# Instituto Juan March de Estudios e Investigaciones

# 38 CENTRO DE REUNIONES INTERNACIONALES SOBRE BIOLOGÍA

## Workshop on

# Immunodeficiencies of Genetic Origin

Organized by

A. Fischer and A. Arnaiz-Villena

F. W. Alt A. Arnaiz-Villena R. M. Blaese H. von Boehmer A. O. Carbonara M. Cooper A. L. Corbí A. Corell A. Fischer R. A. Gatti

IJM

38

Wor

R. S. Geha
W. J. Leonard
B. Mach
T. W. Mak
H. D. Ochs
J. R. Regueiro
F. S. Rosen
G. de Saint Basile
C. I. E. Smith

## Instituto Juan March de Estudios e Investigaciones

CENTRO DE REUNIONES INTERNACIONALES SOBRE BIOLOGÍA

# 38



Workshop on

Immunodeficiencies of Genetic Origin

Organized by

A. Fischer and A. Arnaiz-Villena

F. W. Alt A. Arnaiz-Villena R. M. Blaese H. von Boehmer A. O. Carbonara M. Cooper A. L. Corbí A. Corell A. Fischer R. A. Gatti R. S. Geha W. J. Leonard B. Mach T. W. Mak H. D. Ochs J. R. Regueiro F. S. Rosen G. de Saint Basile C. I. E. Smith

The lectures summarized in this publication were presented by their authors at a workshop held on the 6th and 7th of March, 1995, at the Instituto Juan March.

Depósito legal: M. 16.682/1995 I. S. B. N.: 84-7919-538-X Impresión: Ediciones Peninsular. Tomelloso, 27. 28026 Madrid.

## INDEX

A. Fischer and A. Arnaiz-Villena: Introduction	7
FIRST SESSION: T-CELL DIFFERENTIATION AND DEFICIENCIES Chairman: Max Cooper	11
Tak W. Mak: T Lymphocyte function and ontogeny in gene- targeted mutant mice	13
Frederick W. Alt: Identification of 3 different genes that encode products involved in V(D)J recombination and DNA repair	15
Antonio Arnaiz-Villena: Papillon-Lefevre immunodeficiency	17
Alain Fischer: Apo-1/Fas/CD95 mutations in human lymphoproliferative syndrome associated with autoimmunity and defective apoptosis	18
Bernard Mach: MHC class II deficiency, a disease of gene regulation	19
José R. Regueiro: CD3 deficiencies in man and mouse	21
<b>Geneviève de Saint Basile:</b> X-SCID caused by IL2Rγ mutations	22
Warren J. Leonard: The role of the common cytokine receptor $\gamma$ chain in X-linked severe combined immunodeficiency and in cytokine signaling	24
FIRST SESSION: T-CELL DIFFERENTIATION AND DEFICIENCIES (continuation) Chairman: Alain Fischer	27
Raif S. Geha: CD40/CD40L interaction and immunodeficiency	29
Fred S. Rosen: Wiskott-Aldrich syndrome	30
Hans D. Ochs: Mutations of the WASP gene cause classic Wiskott-Aldrich gene and X-linked thrombocytopenia	31
Short oral presentations: Ignacio J. Molina: A new experimental system for studying the pathogenesis of the Wiskott-Aldrich syndrome and the function of the WASP gene	33

Instituto Juan March (Madrid)

