, we are investigating the molecular cascade that orchestrates neurotransmitter release. We are trying, in conjunction with other laboratories, to achieve a systematic description of all of the molecular components that mediate neurotransmitter release, and to analyse these components structurally and functionally. Although far from complete, these studies have provided an initial description of the molecular machinery that mediates neurotransmitter release, and provide clues to how the speed, precision, and plasticity of a synapse are achieved. For example, candidate Ca^{2+} -sensors for exocytosis have been identified in synaptotagmins, and integrators of presynaptic signalling at the active zone were described in RIM and Munc13 proteins. In my contribution I will discuss the current state of the project and its implications for a wider understanding of brain function.

References:

Südhof, T.C. (1995) The Synaptic Vesicle Cycle: A cascade of protein-protein interactions. <u>Nature</u> 375, 645-653.
 Verhage, M., Maia, A.S., Plomp, J.J., Brussaard, A.B., Heeroma, J.H., Vermeer, H., Toonen, R.F., Hammer, R.E., van den Berg, T.K., Missler, M., Geuze, H.J., Südhof, T.C. (2000) Synaptic assembly of the brain in the absence of neurotransmitter secretion. <u>Science</u> 287, 864-869.

3. Fernandez-Chacon, R., Konigstorffer, Gerber, S.H., Garcia, J., Matos, M.F., Stevens, C.F., Brose, N., Rizo, J., Rosenmund, C. and Südhof, T.C. (2001) Synaptotagmin I functions as a Ca²⁺-regulator of release probability. <u>Nature</u> 410, 41-49.

4. Schoch, S., Castillo, P.E., Jo, T., Mukherjee, K., Geppert, M., Wang, Y., Schmitz, F., Malenka, R.C., and Südhof, T.C. (2002) RIM1α forms a protein scaffold for regulating neurotransmitter release at the active zone. Nature 415, 321-326.

5. Castillo, P.E., Schoch, S., Schmitz, F., Südhof, T.C., and Malenka, R.C. (2002) RIM1 α is required for presynaptic long-term potentiation. <u>Nature</u> 415, 327-330.

6. Biederer, T., Sara, Y., Mozhayeva, M., Atasoy, D., Liu, X., Kavalali, E.T., and Südhof, T.C. (2002) SynCAM, A synaptic cell adhesion molecule that drives synapse assembly. <u>Science</u> 297, 1525-1531.

Increased GLT-1 expression in the prefrontal cortex of schizophrenics

F. Conti, C. Matute, M. Melone, A. Vallejo-Illarramendi and Fiorenzo Conti1, M.D.

High-affinity glutamate transporters (GluTs) are important regulators of glutamatergic transmission (Conti and Weinberg, 1999); increased Glu uptake may thus reduce glutamatergic transmission. The possibility that increased Glu uptake plays a role in the pathophysiology of schizophrenia is suggested by the demonstration that in rat frontal cortex clozapine reduces the expression and function of GLT-1 – the astrocytic GluT responsible for the greatest proportion of Glu transport - in a strong and selective manner, thereby raising Glu levels (Melone et al., 2001, 2002).

We thus studied the prefrontal cortex (PFC) of 2 drug-free schizophrenic subjects to verify whether the reduced efficacy of glutamatergic transmission reported in schizophrenia depends on the increased expression of GLT-1, the transporter responsible for most glutamate (Glu) transport.

Samples from areas 9/46 were used to study GLT-1 mRNA with DNA microarray and GLT-1 protein by immunocytochemistry and immunoblotting, and for measuring L-3H-Glu uptake. Gene expression and immunocytochemical studies showed that both GLT-1 mRNA and protein was robustly upregulated compared with controls. Functional studies revealed that Glu uptake was nearly 4-fold higher than in controls.

The hypothesis that GLT-1 is abnormally high in the PFC of schizophrenics is tenable and merits large-scale studies.

References:

⁻ Conti F, and Weinberg RJ (1999) Trends in Neurosciences, 22: 451-458.

⁻ Melone M, Vitellaro Zuccarello L, A Vallejo-Illarramendi, A Perez-Samartin, C Matute, A Cozzi, DE Pellegrini-Giampietro, JD Rothstein, Conti F (2001) Molecular Psychiatry, 6: 380-386.

⁻ Melone M, Bragina L, Conti F (2002) Molecular Psychiatry, in press.

A mouse model of mental illness based on a known risk factor

Limin Shi and Paul H. Patterson

Biology Division, California Institute of Technology, Pasadena CA 91125

There is considerable evidence that exposure of pregnant women to viral infection during a critical period can increase the incidence of schizophrenia and possibly autism in their offspring (reviewed in 1). We find that inducing an influenza virus respiratory infection in pregnant mice cause highly abnormal behavioral responses in their offspring, tested as adults (2). These tests include open field, novel object, social interaction and prepulse inhibition (PPI) of the acoustic startle response, and the results are consistent with abnormalities observed in subjects with schizophrenia and autism. Moreover, the deficit in PPI is corrected by anti-psychotic drug treatment, and exacerbated by psychomimetic drug treatment.

A variety of abnormalities have been observed in the cortex and hippocampus of mice born to infected mothers, including the distribution of SNAP-25 and reelin, and shrinkage of pyramidal neurons (3-5). Many of these findings are consistent with those reported for schizophrenic brain. We now find that these offspring also display a loss of Purkinje cells, specifically in lobules VI and VII of the cerebellar vermis (6). We also occasionally see what appear to be misplaced Purkinje cells in the white matter of lobule VII. In addition, the molecular layer of lobule VII appears to be thinner in these mice. This highly specific neuropathology is of particular interest because it coincides with that reported in both imaging and postmortem studies of autistic brains. This finding is also consistent with histological and behavioral abnormalities that have been linked to the cerebellum in schizophrenia.

We hypothesize that the abnormal brain development seen in the mice born to infected mothers is due to the maternal immune response, rather than to influenza infection of the fetal brain. In support of this idea, we find no evidence of viral RNA in the fetal brains of exposed mice. Moreover, a PPI deficit is found in adult mice born to mothers that were treated with poly(I:C), a synthetic, double stranded RNA that is known to evoke an anti-viral like immune response (2).

References:

6. Patterson, PH, Shi, L (2002) Internl Mtg Autism Res 2:S3.3.4.

Supported by Ginger and Ted Jenkins, the Mettler Autism Fund and the Stanley Medical Research Institute.

^{1.} Patterson, PH, Curr Opin Neurobiol 12:115-118, 2002.

^{2.} Shi, L, Fatemi, SH, Sidwell, RW, Patterson, PH, J Neurosci, in press.

^{3.} Fatemi, SH, Sidwell, RW, Kist, D, Akhter, P, Meltzer, HY, Bailey, K, Thuras, P, Sedgewick, J (1998) Brain Res 800:1-9.

^{4.} Fatemi, SH, Emamian, ES, Kist, D, Sidwell, RW, Nakajima, K, Akhter, P, Shie, A, Sheikh, S, Bailey, K, (1998) Molec Pyschiat 4:145-154.

^{5.} Fatemi, SH, Earle, J, Kmodia, R, Kist, D, Emamian, ES, Patterson, PH, Shi, L, Sidwell, R (2002) Cell Molec Neurobiol 22:25-33, 2002.

Determinants of developmental trajectory: Models of early insult leading to delayed or long-lasting psychopathology

P. Levitt^{1,2} G.D. Stanwood^{1,2} and K. Koshibu³

¹J.F. Kennedy Ctr for Research on Human Development & ²Dept Pharmacology, Vanderbilt Univ, Nashville, TN, USA; ³Dept Neurobiology, Univ Pittsburgh School of Medicine, Pittsburgh, PA, USA

A primary pathogenetic event of relatively short duration causing pre- and/or postnatal disruptions of brain development has been proposed to underlie schizophrenia. If this neurodevelopmental hypothesis is correct, then the resultant behavioral and cognitive signs and symptoms remain dormant for some time after the primary insult. This hypothesis of schizophrenia is attractive for a variety of reasons, and there are several converging lines of evidence to support such a model. However, few examples in which delayed-onset disruption of structure-function relationships have been documented. We will discuss two models, one genetic and one environmentally-induced, that result in different developmental trajectories and adaptive changes in brain structure and function.

In our investigations of a mouse (waved-1; wa-1) carrying a single gene mutation that causes gradual, postnatal hypomorphic expression of transforming growth factor-alpha (TGFa, peripubertal changes in brain structure and in learned fear conditioning occur, and are expressed with a gender bias¹. It is noteworthy that in the full TGF null mouse line in which expression of this growth factor is completely removed throughout development does not exhibit any functional or structural brain abnormalities. In contrast, we have used highresolution structural magnetic resonance imaging of wa-1 mice to document selective enlargement of ventricles in the absence of major loss of tissue parenchyma, changes that are reminiscent of schizophrenia. Moreover, alterations in learning appear to be at least partially due to adolescent onset of abnormal stress responsiveness in wa-1 male mice. We have assessed the levels of transcript encoding TGF α and EGFR, the receptor for TGF α , and have observed a gender dependent mRNA expression pattern in select HPA structures. In contrast, gender specificity was not observed in transcript expression patterns in learning associated brain regions such as the amygdala and hippocampus. This structure-dependent pattern of mRNA expression supports the importance of the HPA axis in assigning the gender specific phenotype in wa-1 that develops during adolescence.

In a second group of studies, we have used psychostimulant exposure to alter monoamine neurotransmitter function during prenatal brain development. In this model of gestational exposure to cocaine, administration of the drug for a very limited period of time during a "sensitive period", results in disrupted dopamine (DA) D_1 receptor signaling that remains permanently altered for the life of the animal^{2,3}. The disrupted signaling also causes abnormal development of certain neurons in regions of the cerebral cortex that receive dense DA innervation, including the prefrontal cortex and anterior cingulate. Strikingly, the offspring are permanently desensitized to the locomotor effects of psychostimulants, whereas adults exposed to the same level of cocaine instead exhibit intense sensitization, suggesting that the biological response of the CNS to this pharmacological insult varies not only in magnitude, but even in direction, depending on the developmental maturity of the nervous system when the insult is received.