PAGE

#### PAGE

Antonio Celada: The transcription factor PU.1 is necessary for macrophage proliferation	35
<b>Balbino Alarcón:</b> A tyrosine-containing motif mediates ER retention of CD3- $\epsilon$ and adopts a helix/turn structure	36
SECOND SESSION: B CELL DIFFERENTIATION DEFICIENCIES Chairman: Fred S. Rosen	37
<b>Max Cooper:</b> B cell differentiation defects in X-linked agammaglobulinemia, IgA deficiency and common variable immunodeficiency	39
<b>C.I. Edvard Smith:</b> X-linked agammaglobulinemia: A tyrosine kinase disease	40
Angelo Oscar Carbonara: Unequal crossing over as source of IGHC locus variability	42
Short oral presentation: Teresa Español: IgA deficiency. Clinical and immunological heterogeneity	43
THIRD SESSION: GENE THERAPY Chairman: Maxime Seligmann	45
<b>R. Michael Blaese:</b> T lymphocyte gene therapy for ADA deficiency (SCID): Results of the initial trial with 4 years observation	47
Harald von Boehmer: Thymic Selection	49
FOURTH SESSION: OTHER IMMUNODEFICIENCIES Chairman: Maxime Seligmann	51
Alfredo Corell: Virus infection models affecting CD3 activation	53
Angel L. Corbí: Leukocyte adhesion deficiency (LAD)	54
<b>R.A. Gatti:</b> Ataxia-telangiectasia: Pathogenesis and positional cloning	56
Short oral presentation: Miguel A.R. Marcos: Two early phases of lymphohematopoiesis in the mouse embryo	58

### PAGE

POSTERS	59
Ana C. Carrera: Structure/function analysis of the tyrosine kinase pp <sup>56kk</sup> involved in the establishment of immunocompetence	61
Manuel Hernández: CD40L expression in hyper IgM syndrome and CVID	62
Manel Juan: BamHI and PstI polymorphism of the human CD50 (ICAM-3) gene	63
Nathalie Nicolas: The human autosomal SCID and Omenn's syndrome: An increased radiosensitivity without DNA repair defect	64
Lourdes Tricas: Clinical and immunological study from a hyper-IgM immunodeficiency syndrome	65
<b>Igor Vořechovský:</b> A rapid PCR-SSCP mutation analysis of the Bruton's tyrosine kinase gene in X-linked agammaglobulinaemia	67
Igor Vořechovský: Pilot linkage mapping of selective immunoglobulin a deficiency and common variable immune deficiency: Lack of evidence for an involvement of	
the MHC region in families with autosomal dominant segregation	68
Henning Walczak: APO-1/Fas (CD95) mediated T cell receptor induced suicide	69
A. Fischer and A. Arnaiz-Villena: Perspectives	71
LIST OF INVITED SPEAKERS	75
LIST OF PARTICIPANTS	79

## INTRODUCTION

## A. Fischer and A. Arnaiz-Villena

This booklet is a collection of summaries of presentations at the "Immunodeficiencies of Genetic Origin" workshop held in Madrid on 6-7 March 1995.

The idea to celebrate such a meeting was put forward because of the overwhelming information published in the last years on the precise molecular and genetic anomalies which underlay many inmunodeficiencies. Abnormal surface molecules, defective tyrosin-kinases and mutated genes among others have been spotted in children with immune system dysfunctions.

Andrés González rapidly materialized the proposal and Alain Fischer and Antonio Arnaiz-Villena called most of the scientists who have reported the molecular basis of inherited deficiencies, and others who are in the front line of research on gene therapy, thymic ontogeny and immune genes "knock out" mice. The scientists response was good and most of them accepted the invitation. The magnificent and internationally reputed organization of these series of meetings was maintained at its highest level by Andrés González, as usual.

Only inherited immunodeficiencies affecting lymphocytes were studied in the workshop. Theoretically it is possible to find out the genetic defects at any level of immunological ontogeny and functionality; thus, examples of deficiencies were presented at the following immune response steps:

- 1.- Antibody presenting cells or B-T cell contacts: mutations at CD18 (LFA-1) molecule or CD40 ligand; "bare lymphocyte syndrome" with absence of HLA class II presenting molecules and TAP-2 deficiency (peptide transporter molecule for HLA class I antigens) may also be classified within this item.
- 2.- B-T cell specific genes rearrangement defects: progress in recombinase defects was shown and multiple enzymes with genes in different chromosomes were involved.
- 3.- **B cell antibody production:** Bruton agammablobulinemia due to a defect of *btk* (Bruton tyrosin-kinase).
- 4.- **T cell recognition:** defect at the CD3 complex has been described;  $\epsilon$  and  $\gamma$  chain mutant families there exist.
- 5.- **T cell response:** a defect in ZAP-70 tyrosin-kinase coupled to CD3 is responsible for a defective transmission signal and a severe immunodeficiency is associated to a defective  $\gamma$  subunit which is common to the receptors for IL-2, IL-4, IL-7, IL-9 an IL-15.
- 6.- T cell cooperation: CD40 ligand deficiency could also be included here. In addition, abnormal IL-2 production and reduction/lack of NF-AT have been detected without knowing the precise gene involved.
- 7.- Toxic metabolites: accumulated in ADA and PNP deficiencies result in a serious immunodeficiency, which may be treated by adequate gene therapy.
- 8.- There are other inmunodeficiencies, whose genetic defect is pin pointed but the mechanisms by which immunodeficiency is established are still obscure; these are the cases of the Wiskott-Aldrich syndrome and the Fas deficiency.

In addition, artificially impaired genes in mice ("knock out" mice) showed how a lack of certain surface proteins and interleukins affected the immune function; the last advances in T-cell ontogeny were also detailed (i.e.: The precise nature of pre-T cell  $\alpha$  chain).

The problems of gene therapy were also stressed by the example of the adenosindeaminase deficiency treatment; the infusion of culture - expanded peripheral blood T cells modified by retroviral mediated ADA gene-transfer can be an effective therapy for this severe immunodeficiency.

In summary, the precise physiology of the immune system is being clarified by these inherited natural (human) or "knock out" (mouse) occurring genetic errors.

It is likely that the study of immunodeficiencies of genetic origin will be one of the most productive field in biology. This study has been possible due to the widespread availability of new technology, including the existence of specific monoclonal antibodies against surface molecules, molecular genetics, easy gene amplification, Ca<sup>++</sup> and tyrosin-kinases study methodology.

Immunology and physiology in general will benefit of the discoveries and preventive and even curative medicine will be used to fight anomalies.

# FIRST SESSION: T-CELL DIFFERENTIATION AND DEFICIENCIES

Chairman: Max Cooper

# T LYMPHOCYTE FUNCTION AND ONTOGENY IN GENE-TARGETED MUTANT MICE

Tak W. Mak

Ontario Cancer Institute, Toronto, Ontario/Canada

T lymphocytes recognize their antigen peptides and Major Histocompatibility Complex products with the use of their T ceil antigen receptors (TeR). In addition to the  $\alpha$  and  $\beta$  ohains of TeR, the interaction between T cells and their target cells or antigen presenting cells is also assisted by a series of other cell surface polypeptides. Most notable of these are CD4 and CD8, which are selectively expressed on mature helper/induces and killer/suppressor T cells, respectively. Upon engagement of their ligands, a series signals are being transduced intracytoplasmically via some of these molecules and their associated proteins. Perhaps the most important enzyme in this signal transduction process is the lymphocytes specific tyrosine kinase lck. Another important component is the cell surface tyrosine phosphatase CD45. This molecule is alternatively spliced and the different isoforms are expressed on the various hemopoietic and lymphopoictic cells. Signaling through the TcR-CD4/CD8-lck-CD45 complex is thought to be insufficient to activate T lymphocytes. A co-stimulatory signal is believed to be essential. Many investigators have suggested that CD28, a ligand for B7/BB1 is an essential co-stimulatory signal. In an attempt to gain better understanding on the roles of these molecules in T lymphocyte functions and ontogeny, we generated a series of mutant mice with disruptions in these genes. These mutant mice are being analyzed in order that we can evaluate the importance of these genes in T cell development.

References:

1) Fung-Leung, W.-P., Schilham, M., Rahemtulla, A., Vollenweider, M., Potter, J., van Ewijk, W. & Mak, T.W. (1991) CD8 is needed for development of cytotoxic T cells but not helper T cells. Cell 65: 443-449.

2) Rahemtulla, A., Fung-Leung, W.-P., Schilham, M.W., Kundig, T.M., Sambhara, S.R., Narendran, A., Arabian, A., Wakeham, A., Paige, C., Zinkernagel, R.M., Miller, R.G. & Mak, T.W. (1991) Mice lacking CD4 have normal development and function of CD8<sup>+</sup> cells but have markedly decreased helper cell activity. Nature 353: 180-184.