Although the particular insults to the CNS used in these studies are not intended to model any specific causes of schizophrenia, these findings reflect the importance of timing on the disruption of neurodevelopment, and illustrate the vast diversity of resultant neuroadaptations. The study of such alterations in developmental trajectories are vital steps in understanding the complex additive and interacting effects that can lead to the eventual expression of the hallmark features of schizophrenia. Such developmental events can result in highly selective outcomes that reflect the culmination of adaptive (and sometimes maladaptive) changes that may underlie the pathophysiology of schizophrenia.⁴

Supported by: NIMH grant MH45507(PL) and NIDA grant DA11195 (PL and GS) and IGERT predoctoral grant (KK).

References:

- Burrows, R.C., Levitt, P., and Shors, T.J. Postnatal decrease in TGFα is associated with enlarged ventricles, deficient amygdaloid vasculature and performance deficts. Neuroscience <u>96</u>: 825-836, 2000.
- Jones, L.B., Stanwood, G.D., Reinoso, B.S., Washington, R.A., Wang, H.-Y., Friedman, E. and Levitt, P. In utero cocaine-induced dysfunction of dopamine D₁ receptor signaling and abnormal differentiation of cerebral cortical neurons. J. Neurosci. <u>20</u>: 4606-4614, 2000.
- Stanwood, G.D. and Levitt, P. The effects of cocaine on the developing nervous system. In: Handbook of Developmental Cognitive Neuroscience. C. Nelson and M. Luciana, (eds.), MIT Press, New York, <u>VII</u>: 519-536, 2001.
- Lewis, D.A. and Levitt, P. Schizophrenia as a disorder of neurodevelopment. Ann. Rev. Neurosci. 25:409-432.

POSTERS



Alantamine induces neurotransmitter release likely by blocking SK channels

Eva Alés, Francesca Gullo, Esperanza Arias, Antonio G. García and Manuela G. López

Galantamine has shown increased clinical efficacy and safety for the treatment of Alzheimer's disease. This drug has been described as a selective acetylcholinesterase inhibitor that allosterically modulates nicotinic receptors. However, here we present evidence of a new mechanism for galantamine, i.e. blockade of SK channels (Ca^{2+} -activated K⁺ channel).

These channels are activated by an increase of intracellular calcium and their activation causes membrane hyperpolarization, which inhibits cell firing and limits the firing frequency of repetitive action potentials.

This process termed spike-frequency adaptation is essential for normal neurotransmission.

We have observed that galantamine can inhibit SK channel currents in a concentration-dependent manner and, as apamin (a selective blocker of SK channels), can potentiate the secretory responses induced by brief acetylcholine and K^+ pulses (10 s) applied to fast-superfused bovine adrenal chromaffin cell populations. At the single cell level, galantamine caused a mild increase of the cytosolic calcium and catecholamine release, measured simultaneously using a fura-2 fluorescence dye and an amperometric carbon fiber microelectrode. This effect could not be reproduced in all tested cells. However, this action when observed, was clearly dependent of calcium and galantamine concentrations. Furthermore, a potentiation of exocytosis by galantamine could be observed when acetylcholine and potassium were applied.

Our hypothesis is that galantamine can block SK channels and inhibit the hyperpolarization induced by acetylcholine or depolarizing stimuli.

Therefore, galantamine could potentiate the rate of action potential firing and consequently, neurotransmitter release; this is a process that has been described to be depressed in Alzheimer's patients and other phycopathological diseases.

Catecholamine innervation of identified pyramidal neurons in the human temporal cortex

R. Benavides-Piccione and Javier DeFelipe. Cajal Institute, Madrid, Spain

Catecholamines have been shown to be involved in a variety of processes and pathologies such as Alzheimer's disease and schizophrenia. Previous studies have established that catecholaminergic innervation of both pyramidal and non-pyramidal cells occurs, and that this is selective for area, laminar and neuronal cell types. In the present study we have quantified catecholamine inputs to identified pyramidal cells in the human temporal cortex. To this purpose, we filled individual pyramidal neurons with Lucifer Yellow in fixed cortical tissue and immunostained with tyrosine hydroxylase (the rate limiting enzyme in catecholamine synthesis). Pyramidal cells (n=8) were reconstructed from serial confocal microscope images, and putative contacts between tyrosine hydroxylase immunoreactive (TH-ir) axon terminals and dendritic spines and shafts of labeled pyramidal cells were quantified at high magnification. After dividing the basal and apical dendritic fields of pyramidal cells in three portions (proximal, intermediate and distal), catecholamine contacts were located mainly in the intermediate portion. Moreover, pyramidal neurons received a remarkable narrow variation of 15 to 31 (average of 23) putative contacts from TH-ir axons per neuron. These results suggest that pyramidal neurons recieve an optimal number of cathecolaminergic synapses and that these synapses are stereotypically organized in the human temporal cortex.

Cortical AMPA and kainate receptor subunit expression in schizophrenia, bipolar disorder, and major depressive disorder

M. Beneyto and J.H. Meador-Woodruff. Mental Health Research Institute and Department of Psychiatry, University of Michigan, Ann Arbor, MI. U.S.A.

Several lines of evidence suggest abnormal expression of ionotropic glutamate receptors in schizophrenia. Recently, converging studies have found that disturbances in glutamatergic neurotransmission may also play a role in the pathophysiology of other psychiatric illnesses. Although alterations of expression of ionotropic glutamate receptors have been reported in limbic cortex and medial temporal lobe structures in schizophrenia, possible changes in the expression of these receptors in mood disorders have not been well studied. In the present study, we have determined AMPA and kainate receptor subunit expression in the dorsolateral prefrontal (dlPFC) and anterior cingulate (aCg) cortex of brains from subjects with schizophrenia, bipolar disorder, major depression, and a comparison group, using samples from the Stanley Foundation Neuropathology Consortium. We have studied these receptors by using in situ hybridization for subunit transcript expression and by receptor autoradiography. We found differences between the alterations that occur in the dlPFC versus ACg. There were no differences between the subjects with any mental illness and the control group in the Acg.

Interestingly, in dIPFC, a significant decrease of GluR2 and KA2 transcripts expression was observed, but only in major depression. These results suggest abnormal glutamatergic neurotransmission in the cerebral cortex in major depressive disorder, specific to dIPFC and not seen in aCg. Interestingly, these two receptor families are spared in these cortical areas in schizophrenia and bipolar disorder in this particular cohort of subjects.

Supported by MH53327.

Neurobiological and behavioural abnormalities induced by prenatal stimulation of dopamine D1 receptors

Sajnani, G., Meyer, G., Abreu, P. and Castro, R.

The fact that dopamine receptors are found early in brain development, prior to the formation of subcortical and cortical synaptic connections, suggests that dopamine, acting through dopamine receptors, may play an important function in neural development (1,2,3). We studied the long-term effects of prenatal SKF-38393 (a D1 receptor selective agonist) on monoamines metabolism in mesolimbic and mesostriatal systems of puber male rats. We also studied the behavioural consequences of such a manipulation.

Our results showed that low doses of SKF-38393 (0.05 mg/Kg s.c.) induced a selective reduction of either serotonin innervation in dorsal hippocampus.

Moreover, there was either a significant reduction of serotonin and 5-HIAA content, and GAP-43 expression levels, in hippocampus. These changes were followed by a selective decrease in dopamine and Dopac levels in mesolimbic system regions. Finally, a diminished vertical-locomotor activity and an altered stress-response was also found in puber rats.

These results suggest that early abnormal occupation of dopamine D1 receptors may have long-term consequences on development and function of brain limbic regions. Neuropathological and neuroimaging abnormalities have been reported in the same limbic regions in schizophrenia.

References:

1. Lankford, K.L., DeMello, F.G. and Klein, W.L. Proc. Natl. Acad. Sci. USA 85: 4567-4571 (1988).

- 2. Castro, R., Brito, B., Segovia, J., Martín-Trujillo, J.M. and Notario, V. Mol. Brain Res. 26:74-80 (1994).
- 3. Stanwood, G.D., Washington, R.A., Shumsky, J.S. and Levitt, P. Neuroscience 106:5-14 (2001).

Altered gene expression of a cell adhesion molecule subfamily in schizophrenia

Philip Ebert, Frank Middleton, Karoly Mirnics, David Lewis, and Pat Levitt

Schizophrenia is a complex disorder affecting $\sim 1\%$ of the population. Current evidence suggests that aberrant synaptic function may mediate in part the onset and progression of this disease; the functional deficits associated with schizophrenia, in addition to neuroanatomical studies, suggest that the defects underlying synaptic dysfunction are not global in nature, instead originating due to aberrant expression or function of molecules expressed in restricted regions and circuits. Members of the IgLON(K) family of genes (LSamp, OBCAM, Neurotrimin and Kilon), expressed in restricted regions of the brain (including the limbic system), encode GPI-anchored cell adhesion molecules of the Ig superfamily. Recent reports have revealed that Ig superfamily cell adhesion molecules such as SynCAM1 and neuroplastin2 are involved in synaptic assembly and maintenance, respectively, and therefore could play a role in the synaptic dysfunction associated with schizophrenia. We investigated by gene microarrays and in situ hybridization the expression of the IgLON(K) family in subjects with schizophrenia, focusing on the dorsal prefrontal cortex. The analysis revealed that the expression of the IgLON(K) family is consistently and significantly down-regulated in schizophrenia, with Kilon and OBCAM being the most robustly changed. The analysis further revealed distinct patterns of expression for each molecule in layers 5 and 6 of the human cortex. We suggest that alterations in expression of the IgLON(K) molecules reflect more global problems with synaptic function in schizophrenia.

References:

1) Biederer et al, Science 2002 Aug 30;297(5586):1525-31

2) Smalla et al, Proc Natl Acad Sci U S A 2000 Apr 11;97(8):4327-32

The effect of antipsychotic medication on the occupancy of the D2 receptor by endogenous dopamine

W. Gordon Frankle, Mark Slifstein, Anissa Abi-Dargham and Marc Laruelle

All pharmacological treatments which improve the psychotic symptoms of schizophrenia act as antagonists at the dopamine D2 receptor. The exact mechanism by which D2 antagonism improves psychotic symptoms is unclear. One hypothesis is that blockade of the D2 receptors by antipsychotic medications decreases the occupancy of these receptors by endogenous dopamine. Imaging studies have determined that the risk of extrapyramidal side effects increases when the striatal D2 receptor occupancy by an antagonist is greater than 80% [1]. However, the level of occupancy at which antipsychotic medications are effective is less well established. Some medications, notably clozapine, achieve efficacy at lower D2 occupancy levels than other medications [2]. The goal of the present study was to better characterize the degree of receptor occupancy by antipsychotic medications required to reduce occupancy of the D2 receptor by endogenous dopamine to control levels.