3) Molina, T., Kishihara, K., Siderovski, D., van Ewijk, W., Paige, C., Hartmann, K., Veillette, A., Davidson, D. & Mak, T.W. (1992) Profound block in thymocyte development in mice lacking p56<sup>1</sup>Ck. Nature 357: 161-164.

4) Koh, D.-R., Fung-Lcung, W.-P., Ho, A., Gray, D., Acha-Orbea, H. & Mak, T.W. (1992) Less mortality but more relapses in experimental allergic encephalomyelitis (EAE) in CD8<sup>-/-</sup> mice. Science 256: 1210-1213.

5) Penninger, J., Kishihara, K., Molina, T., Wallace, V.A., Timms, F., Hedrick, S.M. & Mak, T.W. (1993) Requirement for tyrosine kinase p56lck for thymic development of transgenic γδ T cells. Science 260: 358-361.

6) Pfeffer, K., Matsuyama, T., Kundig, T.M., Wakeham, A., Kishihara, K., Shahinian, A., Wiegmann, K., Ohashi, P.S., Kronke, M. & Mak, T.W. (1993)

7) Kishihara, K., Penninger, J., Wallace, V.A., Kundig, T.M., Kawai, K., Wakeham, D., Timms, E., Pfeffer, K., Ohashi, P., Thomas, M.L., Furlonger, C., Paige, C.J. & Mak, T.W. (1993) Normal B lymphocyte development but impaired T cell maturation in CD45-exon6 tyrosine phosphatase deficient micc. Cell 74:143-156.

8) Matsuyama, T., Kimura, T., Kitagawa, M., Pfeffer, K., Kawakami, T., Watanabe, N., Kundig, T.M., Amakawa, R., Kishihara, K., Wakeham, A., Potter, J., Furlonger, C.L. Narendran, A., Suzuki, H., Ohashi, P.S., Paige, C.J., Taniguchi, T. & Mak, T.W. (1993) Targeted disruption of IRF-1 or IRF-2 results in abnormal type I IFN gene induction and aberrant lymphocyte development. Cell 75: 1-20.

9) Shahinian, A., Pfeffer, K., Lee, K., Kundig, T., Kishihara, K., Wakeham, A., Kawai, K., Ohashi, P., Thompson, C. & Mak, T.W. (1993) Differential T cell costimulating requirements in CD28 deficient mice. Science 261: 609-612.

10) Tanaka, N., Ishihara, M., Kitagawa, M., Harada, H., Kimura, T., Matsuyama, T., Lamphier, M.S., Aizawa, S.H., Mak. T.W. & Taniguchi, T. (1994) Cellular commitment to oncogene-induced transformation or apoptosis is dependent on the transcription factor IRF-1. Cell 77: 829-39. Identification of 3 Different Genes that Encode Products Involved in V(D)J Recombination and DNA Repair. Guillermo E. Taccioli, Zhiying Li, #Tanya M. Gottlieb, \*Tracy Blunt, #Nicholas J. Finnie, Jocelyne Demengeot, \*Anne Priestley, Ryushin Mizuta, Yijie Gao, +Tomas Otevrel, \*Alan Lehman, +Thomas Stamato, \*Penny Jeggo, #Stephen P. Jackson, and Frederick W. Alt. The Howard Hughes Medical Institute, Children's Hospital, Center for Blood Research and Department of Genetics, Harvard University Medical School, Boston. #Wellcome/CRC Institute and Department of Zoology, Cambridge University, UK. \*Medical Research Council, Cell Mutation Unit, University of Sussex, Brighton, UK.\*Lankenau Medical Research Center, Wynnewood, Pa.

The V(D)J recombination reaction involves recognition of conserved recombination sequences (RS) that flank each germline V, D, or J segment, introduction of double strand breaks at the RS/coding sequence junctions, loss and/or addition of nucleotides at the coding junctions, and additional activities (e.g. polymerization and ligation) to complete the joining process. V(D)J recombination appears to be carried out by both pre-lymphocyte specific factors such as RAG-1, RAG-2 and TdT as well as generally expressed factors including gene products involved in double strand DNA break repair (DSBR) pathways.

We have previously shown that three independent Chinese hamster ovary (CHO) mutant cell lines that have defects in double strand DNA break repair (DSBR) also have defects in V(D)J recombination as assayed with transient V(D)J recombination substrates following introduction of the RAG-1 and 2 genes into these cell lines. These findings suggest that the DSBR and V(D)J recombination processes share common factors. In combination with its V(D)J defect, the mutant V-3 CHO line has a defect that affects V(D)J recombination in a manner essentially identical to that found in the context of the homozygous murine *scid* mutation : V(D)J recombination is properly initiated, RS joining occurs normally, but coding join formation is impaired. We have shown by cell complementation studies that the murine *scid* and V-3 mutations probably affect the same gene. Two additional, independent CHO cell DSBR mutants (xrs-6 and XR-1) can initiate V(D)J recombination properly, but are blocked in ability to form both RS and coding joins.

We have obtained both genetic and biochemical evidence which demonstrates that the human gene (XRCC-5; chromosome 2) which complements the xrs-6 mutant encodes the 80 kDa subunit of the Ku complex. The complex of the 80 and 70kDa Ku subunits binds to free double stranded DNA ends and forms the DNA binding component of the DNA-dependent protein kinase (DNA-PK). The large catalytic subunit of DNA-PK (DNA-PKcs; approximately 450kDa), once activated through its interaction with DNA bound Ku can phosphorylate a number of proteins *in vitro* including transcription factors and p53. A variety of biochemical data indicate that DNA-PK activity is defective in the V-3/Scid cell lines due to a defect in the DNA-PKcs molecule. Furthermore, YACs containing the human DNA-PKcs gene can complement the defective DNA-PK activity, DSBR defect, and V(D)J recombination defect in the V-3 line. This and other data suggest that the DNA-PKcs is the XRCC-7 gene (human chromosome 8) which complements the V-3 (murine Scid) defect.

Finally, we have recently cloned a human cDNA sequence, which encodes a unique protein, based on its ability to restore normal V(D)J recombination activity to the XR-1 cell line. Expression of this cDNA sequence in XR-1 cells also substantially complements their DSBR defect. We have mapped the gene (XRCC-4 gene) encoding this cDNA to human chromosome 5. Current efforts are aimed at clucidating potential functions of the XRCC-4 gene product in DSBR and V(D)J recombination.

#### References.

1. Taccioli, G.E., Rathbun, G., Oltz, E., Stamato, T., Jeggo, P.A., and Alt, F.W. (1993) Impairment of VDJ recombination in double strand break repair mutants. *Science* 620, 207-210.

2. Taccioli, G.E., Cheng, H.-W., Varghese, A.J., Whitmore, G., and Alt, F.W. (1994) A DNA Repair Defect in Chinese Hamster Ovary Cells Affects V(D)J recombination similarly to the murine *Scid* mutation. J. Biol. Chem. 269:7439-7442.

3. Taccioli, G.E., Gottlieb, T.M., Blunt, T., Pricstly, A., Demengcot, J., Mizuta, R., Lehmann, A.R., Alt. F.W., Jackson, S.P., and Jeggo, P.A. (1994) Ku80 is the product of the XRCC5 gene: A direct link between DNA repair and V(D)J recombination. Science 265:1442-1445.

4. Blunt, T., Finnie, F.J., Taccioli, G.E. Smith, G.C.M., Demengeot, J., Gottlieb, T.M., Mizuta, R., Varghese, A.J., Alt, F.W., Jeggo, P.A., and Jackson. S.P. (1995) Defective DNA-dependent protein kinase activity is linked to V(D)J recombination and DNA repair defects associated with the murine *SCID* mutation. Submitted for Publication.

#### Papillon-Lefevre Immunodeficiency

Antonio Arnaiz Villena, Hospital Universitario 12 de Octubre, Universidad Complutense, 28041 MADRID

The Papillon-Lefevre (P.F.) syndrome is caused by an unknown autosomic recessive genetic anomaly and is characterized by a severe periodontal inflammation and palmoplantar hyperkeratosis. 30 % patients show increased susceptibility to bacterial . infections.