Our group, by examining the effect of dopamine depletion on [¹²³I]IBZM binding with SPECT, has previously demonstrated that baseline striatal dopamine levels are elevated in individuals with schizophrenia [3]. The present study utilized data of the18 healthy controls and 18 medication-free schizophrenic subjects from this prior study. The average free dopamine in both controls and schizophrenic subjects was calculated using the flowing formula [4]: (1) Free dopamine = $K_D * (BP_2/BP_1 - 1)$, where K_D is the dissociation rate constant of dopamine at the D2 receptor and BP₁ and BP₂ are the binding potentials pre- and post- dopamine can be calculated using this free concentration and the equation: (2) Occupancy = $F/(K_D + F)$, where F is concentration (nM) of free dopamine and K_D is the dissociation rate constant of dopamine at the D2 receptor (160 nM). Using this formula the calculated occupancy by dopamine of the D2 receptors is 15.8% in medication-free schizophrenic individuals and 8.8% in healthy individuals.

Equation (2) can be expanded to allow for the calculation of receptor occupancy by one species (e.g. dopamine) in the presence of another (e.g. antipsychotic). The expanded equation is: (3) Occupancy = $(F/K_D)/(1 + F_{ANT}/K_{DANT} + F/K_D)$, where F, F_{ANT} , K_D and K_{DANT} are the free concentrations and the dissociation rate constants of dopamine and the antipsychotic, respectively. With equation (3) we can calculate the striatal D2 occupancy by an antipsychotic necessary to reduce the 15.8% D2 occupancy by dopamine in schizophrenics to control levels of 8.8%. Assuming a starting level of free dopamine of 30 nM in schizophrenic subjects (15.8% occupancy) the level of D2 occupancy by an antagonist that would reduce the dopamine occupancy to that of healthy controls (8.8%) is 47%. Based on

these results it is unlikely that the sole mechanism of action of antipsychotic medications is to reduce D2 occupancy by dopamine, since, with the exception of clozapine, the D2 occupancy level at which antipsychotic medication is effective is generally greater than 50%.

References:

 Farde, L. and A.L. Nordstrom, PET examination of central D2 dopamine receptor occupancy in relation to extrapyramidal syndromes in patients being treated with neuroleptic drugs. Psychopharmacol Ser, 1993. 10: p. 94-100.

2. Farde, L., et al., Positron emission tomographic analysis of central D1 and D2 dopamine receptor occupancy in patients treated with classical neuroleptics and clozapine. Relation to extrapyramidal side effects. Arch Gen Psychiatry, 1992. 49(7): p. 538-44.

 Abi-Dargham, A., et al., From the cover: increased baseline occupancy of D2 receptors by dopamine in schizophrenia. Proc Natl Acad Sci U S A, 2000. 97(14): p. 8104-9.

4. Laruelle, M., et al., Imaging D-2 receptor occupancy by endogenous dopamine in humans. Neuropsychopharmacology, 1997. 17(3): p. 162-174.

Lateral association of Neuroligin mediate synaptic adhesion with Neurexin

and induce the formation of presynaptic terminals

Francisco G. Scholl, Camin Dean, and Peter Scheiffele

The precision of synaptic connectivity is essential for normal brain function and the formation of wrong connections leads to specific disease states such as schizophrenia where synaptic activity is upregulated. The understanding of the mechanisms that rule synaptogenesis is essential to overcome defects in synaptic activity.

Previous work has demonstrated that neuroligins, post-synaptic transmembrane proteins, can trigger the assembly of presynaptic specializations in axons contacting neuroligin transfected non-neuronal cells (1). Now, we have analyzed the mechanisms of neuroligin induced synaptogenesis in the CNS. Beta-Neurexins are plasma membrane proteins that associate in vitro with the extracellular domain of Neuroligins (2). However direct evidence of beta-neurexins as functional neuroligin receptors during synapse formation is still missing. In fact, the subcellular localization of neurexins has remained controversial. Here we demonstrate that neurexins localize to synaptic puncta in mouse cerebellar neurons. Overexpression of neuroligin-1 in neuronal cells results in a 5 fold increase in the number of presynaptic terminals contacting the transfected cells. Neurexins are recruited to the cell-cell contact site of such neuroligin-induced synapses. Neuroligin mutants lacking synaptogenic activity fail to aggregate beta-neurexin-expressing cells, however the ability of these mutants to bind soluble neurexins is not altered. The inactivating mutations map to a loop in the neuroligin extracellular domain that is required to form oligomers. These and additional results suggest that lateral association of neuroligin molecules in the postsynaptic membrane induces strong adhesion to neurexins in the presynaptic cells which trigger the formation of presynaptic terminals.

We are now studying the pathway downstream of neurexin in the presynaptic terminal. The cytoplasmic domain of neurexins link to the actin/spectrin cytoskeleton *in vitro*(3). To analyze the function neurexins might play in the assembly of the actin cytoskeleton, we microinjected beta-neurexin cDNA into NIH 3T3 cells. Neurexin expression induces a dramatic reorganization of the actin filaments. Microinjected cells form actin patches underneath plasma membrane domains containing beta-neurexin proteins. We speculate that neurexins may have a functional role on the regulation of the actin cytoskeleton at the synapse.

References:

- 1. Scheiffele P et al. Cell. 2000. 101(6):657-69
- 2. Ichtchenko K et al. Cell. 1995. 81(3):435-43.
- 3. Biederer T, and Sudhof TC. J Biol Chem. 2001. 276(51):47869-76.

Altered gene expression for BDNF signaling in the prefrontal cortex of subjects with schizophrenia

T. Hashimoto; D. W. Volk; S. E. Buchheit; D. A. Lewis

Gene expression for GAD67 and GABA transporter-1, markers of inhibitory neurotransmission, appears to be selectively altered in the parvalbumin (PV)-containing subclass of GABA neurons in the prefrontal cortex (PFC) of subjects with schizophrenia. Brain-derived neurotrophic factor (BDNF) has been shown to regulate the expression of these markers of GABA neurotransmission. To test the involvement of BDNF signaling in altered GABA-related gene expression in schizophrenia, we analyzed mRNA expression for BDNF and its receptor tyrosine kinase, TrkB, in PFC area 9 from 15 pairs of schizophrenic and control subjects, individually matched for age, sex and postmortem interval, using *in situ* hybridization. Expression of both BDNF and TrkB mRNAs, as assessed by film autoradiography, was significantly decreased in subjects with schizophrenia, whereas no change in mRNA expression was detected for TrkC, a receptor tyrosine kinase for NT-3. The magnitudes of the decreases in BDNF and TrkB mRNA expression were similar and significantly correlated across subject pairs.

Furthermore, these changes were correlated with the changes in mRNA expression for GAD67 and PV. TrkB mRNA was expressed abundantly in PV mRNA-positive neurons but not in calretinin mRNA-positive neurons in control subjects, suggesting that PV-containing neurons are most affected, compared to other PFC GABA neurons, by altered BDNF signaling in schizophrenia. The absence of the altered BDNF or TrkB mRNA expression in monkeys treated with haloperidol suggests that the gene expression changes are related to the disease process, but not to the treatment of schizophrenia. Altered BDNF signaling might affect the maintenance of inhibitory synaptic functions in the PFC of subjects with schizophrenia.

Supported by NIMH grants MH45156 and MH43784.

Evidence for an interaction between N-terminal redox sensitive cysteines and a polyamine modulatory site of the NR1/NR2A NMDA receptor

Herin, G.A., Zheng, F., Le, P., and Aizenman, E.

The NMDA subtype of glutamate receptor is sensitive to reducing and oxidizing agents in a subtype specific-manner. Cysteines 744 and 798 on NR1a are responsible for most of the redox sensitivity of NR1a/NR2B and NR1a/NR2C receptors. However, NR1a(c744a/c798a)/NR2A receptors remain sensitive to redox modulation. This has been attributed to cysteines in the amino terminal domain (ATD) of NR1 and NR2A (Choi et al., J. Neurosci. 21:392;2001). We have previously shown that 30uM spermine blocks the effects of DTT in NR1a(c744a,c798a)/NR2A receptors (Herin et al., SFN Abstracts 703.12; 2001), in a tricine-insensitive manner. This suggests a zinc-independent effect of spermine on redox modulation at the N-terminal redox-sensitive cysteines. Evidence exists that spermine alone modulates NR1a/NR2B (but not NR1a/NR2A) receptors via the N-terminal regulatory domain of NR1a. We hypothesized that spermine block of DTT potentiation in NR1a(c744a/c798a)/NR2A could be modified by alterations of the ATD of NR1a. Mutation of cys79 in NR1a(c744a/c798a)/NR2A receptors resulted in a decrease of DTT potentiation of NMDA-induced currents, as reported earlier (Choi et al., ibid.). In support of our hypothesis, the inhibitory actions of spermine on DTT potentiation were markedly attenuated in this construct. These results suggest an interaction between cvs79 of NR1a and a spermine modulatory site in NR1a/NR2A receptors.