We have shown that patients with P.F. syndrome show usually normal percentages of peripheral blood lymphocytes analyzed by many CDs including integrins, CD2 and CD3. However, slightly low CD45 R0 and CD29 number of lymphocytes were recorded during a 7 year follow up, and all times a reduced number of CD18, CD11a and CD2 molecules were obtained in the corresponding positive cells; the high density LFA-1 and CD2 cells were strongly diminished. LFA-1 and CD2 molecules were normal as assessed by immunoprecipitation. It is postulated the existence of a common mechanism that regulate CD2 and LFA-1 expression, which is impaired in P.F. patients.

## Apo-1/Fas/CD95 MUTATIONS IN HUMAN LYMPHOPROLIFERATIVE SYNDROME ASSOCIATED WITH AUTOIMMUNITY AND DEFECTIVE APOPTOSIS

J-P. DE VILLARTAY\*, F. RIEUX-LAUCAT\*, F. LE DEIST\*, C. HIVROZ\*, R. DE CHASSEVAL\*, I.A.G. ROBERTS#, K.M. DEBATIN\*\*, A. FISCHER\* \* INSERM U 429, Hôpital Necker-Enfants Malades, Paris, France \*\* University children's hospital, Heidelberg, Germany # Dept. Haematology, Hammersmith Hospital, London, UK

The Apo-1/Fas/CD95 antigen has been identified as a key cell surface receptor involved in apoptotic cell death. It was found defective in the lpr mouse. Fas expression and function were analysed in three children (including two siblings) with a lymphoproliferative syndrome and autoimmune evidence (thrombocytopenia, hemolytic anemia and neutropenia) in two of them. A restricted  $\beta$  T-cell repertoire was detected in one of the 3 children both in double negative and single positive T-cells. A large genomic deletion encompassing the terminal part of the intracellular domain-encoding exon was responsible for the lack of detectable Fas cell surface expression in the most affected patient. Less severe clinical manifestation was found in the two related patients. Fas expression was normal in these patients but Fas-mediated T and B cell apoptosis was impaired because of a truncated protein resulting from a 2bp deletion within the intracytoplasmic domain.

This Fas gene-defect in humans is to our knowledge the first description of a complete Fas deficiency (the lpr mice have residual Fas expression) and confirms the crucial regulatory role of this molecule in lymphocyte cell death and may provide a molecular basis for susceptibility to autoimmune diseases in humans.

F.RIEUX-LAUCAT et al submitted.

#### MHC CLASS II DEFICIENCY, A DISEASE OF GENE REGULATION Bernard Mach, M.D., Ph.D., University of Geneva Medical School

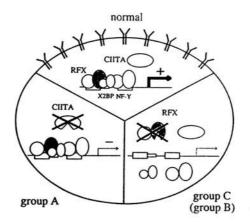
When it was shown that a form of primary immunodeficiency, MHC class II deficiency (also referred to as the Bare Lymphocyte Syndrome, BLS), was in fact a disease of gene regulation, involving transacting regulatory factor(s) (1), there was hope that solving the genetic basis of that rare disease might lead to the identification of essential regulators of MHC class II gene expression. MHC class II genes are indeed very tightly regulated and the control of their expression in certain selected cell types is an essential component of the control of the immune response.

MHC class II deficiency is an autosomal recessive disease characterized by severe immunodeficiency with multiple infections (2). The clinical picture results from an absence of HLA-DR, -DQ and -DP molecules on all cells of these patients. This unusual phenotype was in turn shown to result from a lack of transcription from the promoters of the different MHC class II genes. Although clinically homogeneous, the disease is genetically heterogeneous, with three complementation groups described, A,B and C (3,4).