Neurotransmitters action before synaptogenesis: GABA and glutamate modulate the migration of hippocampal pyramidal cells

J. B. Manent, M. Demarque, Y. Ben-Ari, L. Aniksztejn & A. Represa

Previous studies suggested that neuronal migration in the neocortex is modulated by neurotransmitters, mainly GABA and glutamate. We recently report that a calciumand SNARE-independent release of transmitters underlies a paracrine mode of communication before synapse formation (Demarque et al., 2002). This results in the presence of tonic, spontaneous and evoked currents in embryonic and neonatal CA1 neurons mediated primarily by the activation of GABA-A receptors. These currents persist in the presence of calcium channel blockers, botulinium toxin and are observed in Munc18-1 deficient mice in which vesicular release is abolished (Verhage et al., 2000). We postulate that such a communication may be involved in the regulation of developmental processes. In the present report we analysed if this paracrine action of transmitters contributes to modulate the migration of hippocampal neurons. Cell migration was evaluated on organotypic explants from E18 rats or mice (wild type or KO for Munc 18-1, kindly provided by Dr. TC Südhof). To follow migration two procedures were used: 1) co-culture of explants from green mice (EGFP; a gift from Dr. M. Okabe) and non-fluorescent mice, what allowed the visualisation of fluorescent migrating cells in a non-fluorescent environment. Migrating cells were recorded using the patch-clamp technique in the whole-cell configuration. 2) BrdU labelling of pregnant animals 2 hours before dissection. Slices were treated with GABA(30 μ M) or GABA-A receptor antagonist (50 µM Bicuculline) or glutamate (300 µM) or NMDA receptor antagonist (10 µM MK801). We observed that bath application of GABA and glutamate evoked currents in migrating cells, indicating that these cells already express functionnal ionotropic receptors to GABA and glutamate. Furthermore, the application of GABA-A and NMDA receptor antagonists during 1 day in culture dramatically reduces pyramidal cells migration. Since transmitters are thought to be mainly released from axonal growth cones through a vesicular mechanism, pyramidal cells migration was also investigated in co-cultures of green mice and Munc-18-1 KO mice explants. As reported before (Verhage et al., 2000) CA1 pyramidal cells migration persists in Munc 18-1 KO mice, suggesting that a synaptic release of transmitters is not required for neuronal migration. Our results suggest that GABA and glutamate released through a non-synaptic, non-conventional pathway modulate pyramidal cells migration.

Effects of the antipsychotic drug clozapine on the expression and function of the glutamate transporters GLT-1, GLAST and EAAC1 in the frontal cortex of rats

M Melone1, A Vallejo-Illarramendi2, A Perez-Samartin2, A Cozzi3, L Bragina1, DE Pellegrini-Giampietro3, C Matute2, F Contil 1Inst Hum

I Univ of Ancona, Ancona, Italy; 2 Dept of Neuronsci, Univ of the Basque Country, Bilbao Spain; 3 Dept of Pharmaco

To evaluate the potential role of glutamate transporters in the pathophysiology of schizophrenia, we studied the effects of the antipsychotic clozapine on GLT-1, GLAST and EAAC1 in the neocortex of normal rats which were given clozapine in drinking water (0,5 mg/ml; 25/35 mg/Kg/day; estimated clozapine plasma levels 20-60 ng/ml) for 9 weeks.

Immunocytochemical and immunoblotting studies showed that clozapine induced a strong reduction of GLT-1 expression in the frontal cortex (42,5%-70%) whereas GLAST and EAAC1 expression in the same cortical region were unchanged compared to controls. Glu transport in *Xenopus* oocytes injected with mRNA from the frontal cortex of treated animals was decreased by 60%. In the frontal cortex of treated animals, basal and KCI-evoked Glu output was significantly higher than in controls (about 4-fold increase for basal Glu output and up to 4-fold increase for KCI-evoked Glu output).

Our data show that clozapine administration strongly reduces GLT-1 but not GLAST and EAAC1 expression in the frontal cortex of normal rats. Thus, the robust decrease of Glu uptake and the subsequent increase in Glu levels can be attribuited to a specific effect of clozapine on GLT-1. These findings suggest that clozapine exerts at least partly its therapeutic effect by acting on GLT-1 expression and function thereby potentiating glutamatergic transmission. Furthermore, the present data suggest that glutamatergic hypofunction in schizophrenia may partly result from changes in GLT-1 expression and/or activity.

C-fos expression after cocaine or morphine sensitization in CB1 knockout mice

Patricia Murtra, Miquel Martin, Catherine Ledent, Marc Parmentier and Rafael Maldonado

Repeated intermittent treatment with psychostimulants or opioids sensitizes animals to the locomotor activating effects of these drugs. Such sensitization processes may have important implications for the understanding of addiction, mental illness, and the cellular basis of memory, but the neurobiological mechanisms involved remain only partially understood [5].

Sensitization to the locomotor responses after a chronic treatment with morphine or cocaine, and the rewarding effects of these compounds have been evaluated in CB1 knockout mice [2]. While the a cute locomotor effects of morphine were preserved in these mice, in agreement with previous studies [1], some adaptive responses to chronic morphine treatment seemed to be suppressed in CB1 knockout mice. Thus, the sensitization to hyperlocomotor activity produced by chronic morphine in wild-type animals was abolished in CB1 knockout mice. Furthermore, repeated morphine administration did not exhibit rewarding responses in the conditioned place preference paradigm in CB1 knockout mice, whereas wild-type animals showed a clear preference for the morphine-associated compartment. These data indicated that stimulation of the CB1 receptors are necessary for expressing morphine-induced motivational responses, such as the sensitization, acute locomotor effects of this compound and the adaptive responses to repeated cocaine administration were preserved in CB1 knockout mice. Thus, chronic cocaine responses was similar in both genotypes.

On the other hand, the protooncogene c-fos produces a phosphoprotein, Fos, which regulates gene transcription processes. In neuronal systems, Fos has been proposed to couple synaptic transmission to changes in gene expression by acting in the cell nucleus in concert with other proteins to form complexes in the promoter regions of target genes in response to extrinsic signals [3,4]. Therefore, mice were treated with either morphine or cocaine by using the same schedule than for the locomotor sensitisation procedure, and were then tested for induction of c-fos. We reasoned that expression of Fos could serve as a sensitive indicator of differential gene activation by drugs of abuse in a region of the brain that is directly implicated in the effects of these drugs.

References:

1. Ledent C., Valverde O., Cossu G., Petite F., Aubert J.-F., Beslot F., Bohme G.A., Imperato A., Pedrazzini T., Roques B.P., Vassart G., Fratta W. and Parmentier M. (1999) Unresponsiveness to cannabinoid and reduced addictive effects of opiates in CB-1 receptor knockout mice. Science, 283: 401-404.

2. Martin M., Ledent C., Parmentier M., Maldonado R. and Valverde O. (2000) Cocaine, but not morphine, induces conditioned place preference and sensitization to locomotor responses in CB1 knockout mice. Eur. J. Neurosci., 12: 4038-4046.

3. Sagar S.M., Sharp F.R. and Curran T. (1988) Expression of c-fos protein in brain: metabolic mapping at the cellular level. Science, 240:1328-1331.

4. Sonnenberg J.L., Macgregor-Leon P.F., Curran T. and Morgan J.I. (1989) Dynamic alterations occur in the levels and composition of transcription factor AP-1 complexes after seizure. Neuron, 3:359-365.

5. Wise R.A. and Leeb K. (1993) Psychomotor-stimulant sensitization: a unitary phenomenon? Behav. Pharmacol. 4: 339-349. Instituto Juan March (Madrid)

Reduced flow through NMDA-receptor channels as a basis for cognitive disorganisation in schizophrenia

W. A. Phillips, Steven M Silverstein, P. Uhlhass, & S. Anandaciva

Centre for Cognitive and Computational Neuroscience, Department of Psychology, University of Stirling, FK9 4LA, UK Weill Medical College, Cornell University, USA

Cognitive disorganisation involves impairment of dynamic organizational processes such as contextual modulation and dynamic grouping. There is both neurophysiological and psychological evidence for these processes, and they are at the centre of the theory of cortical computation known as Coherent Infomax (Phillips & Singer, *Behavioral and Brain Sciences* 1997, 20: 657-722). This theory has been developed both analytically and by computer simulations. Dynamic organisation could be implemented by those long-range connections within and between cortical regions that operate via NMDA-receptor channels, and which control gain as a result of their voltage-dependence. An impairment of these mechanisms is central to PCP-psychosis, and cognitive capabilities that they could provide are impaired in schizophrenic patients. Evidence for the hypothesis that schizophrenia involves impaired cognitive coordination due to NMDA-receptor hypofunction will be summarized (Phillips & Silverstein, *Behavioral and Brain Sciences*, in press).

Dopamine D1 receptors in the macaque monkey thalamus: selectivity for thalamic projection neurons

Margarita Rodríguez-Moral and Carmen Cavada

The monkey thalamus is notably innervated by dopamine (DA), mostly nuclei connected with associative and limbic cortical areas, neostriatum, and amygdala. This suggests that thalamic DA is involved in the modulation of cognitive, motor and emotional functions. With the purpose of searching for the receptors mediating DA neurotransmission in the thalamus, we studied the localization of dopamine D1 receptor in the brains of adult *Macaca nemestrina* monkeys using a subtype specific antibody.

Surprisingly, we found that D1 is present in large neuronal somata widespread throughout all thalamic nuclei. Their large size and the lack of expression of GABA or GAD indicated that the D1-immunoreactive neurons are projection neurons, not interneurons. This was confirmed by retrograde labeling from the cerebral cortex together with D1 immunodetection.

Finally, the D1 immunoreaction exhibits a perikaryal localization and does not appear associated to the plasmatic membrane of neuronal bodies, suggesting that D1 may be associated to the plasmalemma away from the neuronal body.

We propose that D1 is present in thalamocortical axon terminals based on: 1) the localization of the receptor protein in the perikaryon of thalamocortical projection neurons, and 2) the presence, previously described by others, of presynaptic D1 in axon terminals of the macaque cerebral cortex forming asymmetric synapses. Because D1 receptors have been proposed to modulate the release of neurotransmitters, including glutamate, and because thalamocortical neurons are considered glutamatergic, D1 may function as a presynaptic modulator of glutamate neurotransmission in thalamocortical systems.

Supported by DGESIC PB98-0064.

Electrophysiological study of motor function in a-Synuclein null-mice

L. Tabares#, G. Vergara and A. Rosenthal Rinat

Neuroscience. Palo Alto, 94304 CA. (USA.), and #Department of Physiology and Biophysics. School of Medicine. University of Seville. Seville (Spain)

a-Synuclein (a-Syn) is a 14 kDa protein of the mature synapses which has been implicated in the physiopathology of Parkinson's disease. However, the physiological functions of a-Syn are not yet well understood. In hippocampal neurons in culture, downregulation of a-Syn by antisense has been shown to reduce the number of synaptic vesicles, indicating that this protein may regulate the size of the presynaptic vesicular pool (1).

Expression of mutant human a-Syn forms in mice generates animals with brainstem neuronal degeneration and motor neuron pathology, this last finding suggesting that a-Syn has also a role on the synapses maintenance of the neuromuscular junction (2).

We have used mice carrying a knock-out of the Snca gene (3) to study the effect of the lack of a-Syn on motor function. The amplitude of the compound motor action potentials (CMAP) on the EDL and GN muscles in response to prolonged repetitive stimulation of the sciatic nerve were compared in knock-out and wild-type mice. In both groups, the initial amplitudes of CMAPs were similar and the decline on CMAPs amplitudes during trains of either short or prolonged high-frequency stimulations (20-100 Hz) presented similar time course. The rates of recovery of the amplitude of the CMAPs were also very similar in wild-type and knock-out muscles. Estimation of the number and size of motor units on the GN and EDL shows not differences between null- and wild-types. We also examined motor performance in rotating rod experiments (progressive speed acceleration up to 36 rpm, 5 minutes test). The motor performance of the two groups revealed no differences.

In summary, our study shows no gross sings of impair performance of the motor system on a-Syn null mice, suggesting that, at least, at this level other synapses proteins may compensate for that deficit.

References:

- 1. Murphy et al., 2000. J. Neurosc., 20:3214-3220
- 2. van der Putten et al., 2000. J. Neurosc., 20:6021-6029
- 3. Abeliovich et al., 2000. Neuron, 25: 239-252.

Antigliosis: Evidence for glial effects on synaptic plasticity in schizophrenia

Keith A. Young, Paul B. Hicks and Leigh A. Holcomb

Most investigations of glial pathology have failed to find evidence for activation or proliferation of glia (gliosis) in schizophrenia. We have used stereological techniques to estimate total glial cell populations in the anterior thalamus in subjects from the Stanley Foundation Neuropathology Consortium. There is a significant reduction in the normal glia/neuron ratio in this nucleus in schizophrenic, but not bipolar or depressed subjects. This pattern of antigliosis has also been reported in frontal cortical regions in schizophrenia (Pakkenberg, 2001). Because the schizophrenic subjects in the Stanley Foundation collection were younger than many other post-mortem studies, our data provide evidence for early glial insults in schizophrenia. We are currently in the process of studying specific glial cell subtypes to determine whether glial cell number reductions are specific for microglia, astrocytes and/or oligodendrocytes.

During the past decade, there has been a reformation in our understanding of the role of glia in CNS neurotransmission. It is well established that glia perform important developmental and trophic actions for neurons. But in addition to these functions, recent findings implicate glia as partners in local signaling activity through glial calcium gradients, and have established microglia and astrocytes as active participants in neurochemical modulation. Most recently, it has been demonstrated that normal synaptic plasticity is dependent on glial cell activity. For instance, glial-derived TNF-alpha is necessary for maintenance of normal synaptic strength of excitatory synapses (Beattie et al., 2002).

Antigliosis in schizophrenics is therefore a prime candidate in the impairment of synaptic plasticity in this mental illness. Genetic findings from our VA Cooperative Studies cohort (Skol et al., 2002) and other recent studies (Steffansson et al., 2002) have provided further support for glial involvement in schizophrenia by identifying candidate genes that participate in the determination of glial cell fates and glial activity. In combination with microarray identification of abnormal glial gene products in schizophrenic brain tissue, these recent findings suggest that antigliosis may be a intimately involved in the pathophysiology of schizophrenia.

LIST OF INVITED SPEAKERS

Guillermo Alvarez de Toledo	Department of Physiology & Biophysics. University of Seville. Avda. Sánchez Pizjuán 4, 41009 Sevilla (Spain). Tel.: 34 95 455 9856. Fax: 34 95 455 1769. E-mail: gat@us.es
Yehezkel Ben-Ari	Institut de Neurobiologie de la Méditerranée (INMED). INSERM U29. Parc Scientifique de Luminy, 13273 Marseille Cedex 09 (France). Tel.: 33 4 91 82 81 24. Fax: 33 4 91 82 81 01. E-mail: ben-ari@inmed.univ-mrs.fr
Jean P. Bourgeois	Laboratoire «Récepteurs et Cognition». Départment des Neurosciences. Institut Pasteur. 25 rue du Dr. Roux, 75724 Paris Cedex 15 (France). Tel.: 33 1 45 68 88 08. Fax: 33 1 45 68 88 36. E-mail: jpbourg@pasteur.fr
Hollis T. Cline	Cold Spring Harbor Lab. 1 Bungtown Road, Cold Spring Harbor, NY. 11724 (USA). Tel.: 1 516 367 8897. Fax: 1 516 367 6805. E-mail: cline@cshl.org
Javier DeFelipe	Cajal Institute. Avenida Dr. Arce 37, 28002 Madrid (Spain). Tel.: 34 91 585 47 35. Fax: 34 91 585 47 54. E-mail: defelipe@cajal.csic.es
Jean-Marc Fritschy	Institute of Pharmacology and Toxicology, University of Zurich. Winterthurerstrasse 190, 8057 Zürich (Switzerland). Tel.: 41 1 635 5926. Fax: 41 1 635 6874. E-mail: fritschy@pharma.unizh.ch
Patricia Gaspar	INSERM U 106, Hôpital Salpêtrière, 75651 Paris cedex 13 (France). Tel.: 33 1 53 61 26 46. Fax: 33 1 45 70 9990. E- mail: patricia.gaspar@u106.eu.org
Patricia S. Goldman-Rakic	Department of Neurobiology, Yale University School Medicine. 333 Cedar St., New Haven, CT. 06510 (USA). Tel.: 1 203 785 4808. Fax: 1 203 785 5263. E-mail: patricia.goldman-rakic@yale.edu
Stephan Heckers	Department of Psychiatry, Massachusetts General Hospital, Harvard Medical School, Boston, MA. 02115 (USA). Tel.: 1 617 724 6141. Fax: 1 617 855 3199. E-mail: heckers@ psych.mgh.harvard.edu
Marc Laruelle	Columbia Univ. College of Physicians and Surgeons. New York State Psychiatric Institute, Unit 31. 1051 Riverside Drive, New York, NY. 10032 (USA). Tel.: 1 212 543 5388. Fax: 1 212 568 6171. E-mail: ml393@columbia.edu

Juan Lerma	Instituto Cajal, CSIC. Av. Doctor Arce 37, 28002 Madrid (Spain). Tel.: 34 91 585 47 10. Fax: 34 91 585 47 54. E- mail: lerma@cajal.csic.es
Pat Levitt	J.F. Kennedy Ctr for Research on Human Development, Vanderbilt University. 230 Appleton Place, Nashville, TN. 37203 (USA). Tel.: 1 615 322 82 42. Fax: 1 615 322 59 10. E-mail: Pat.Levitt@Vanderbilt.Edu
David A. Lewis	Departments of Psychiatry and Neuroscience. University of Pittsburgh. 3811 O'Hara Street, W1650 BST, Pittsburgh, PA. 15213 (USA). Tel.: 1 412 624 39 34. Fax: 1 412 624 99 10. E-mail: lewisda@msx.upmc.edu
Jeffrey A. Lieberman	University of North Carolina School of Medicine. 7025 Neurosciences Hospital, Chapel Hill, NC. 27599-7160 (USA). Tel.: 1 919 966 8990. Fax: 1 919 966 8994. E-mail: jeffrey-lieberman@med.unc.edu
Károly Mirnics	Depts. of Psychiatry and Neurobiology, University of Pittsburgh. 3811 O'Hara Street. W1650 BST, Pittsburgh, PA. 15213 (USA). Tel.: 1 412 648 9788. Fax: 1 412 624 9910. E-mail: karoly@pitt.edu
Paul H. Patterson	Biology Division, California Institute of Technology. 391 S. Holliston, Pasadena, CA. 91125 (USA). Tel.: 1 626 395 6826. Fax: 1 626 585 8743. E-mail: php@its.caltech.edu
Thomas C. Südhof	Center for Basic Neuroscience, Dpt. of Molecular Genetics, and Howard Hughes Medical Institute, UT Southwestern Medical Center. 6000 Harry Hines Blvd., Dallas, TX. 75390 (USA). Tel.: 1 214 648 1876. Fax: 1 214 648 1879. E-mail: Thomas.Sudhof@UTSouthwestern.edu
Rafael Yuste	Dept. Biological Sciences, Columbia University. 1002 Fairchild, New York, NY. 10027 (USA). Tel.: 1 212 854 2354. Fax: 1 212 865 8246. E-mail: rmy5@columbia.edu

Instituto Juan March (Madrid)

Juan March (Madrid)

LIST OF PARTICIPANTS

Eva Alés	Dpto. Farmacología y Terapéutica. Facultad de Medicina. UAM. C/ Azobispo Morcillo, 4, 28029 Madrid (Spain). Tel.: 34 91 397 5387. Fax: 34 91 397 5380. E-mail: eva. ales@uam.es
Lidia Alonso	Instituto Cajal (CSIC). Avda. Dr. Arce, 37, 28002 Madrid (Spain). Tel.: 34 91 585 47 34. Fax: 34 91 585 47 54. E- mail: aidil@cajal.csic.es
Inmaculada Ballesteros-Yañez	Instituto Cajal (CSIC). Avda. Dr. Arce, 37, 28002 Madrid (Spain). Tel.: 34 91 585 47 35. Fax: 34 91 585 47 54. E- mail: inby_10@yahoo.com
Ruth Benavides-Piccione	Dpto. de Neuroanatomía y Biología Celular. Instituto Cajal. CSIC. Avda. Dr. Arce, 37, 28002 Madrid (Spain). Tel.: 34 91 585 4735. Fax: 34 91 585 4754
Monica Beneyto	Mental Health Research Institute and Department of Psychiatry. University of Michigan. 205 Zina Pitcher Place, Ann Arbor, MI. 48109 (USA). Tel.: 1 734 936 2056. Fax: 1 734 647 4130. E-mail: mbeneyto@umich.edu
Rafael Castro	Dpto. de Fisiología. Facultad de Medicina. Universidad de La Laguna, 38320 Tenerife (Spain). Tel.: 34 922 31 93 60. Fax: 34 922 31 93 97. E-mail: jrcastro@ull.es
Fiorenzo Conti	Università di Ancona. Via Tronto 10/A. Torrette di Ancona, 60020 Ancona (Italy). Tel.: 39 71 220 6056. Fax: 39 71 220 6052. E-mail: f.conti@popcsi.unian.it
Philip Ebert	Department of Pharmacology. Vanderbilt University. 465 21st Avenue South. MRBIII. Room 8114, Nashville, TN. 37232 (USA). Tel.: 1 615 936 3865. Fax: 1 615 936 3747. E-mail: Philip.Ebert@vanderbilt.edu
W. Gordon Frankle	Division of Functional Brain Mapping. Columbia University. New York State Psychiatric Institute. 1051 Riverside Drive, Unit 31, New York, NY. 10032 (USA). Tel.: 1 212 543 6597. Fax: 1 212 568 6171. E-mail: wf2004@columbia.edu
Francisco G. Scholl	Dept. of Physiology and Cellular Biophysics. Room 1119. Columbia University. 630 W 168th Street, New York, NY. 10032 (USA). Tel.: 1 212 342 0417. Fax: 1 212 305 5775. E-mail: fg2008@columbia.edu