A first clue to the regulatory defect responsible for this disease came from the observation that patients from BLS group B and C exhibit a characteristic defect in the binding of a specific protein complex, called RFX, to the X box motif of MHC class II promoters (5), while patients from complementation group A bind RFX normally. This defines two distinct types of molecular defects. Genetic complementation of cell lines corresponding to each of these two types of defects by cDNA libraries in expression vectors has now allowed the cloning and identification of the regulatory genes involved in each of these two types of defects. Patients from complementation group A (as well as some in vitro generated regulatory mutant lines that belong to the same group) have a normal promoter binding pattern but are mutated in a novel MHC class II transactivator called CIITA (6) (see Figure, bottom left). Patients from group C, characterized by the lack of binding of RFX in vitro and an unoccupied promoter in vivo, are indeed mutated in the gene that encodes the large subunit of the RFX complex (Steimle V, Durand B, Barras E, Zufferey M, Mach B, Reith W, in preparation). In both cases, introduction of the normal, non-mutated, form of the regulatory gene (as cDNA) into MHC class II negative B lymphocytes from BLS patients restores full expression of all MHC class II genes and cell surface molecules in several patients from each of these two groups. Although the gene mutated in complementation group B has not been cloned yet, it is tempting to suggest that it encodes the other subunit of RFX, since purified RFX heterodimers can correct MHC class II transcription in vitro in extracts from group B as well as group C patients (7). The two types of molecular defects identified in promoters of BLS patients, and the two transacting regulatory factors involved are schematically represented in the Figure (bottom left and right), with obvious consequences in terms of cell surface expression of class II molecules. Somewhat unexpectedly, it was discovered that CIITA, the MHC class II transactivator mutated in BLS, is the mediator of this crucial induction process (8). Expression of the CIITA gene is, itself, very tightly controlled, induced for instance by interferon gamma, and it is both necessary and sufficient to activate MHC class II gene expression (8). In fact, CIITA behaves as the main regulator of MHC class II expression in a variety of biological situations.

Because of the importance of MHC class II expression in numerous pathological situations, including in disease that are not T lymphocyte-mediated, the novel MHC class II transactivators discovered in studying BLS patients should probably be considered for novel immunomodulation strategies. Contrary to many transcription factors, RFX and CIITA are both absolutely essential and remarkably specific for MHC class II gene expression. These properties make them excellent targets for immunomodulators that would act at the transcriptional level.

- de Préval C, Lisowska-Grospierre B, Loche M, Griscelli C, Mach B. A trans-acting class II regulatory gene unlinked to the MHC controls expression of HLA class II genes. Nature 1985;318:291-293.
- Griscelli C, Lisowska-Grospierre B, Mach B. Combined immunodeficiency with defective expression in MHC class II genes. In: Rosen FS, Seligman M., eds. Immunodeficiencies. Chur, Switzerland: Harwood Academic Publishers, 1993, pp. 141-54.
- Hume CR, Lee JS. Congenital immunodeficiencies associated with absence of HLA class II antigens on lymphocytes result from distinct mutations in trans-acting factors. Hum Immunol 1989;26:288-309.
- Benichou B, Strominger JL. Class II antigen-negative patient and mutant cell lines represent at least three, and probably four, distinct genetic defects defined by complementation analysis. Proc Natl Acad Sci USA 1991;88:4285-8.
- Reith W, Satola S, Herrero Sanchez C, Amaldi I, Lisowska-Grospierre B, Griscelli C, Hadam MR, Mach B. Congenital immunodeficiency with a regulatory defect in MHC class II gene expression lacks a specific HLA-DR promoter binding protein, RF-X. Cell 1988;53:897-906.
- Steimle V, Otten LA, Zufferey M, Mach B. Complementation cloning of an MHC Class II transactivator mutated in hereditary MHC class II deficiency (or Bare lymphocyte syndrome). Cell 1993;75: 135-146.
- Durand B, Kobr M, Reith W, Mach B. Functional complementation of MHC class II regulatory mutants by the purified X box binding protein RFX. Mol Cell Biol 1994;14:6839-6847.
- Steimle V, Siegrist CA, Mottet A, Lisowska-Grospierre B, Mach B. Regulation of MHC class II expression by interferon-gamma is mediated by the transactivator gene CIITA. Science 1994;265:106-109.