Instituto

2003 WORKSHOPS

Gregory Gasic	Athinoula Martinos Biomedical Imaging Center & Harvard Medical School. Massachusetts General Hospital-East. 149, 13th Street, Charlestown, MA. 02129 (USA). Tel.: 1 617 726 0326. Fax: 1 617 726 7422. E-mail: ggasic@nmr.mgh. harvard.edu
A. Chistina Grobin	University of North Carolina. 7033 Neurosciences Hospital CB#7160, Chapel Hill, NC. 27599-7160 (USA). Tel.: 1 919 843 3794. Fax: 1 919 966 8994. E-mail: grobinac@med. unc.edu
Farid Hamzei-Sichani	State University of New York. 811 New York Ave. Apt. 1007, Brooklyn, NY. 11203 (USA). Tel.: 1 718 270 8895. Fax: 1 718 270 8895. E-mail: fh81@alumni-mail.gs. columbia.edu
Takanori Hashimoto	Department of Psychiatry. University of Pittsburgh. 200 Lothrop Street. Biomedical Science Tower W1606, Pittsburgh, PA. 15213 (USA). Tel.: 1 412 624 9909. Fax: 1 412 624 9910. E-mail: hashimotot@msx.upmc.edu
Greta A. Herin	Department of Neurobiology. University of Pittsburgh. E1405 Biomedical Science Tower, Pittsburgh, PA. 15261 (USA). Tel.: 1 412 648 9480. Fax: 1 412 648 1441. E-mail: geeann+@pitt.edu
Rachel Jones	Nature Reviews Neuroscience. The Macmillan Building, 4 Crinan Street, London N1 9XW (UK). Tel.: 44 20 7843 3632. Fax: 44 20 7843 3629. E-mail: r.jones@nature.com
Jean B. Manent	Universite de la Mediterranee. INMED. 163 route de Luminy, BP13, 13273 Marseille (France). Tel.: 33 491 82 81 14. Fax: 33 491 82 81 01. E-mail: manent@inmed.univ- mrs.fr
Emilie Marcus	Neuron. 1100 Massachusetts Ave., Cambridge, MA. 02138 (USA). Tel.: 1 617 397 2839. Fax: 1 617 397 2810. E-mail: emarcus@cell.com
Marcello Melone	Univ. of Ancona. Via Tronto 10/A Torrette di Ancona, 60020 Ancona (Italy). Tel.: 39 71 220 6056. Fax: 39 71 220 6052. E-mail: m.melone@popcsi.unian.it
Vicente Molina	Dept. of Psychiatry. Hospital Doce de Octubre. Carretera de Andalucía, km 5,4, 28041 Madrid (Spain). Tel.: 34 91 390 8536. Fax: 34 91 390 8538. E-mail: vmolina@eresmas.net
Alberto Muñoz	Dpto. de Neuroanatomia y Biología Celular. Instituto Cajal. CSIC. Avda. Dr. Arce 37, 28002 Madrid (Spain). Tel.: 34 91 585 4734. Fax: 34 91 585 4754. E-mail: amunoz@cajal. csic.es

Patricia Murtra	Neurofarmacología. Universidad Pompeu Fabra. C/ Doctor Aiguader 80, 08003 Barcelona (Spain). Tel.: 34 93 542 28 30. Fax: 34 93 542 2802. E-mail: patricia.murtra@cexs. upf.es
Naveen Nagarajan	Max-Planck Inst. for Biophysical Chemistry. Karl Friedrich Bonhoeffer Inst. University of Göttingen. Am Fassberg, 11, 37070 Göttingen (Germany). Tel.: 49 55 1201 1640. Fax: 49 55 1201 1688. E-mail: nnaveen@gwdg.de
William A. Phillips	Centre for Cognitive and Computational Neuroscience, Department of Psychology, University of Stirling, Stirling, Scotland FK9 4LA (UK). Tel.: 44 1786 467 646. Fax: 44 1786 467 641. E-mail: w.a.phillips@stir.ac.uk
Margarita Rodríguez-Moral	Dpto. de Morfología. Facultad de Medicina. Universidad Autonoma de Madrid. C/Arzobispo Morcillo, s/n, 28029 Madrid (Spain). Tel.: 34 91 397 5355. Fax: 34 91 397 5353. E-mail: margarita.rodriguez@uam.es
Eva M. Romero	Dpto. de Neuroinmunología. Instituto Cajal. CSIC. C/ Dr. Arce 37, 28002 Madrid (Spain). Tel.: 91 585 4741. Fax: 91 585 4754. E-mail: evaromero@cajal.csic.es
Miguel A. Sánchez	Dpto. de Morfología. Facultad de Medicina. Universidad Autónoma de Madrid. C/Arzobispo Morcillo s/n, 28029 Madrid (Spain). Tel.: 34 91 397 5355. Fax: 34 91 397 5353. E-mail: miguel. sanchez@uam.es
John E. Spiro	Nature Neuroscience. Nature Publishing Group. 345 Park Avenue South, New York, NY. 10010 (USA). Tel.: 1 212 726 93 19. Fax: 1 212 696 09 78. E-mail: j.spiro@ natureny.com
Lucia Tabares	Dept. of Physiology and Biophysics. School of Medicine. University of Seville. Avda. Sanchez Pizjuan, 4, 41009 Seville (Spain). Tel.: 34 95 455 1768. Fax: 34 95 455 17 69. E-mail: tabares@rinatneuro.com
Maria Toledo-Rodriguez	Center for Brain & Mind. EPLF. Ecublens, 1015 Lausanne (Switzerland). Tel.: 41 21 693 9697. Fax: 41 21 693 5350. E-mail: maria.toledo@epfl.ch
Keith A. Young	CTVHCS Neuropsychiatry Research Program. Texas AandM University System HSC. 1901 S. 1st St 151N, Temple, TX. 76504 (USA). Tel.: 1 254 899 4033. Fax: 1 254 899 6155. E-mail: Keith.Young@med.va.gov

Texts published in the SERIE UNIVERSITARIA by the FUNDACIÓN JUAN MARCH concerning workshops and courses organized within the Plan for International Meetings on Biology (1989-1991)

*: Out of stock.

- *246 Workshop on Tolerance: Mechanisms and Implications. Organizers: P. Marrack and C. Martínez-A.
- *247 Workshop on Pathogenesis-related Proteins in Plants. Organizers: V. Conejero and L. C. Van Loon.
- *248 Course on DNA Protein Interaction. M. Beato.
- *249 Workshop on Molecular Diagnosis of Cancer. Organizers: M. Perucho and P. García Barreno.
- *251 Lecture Course on Approaches to Plant Development. Organizers: P. Puigdomènech and T. Nelson.
- *252 Curso Experimental de Electroforesis Bidimensional de Alta Resolución. Organizer: Juan F. Santarén.
- 253 Workshop on Genome Expression and Pathogenesis of Plant RNA Viruses. Organizers: F. García-Arenal and P. Palukaitis.
- 254 Advanced Course on Biochemistry and Genetics of Yeast. Organizers: C. Gancedo, J. M. Gancedo, M. A. Delgado and I. L. Calderón.
- *255 Workshop on the Reference Points in Evolution. Organizers: P. Alberch and G. A. Dover.
- *256 Workshop on Chromatin Structure and Gene Expression. Organizers: F. Azorín, M. Beato and A. A. Travers.

- 257 Lecture Course on Polyamines as Modulators of Plant Development. Organizers: A. W. Galston and A. F. Tiburcio.
- *258 Workshop on Flower Development. Organizers: H. Saedler, J. P. Beltrán and J. Paz-Ares.
- *259 Workshop on Transcription and Replication of Negative Strand RNA Viruses. Organizers: D. Kolakofsky and J. Ortín.
- *260 Lecture Course on Molecular Biology of the Rhizobium-Legume Symbiosis. Organizer: T. Ruiz-Argüeso.
- 261 Workshop on Regulation of Translation in Animal Virus-Infected Cells. Organizers: N. Sonenberg and L. Carrasco.
- *263 Lecture Course on the Polymerase Chain Reaction. Organizers: M. Perucho and E. Martínez-Salas.
- *264 Workshop on Yeast Transport and Energetics. Organizers: A. Rodríguez-Navarro and R. Lagunas.
- 265 Workshop on Adhesion Receptors in the Immune System. Organizers: T. A. Springer and F. Sánchez-Madrid.
- *266 Workshop on Innovations in Proteases and Their Inhibitors: Fundamental and Applied Aspects. Organizer: F. X. Avilés.

- 267 Workshop on Role of Glycosyl-Phosphatidylinositol in Cell Signalling. Organizers: J. M. Mato and J. Larner.
- 268 Workshop on Salt Tolerance in Microorganisms and Plants: Physiological and Molecular Aspects.

Organizers: R. Serrano and J. A. Pintor-Toro.

269 Workshop on Neural Control of Movement in Vertebrates. Organizers: R. Baker and J. M. Delgado-García.

Texts published by the CENTRE FOR INTERNATIONAL MEETINGS ON BIOLOGY

- 1 Workshop on What do Nociceptors Tell the Brain? Organizers: C. Belmonte and F. Cerveró.
- *2 Workshop on DNA Structure and Protein Recognition. Organizers: A. Klug and J. A. Subirana.
- *3 Lecture Course on Palaeobiology: Preparing for the Twenty-First Century. Organizers: F. Álvarez and S. Conway Morris.
- *4 Workshop on the Past and the Future of Zea Mays. Organizers: B. Burr, L. Herrera-Estrella and P. Puigdomènech.
- *5 Workshop on Structure of the Major Histocompatibility Complex. Organizers: A. Arnaiz-Villena and P. Parham.
- *6 Workshop on Behavioural Mechanisms in Evolutionary Perspective. Organizers: P. Bateson and M. Gomendio.
- *7 Workshop on Transcription Initiation in Prokaryotes Organizers: M. Salas and L. B. Rothman-Denes.
- *8 Workshop on the Diversity of the Immunoglobulin Superfamily. Organizers: A. N. Barclay and J. Vives.
- 9 Workshop on Control of Gene Expression in Yeast. Organizers: C. Gancedo and J. M. Gancedo.

- *10 Workshop on Engineering Plants Against Pests and Pathogens. Organizers: G. Bruening, F. García-Olmedo and F. Ponz.
- 11 Lecture Course on Conservation and Use of Genetic Resources. Organizers: N. Jouve and M. Pérez de la Vega.
- 12 Workshop on Reverse Genetics of Negative Stranded RNA Viruses. Organizers: G. W. Wertz and J. A. Melero.
- *13 Workshop on Approaches to Plant Hormone Action Organizers: J. Carbonell and R. L. Jones.
- *14 Workshop on Frontiers of Alzheimer Disease. Organizers: B. Frangione and J. Ávila.
- *15 Workshop on Signal Transduction by Growth Factor Receptors with Tyrosine Kinase Activity. Organizers: J. M. Mato and A. Ullrich.
 - 16 Workshop on Intra- and Extra-Cellular Signalling in Hematopoiesis. Organizers: E. Donnall Thomas and A. Grañena.
- *17 Workshop on Cell Recognition During Neuronal Development. Organizers: C. S. Goodman and F. Jiménez.

- 18 Workshop on Molecular Mechanisms of Macrophage Activation. Organizers: C. Nathan and A. Celada.
- *19 Workshop on Viral Evasion of Host Defense Mechanisms. Organizers: M. B. Mathews and M. Esteban.
- *20 Workshop on Genomic Fingerprinting. Organizers: M. McClelland and X. Estivill.
- 21 Workshop on DNA-Drug Interactions. Organizers: K. R. Fox and J. Portugal.
- *22 Workshop on Molecular Bases of Ion Channel Function. Organizers: R. W. Aldrich and J. López-Barneo.
- *23 Workshop on Molecular Biology and Ecology of Gene Transfer and Propagation Promoted by Plasmids. Organizers: C. M. Thomas, E. M. H. Willington, M. Espinosa and R. Díaz Orejas.
- *24 Workshop on Deterioration, Stability and Regeneration of the Brain During Normal Aging. Organizers: P. D. Coleman, F. Mora and M. Nieto-Sampedro.
- 25 Workshop on Genetic Recombination and Defective Interfering Particles in RNA Viruses. Organizers: J. J. Bujarski, S. Schlesinger and J. Romero.
- 26 Workshop on Cellular Interactions in the Early Development of the Nervous System of Drosophila. Organizers: J. Modolell and P. Simpson.
- *27 Workshop on Ras, Differentiation and Development. Organizers: J. Downward, E. Santos and D. Martín-Zanca.
- *28 Workshop on Human and Experimental Skin Carcinogenesis. Organizers: A. J. P. Klein-Szanto and M. Quintanilla.
- *29 Workshop on the Biochemistry and Regulation of Programmed Cell Death. Organizers: J. A. Cidlowski, R. H. Horvitz, A. López-Rivas and C. Martínez-A.

- *30 Workshop on Resistance to Viral Infection. Organizers: L. Enjuanes and M. M. C. Lai.
- 31 Workshop on Roles of Growth and Cell Survival Factors in Vertebrate Development. Organizers: M. C. Raff and F. de Pablo.
- 32 Workshop on Chromatin Structure and Gene Expression. Organizers: F. Azorín, M. Beato and A. P. Wolffe.
- *33 Workshop on Molecular Mechanisms of Synaptic Function. Organizers: J. Lerma and P. H. Seeburg.
- *34 Workshop on Computational Approaches in the Analysis and Engineering of Proteins. Organizers: F. S. Avilés, M. Billeter and E. Querol.
- 35 Workshop on Signal Transduction Pathways Essential for Yeast Morphogenesis and Cell Integrity. Organizers: M. Snyder and C. Nombela.
- 36 Workshop on Flower Development. Organizers: E. Coen, Zs. Schwarz-Sommer and J. P. Beltrán.
- *37 Workshop on Cellular and Molecular Mechanism in Behaviour. Organizers: M. Heisenberg and A. Ferrús.
- 38 Workshop on Immunodeficiencies of Genetic Origin. Organizers: A. Fischer and A. Arnaiz-Villena.
- 39 Workshop on Molecular Basis for Biodegradation of Pollutants. Organizers: K. N. Timmis and J. L. Ramos.
- *40 Workshop on Nuclear Oncogenes and Transcription Factors in Hematopoietic Cells.

Organizers: J. León and R. Eisenman.

- *41 Workshop on Three-Dimensional Structure of Biological Macromolecules. Organizers: T. L Blundell, M. Martínez-Ripoll, M. Rico and J. M. Mato.
- 42 Workshop on Structure, Function and Controls in Microbial Division. Organizers: M. Vicente, L. Rothfield and J. A. Ayala.
- *43 Workshop on Molecular Biology and Pathophysiology of Nitric Oxide. Organizers: S. Lamas and T. Michel.
- *44 Workshop on Selective Gene Activation by Cell Type Specific Transcription Factors. Organizers: M. Karin, R. Di Lauro, P. Santisteban and J. L. Castrillo.
- 45 Workshop on NK Cell Receptors and Recognition of the Major Histocompatibility Complex Antigens. Organizers: J. Strominger, L. Moretta and M. López-Botet.
- 46 Workshop on Molecular Mechanisms Involved in Epithelial Cell Differentiation. Organizers: H. Beug, A. Zweibaum and F. X. Real.
- 47 Workshop on Switching Transcription in Development. Organizers: B. Lewin, M. Beato and J. Modolell.
- 48 Workshop on G-Proteins: Structural Features and Their Involvement in the Regulation of Cell Growth. Organizers: B. F. C. Clark and J. C. Lacal.
- *49 Workshop on Transcriptional Regulation at a Distance. Organizers: W. Schaffner, V. de Lorenzo and J. Pérez-Martín.
- 50 Workshop on From Transcript to Protein: mRNA Processing, Transport and Translation. Organizers: I. W. Mattaj, J. Ortín and J. Valcárcel.
- 51 Workshop on Mechanisms of Expression and Function of MHC Class II Molecules. Organizers: B. Mach and A. Celada.

- 52 Workshop on Enzymology of DNA-Strand Transfer Mechanisms. Organizers: E. Lanka and F. de la Cruz.
- 53 Workshop on Vascular Endothelium and Regulation of Leukocyte Traffic. Organizers: T. A. Springer and M. O. de Landázuri.
- 54 Workshop on Cytokines in Infectious Diseases. Organizers: A. Sher, M. Fresno and L. Rivas.
- 55 Workshop on Molecular Biology of Skin and Skin Diseases. Organizers: D. R. Roop and J. L. Jorcano.
- 56 Workshop on Programmed Cell Death in the Developing Nervous System. Organizers: R. W. Oppenheim, E. M. Johnson and J. X. Comella.
- 57 Workshop on NF-κB/IκB Proteins. Their Role in Cell Growth, Differentiation and Development. Organizers: R. Bravo and P. S. Lazo.
- 58 Workshop on Chromosome Behaviour: The Structure and Function of Telomeres and Centromeres. Organizers: B. J. Trask, C. Tyler-Smith, F. Azorín and A. Villasante.
- 59 Workshop on RNA Viral Quasispecies. Organizers: S. Wain-Hobson, E. Domingo and C. López Galíndez.
- 60 Workshop on Abscisic Acid Signal Transduction in Plants. Organizers: R. S. Quatrano and M. Pagès.
- 61 Workshop on Oxygen Regulation of Ion Channels and Gene Expression. Organizers: E. K. Weir and J. López-Barneo.
- 62 1996 Annual Report
- 63 Workshop on TGF-β Signalling in Development and Cell Cycle Control. Organizers: J. Massagué and C. Bernabéu.
- 64 Workshop on Novel Biocatalysts. Organizers: S. J. Benkovic and A. Ballesteros.

Organizers: M. Barbacid and D. Pulido.

- 66 Workshop on 100th Meeting: Biology at the Edge of the Next Century. Organizer: Centre for International Meetings on Biology, Madrid.
- 67 Workshop on Membrane Fusion. Organizers: V. Malhotra and A. Velasco.
- 68 Workshop on DNA Repair and Genome Instability. Organizers: T. Lindahl and C. Pueyo.
- 69 Advanced course on Biochemistry and Molecular Biology of Non-Conventional Yeasts.

Organizers: C. Gancedo, J. M. Siverio and J. M. Cregg.

- 70 Workshop on Principles of Neural Integration. Organizers: C. D. Gilbert, G. Gasic and C. Acuña.
- 71 Workshop on Programmed Gene Rearrangement: Site-Specific Recombination. Organizers: J. C. Alonso and N. D. F.

Grindley.

- 72 Workshop on Plant Morphogenesis. Organizers: M. Van Montagu and J. L. Micol.
- 73 Workshop on Development and Evolution. Organizers: G. Morata and W. J. Gehring.
- *74 Workshop on Plant Viroids and Viroid-Like Satellite RNAs from Plants, Animals and Fungi. Organizers: R. Flores and H. L. Sänger.
- 75 1997 Annual Report.
- 76 Workshop on Initiation of Replication in Prokaryotic Extrachromosomal Elements.
 Organizers: M. Espinosa, R. Díaz-Orejas, D. K. Chattoraj and E. G. H. Wagner.
- 77 Workshop on Mechanisms Involved in Visual Perception. Organizers: J. Cudeiro and A. M. Sillito.

- 78 Workshop on Notch/Lin-12 Signalling. Organizers: A. Martínez Arias, J. Modolell and S. Campuzano.
- 79 Workshop on Membrane Protein Insertion, Folding and Dynamics. Organizers: J. L. R. Arrondo, F. M. Goñi, B. De Kruijff and B. A. Wallace.
- 80 Workshop on Plasmodesmata and Transport of Plant Viruses and Plant Macromolecules. Organizers: F. García-Arenal, K. J. Oparka and P.Palukaitis.
- 81 Workshop on Cellular Regulatory Mechanisms: Choices, Time and Space. Organizers: P. Nurse and S. Moreno.
- 82 Workshop on Wiring the Brain: Mechanisms that Control the Generation of Neural Specificity. Organizers: C. S. Goodman and R. Gallego.
- 83 Workshop on Bacterial Transcription Factors Involved in Global Regulation. Organizers: A. Ishihama, R. Kolter and M. Vicente.
- 84 Workshop on Nitric Oxide: From Discovery to the Clinic. Organizers: S. Moncada and S. Lamas.
- 85 Workshop on Chromatin and DNA Modification: Plant Gene Expression and Silencing. Organizers: T. C. Hall, A. P. Wolffe, R. J. Ferl and M. A. Vega-Palas.
- 86 Workshop on Transcription Factors in Lymphocyte Development and Function. Organizers: J. M. Redondo, P. Matthias and S. Pettersson.
- 87 Workshop on Novel Approaches to Study Plant Growth Factors. Organizers: J. Schell and A. F. Tiburcio.
- 88 Workshop on Structure and Mechanisms of Ion Channels. Organizers: J. Lerma, N. Unwin and R. MacKinnon.
- 89 Workshop on Protein Folding. Organizers: A. R. Fersht, M. Rico and L. Serrano.

- 90 1998 Annual Report.
- 91 Workshop on Eukaryotic Antibiotic Peptides. Organizers: J. A. Hoffmann, F. García-Olmedo and L. Rivas.
- 92 Workshop on Regulation of Protein Synthesis in Eukaryotes. Organizers: M. W. Hentze, N. Sonenberg and C. de Haro.
- 93 Workshop on Cell Cycle Regulation and Cytoskeleton in Plants. Organizers: N.-H. Chua and C. Gutiérrez.
- 94 Workshop on Mechanisms of Homologous Recombination and Genetic Rearrangements.

Organizers: J. C. Alonso, J. Casadesús, S. Kowalczykowski and S. C. West.

- 95 Workshop on Neutrophil Development and Function. Organizers: F. Mollinedo and L. A. Boxer.
- 96 Workshop on Molecular Clocks. Organizers: P. Sassone-Corsi and J. R. Naranjo.
- 97 Workshop on Molecular Nature of the Gastrula Organizing Center: 75 years after Spemann and Mangold. Organizers: E. M. De Robertis and J. Aréchaga.
- 98 Workshop on Telomeres and Telomerase: Cancer, Aging and Genetic Instability. Organizer: M. A. Blasco.
- 99 Workshop on Specificity in Ras and Rho-Mediated Signalling Events. Organizers: J. L. Bos, J. C. Lacal and A. Hall.
- 100 Workshop on the Interface Between Transcription and DNA Repair, Recombination and Chromatin Remodelling. Organizers: A. Aguilera and J. H. J. Hoeijmakers.
- 101 Workshop on Dynamics of the Plant Extracellular Matrix. Organizers: K. Roberts and P. Vera.

102 Workshop on Helicases as Molecular Motors in Nucleic Acid Strand Separation. Organizers: E. Lanka and J. M. Carazo.

103 Workshop on the Neural Mechanisms of Addiction. Organizers: R. C. Malenka, E. J. Nestler and F. Rodríguez de Fonseca.

- 104 1999 Annual Report.
- 105 Workshop on the Molecules of Pain: Molecular Approaches to Pain Research. Organizers: F. Cervero and S. P. Hunt.
- 106 Workshop on Control of Signalling by Protein Phosphorylation. Organizers: J. Schlessinger, G. Thomas, F. de Pablo and J. Moscat.
- 107 Workshop on Biochemistry and Molecular Biology of Gibberellins. Organizers: P. Hedden and J. L. García-Martínez.
- 108 Workshop on Integration of Transcriptional Regulation and Chromatin Structure. Organizers: J. T. Kadonaga, J. Ausió and E. Palacián.
- 109 Workshop on Tumor Suppressor Networks. Organizers: J. Massagué and M. Serrano.
- 110 Workshop on Regulated Exocytosis and the Vesicle Cycle. Organizers: R. D. Burgoyne and G. Álvarez de Toledo.
- 111 Workshop on Dendrites. Organizers: R. Yuste and S. A. Siegelbaum.
- 112 Workshop on the Myc Network: Regulation of Cell Proliferation, Differentiation and Death. Organizers: R. N. Eisenman and J. León.
- 113 Workshop on Regulation of Messenger RNA Processing. Organizers: W. Keller, J. Ortín and J. Valcárcel.
- 114 Workshop on Genetic Factors that Control Cell Birth, Cell Allocation and Migration in the Developing Forebrain. Organizers: P. Rakic, E. Soriano and A. Álvarez-Buylla.

- 115 Workshop on Chaperonins: Structure and Function. Organizers: W. Baumeister, J. L. Carrascosa and J. M. Valpuesta.
- 116 Workshop on Mechanisms of Cellular Vesicle and Viral Membrane Fusion. Organizers: J. J. Skehel and J. A. Melero.
- 117 Workshop on Molecular Approaches to Tuberculosis. Organizers: B. Gicquel and C. Martín.
- 118 2000 Annual Report.
- 119 Workshop on Pumps, Channels and Transporters: Structure and Function. Organizers: D. R. Madden, W. Kühlbrandt and R. Serrano.om
- 120 Workshop on Cmon Molecules in Development and Carcinogenesis. Organizers: M. Takeichi and M. A. Nieto.
- 121 Workshop on Structural Genomics and Bioinformatics. Organizers: B. Honig, B. Rost and A. Valencia.
- 122 Workshop on Mechanisms of DNA-Bound Proteins in Prokaryotes. Organizers: R. Schleif, M. Coll and G. del Solar.
- 123 Workshop on Regulation of Protein Function by Nitric Oxide. Organizers: J. S. Stamler, J. M. Mato and S. Lamas.
- 124 Workshop on the Regulation of Chromatin Function. Organizers: F. Azorín, V. G. Corces, T. Kouzarides and C. L. Peterson.
- 125 Workshop on Left-Right Asymmetry. Organizers: C. J. Tabin and J. C. Izpisúa Belmonte.
- 126 Workshop on Neural Prepatterning and Specification. Organizers: K. G. Storey and J. Modolell.
- 127 Workshop on Signalling at the Growth Cone. Organizers: E. R. Macagno, P. Bovolenta and A. Ferrús.

- 128 Workshop on Molecular Basis of Ionic Homeostasis and Salt Tolerance in Plants. Organizers: E. Blumwald and A. Rodríguez-Navarro.
- 129 Workshop on Cross Talk Between Cell Division Cycle and Development in Plants. Organizers: V. Sundaresan and C. Gutiérrez.
- 130 Workshop on Molecular Basis of Human Congenital Lymphocyte Disorders. Organizers: H. D. Ochs and J. R. Regueiro.
- 131 Workshop on Genomic vs Non-Genomic Steroid Actions: Encountered or Unified Views. Organizers: M. G. Parker and M. A. Valverde.
- 132 2001 Annual Report.
- 133 Workshop on Stress in Yeast Cell Biology... and Beyond. Organizer: J. Ariño.
- 134 Workshop on Leaf Development. Organizers: S. Hake and J. L. Micol.
- 135 Workshop on Molecular Mechanisms of Immune Modulation: Lessons from Viruses. Organizers: A. Alcami, U. H. Koszinowski and M. Del Val.
- 136 Workshop on Channelopathies. Organizers: T. J. Jentsch, A. Ferrer-Montiel and J. Lerma.
- 137 Workshop on Limb Development. Organizers: D. Duboule and M. A. Ros.
- 138 Workshop on Regulation of Eukaryotic Genes in their Natural Chromatin Context. Organizers: K. S. Zaret and M. Beato.
- 139 Workshop on Lipid Signalling: Cellular Events and their Biophysical Mechanisms. Organizers: E. A. Dennis, A. Alonso and I. Varela-Nieto.
- 140 Workshop on Regulation and Functional Insights in Cellular Polarity. Organizers: A. R. Horwitz and F. Sánchez-Madrid.

- 141 Workshop on The Structure of the Cortical Microcircuit. Organizers: R. Yuste, E. M. Callaway and 'H. Markram.
- 142 Workshop on Control of NF-κB Signal Transduction in Inflammation and Innate Immunity. Organizers: M. Karin, I. M. Verma and J. Moscat.
- 143 Workshop on Engineering RNA Virus Genomes as Biosafe Vectors. Organizers: C. M. Rice, W. J. M. Spaan and L. Enjuanes.
- 144 Workshop on Exchange Factors. Organizers: X. R. Bustelo, J. S. Gutkind and P. Crespo.
- 145 Workshop on the Ubiquitin-Proteasome System. Organizers: A. Ciechanover, D. Finley, T.

Sommer and C. Mezquita.

146 Workshop on Manufacturing Bacteria: Design, Production and Assembly of Cell Division Components.

Organizers: P. de Boer, J. Errington and M. Vicente.

- 147 2002. Annual Report.
- 148 Workshop on Membranes, Trafficking and Signalling during Animal Development.

Organizers: K. Simons, M. Zerial and M. González-Gaitán.

*: Out of Stock.

The Centre for International Meetings on Biology was created within the Instituto Juan March de Estudios e Investigaciones, a private foundation specialized in scientific activities which complements the cultural work of the Fundación Juan March.

The Centre endeavours to actively and sistematically promote cooperation among Spanish and foreign scientists working in the field of Biology, through the organization of Workshops, Lecture Courses, Seminars and Symposia.

> From 1989 through 2002, a total of 176 meetings, all dealing with a wide range of subjects of biological interest, were organized within the scope of the Centre.



Instituto Juan March de Estudios e Investigaciones Castelló, 77 • 28006 Madrid (España) Tel. 34 91 435 42 40 • Fax 34 91 576 34 20 http://www.march.es/biology

The lectures summarized in this publication were presented by their authors at a workshop held on the 10th through the12th of February, 2003, at the Instituto Juan March.

All published articles are exact reproduction of author's text.

There is a limited edition of 400 copies of this volume, available free of charge.