The two types of molecular defects in MHC class II deficiency

## CD3 DEFICIENCIES IN MAN AND MOUSE José R. Regueiro Inmunología, Facultad de Medicina, Universidad Complutense 28040 Madrid, tel/fax 34 1 3941642/1

The CD3 chains of the T lymphocyte antigen receptor (TCR) are multiple monomorphic components of uncertain stoichiometry most likely involved in signal transduction for T cell selection or activation through a range of protein kinases. The recognition of natural human defects (CD3 $\gamma$ , CD3 $\epsilon$ ) and the generation of mouse KOs (CD3 $\delta$ , CD3 $\zeta$ , CD3 $\eta$ ) are helping to understand the redundant and the specialized roles of the invariant chains of the TCR/CD3 complex. The main effects of the CD3 deficiencies on T-cell development, selection and on TCR/CD3 assembly, structure, expression and signaling will be reviewed.

#### References

- Arnaiz-Villena A., Timón M., Corell A., Perez-Aciego P., Martín-Villa JM., Regueiro JR. (1992). Primary immunodeficiency caused by mutations in the gene encoding the CD3- $\gamma$  subunit of the T-lymphocyte receptor. N Engl J Med 327, 529-533.

- Soudais C., De Villartay JP., Le Deist F., Griscelli C., Fischer A., Lisowska-Grospierre B. (1993). Independent mutations of the human CD3€ gene resulting in a T cell receptor/CD3 complex immunodeficiency. Nat Genet 3, 77-81.

- Ohno H., Aoe T., Taki S., Kitamura D., Ishida Y., Rajewsky k., Saito T. (1993). Developmental and functional impairment of T cells in mice lacking CD3ζ chains. EMBO J 12, 4357-4366.

- Love PE., Shores EW., Johnson MD., Tremblay ML., Lee EJ., Grinberg A., Huang SP., Singer A., Westphal H. (1993). T cell development in mice that lack the  $\xi$  chain of the T cell antigen receptor complex. Science 261, 918-921.

- Ohno H., Goto S., Taki S., Shirasawa T., Nakano H., Miyatake S., Aoe T., Ishida Y., Maeda H., Shirai T. (1994). Targeted disruption of the CD3 eta locus causes high lethality in mice: modulation of Oct-1 transcription on the opposite strand. EMBO J 13, 1157-65.

#### X-SCID caused by IL2Ry mutations

## G. de Saint Basile, J. DiSanto, A. Dautry-Varsat, F. Le Deist, M. Cavazzana-Calvo, S.

Hacein-Bey, S. Markiewicks, F. Rieux-Laucat, E. Morellon, A. Fischer

X chromosome-linked severe combined immunodeficiency disease (SCID X1) is a recessive hereditary disorder characterized by a complete absence of immature and mature cells and natural killer (NK) cells, whereas B cells are present in normal or elevated numbers. SCID X1 infants present with severe and persistent infections resulting in failure to thrive and early death in the presence of curative bone marrow transplantation (BMT). Mutation of the  $\gamma$  chain common ( $\gamma$ c) to interleukin (IL) 2, 4, 7, 9 and 15 receptors has been shown to induce the SCID X1. However, although some mutations lead to the typical form of SCID X1, others mutations are responsible to mild forms of SCID where patient present with some T cell differenciation and function.

SCID X1 patients presenting with various phenotypes were therefore analyzed for  $\gamma c$  chain mutation and some functions. In nine patients with typical form of SCID X1, multiple genetic defects in the extracellular region of the  $\gamma c$  chain were detected, which affect in most cases  $\gamma c$  membrane expression and in all cases, high affinity IL2 binding.

In an other patient, an atypical phenotype of SCID X1 was observed consisting in an absence of T and NK cells during the first year of life and in further developpement of an autologous mature T cell population with mild mitogen and antigen response. Raise in T cell counts as well as in T cell function occurs following an allogenic bone marrow transfert which fails to engraft. Identification of a  $\gamma c$  nonsens mutation predicts a truncation of the major part of the intracytoplasmic  $\gamma c$  region which precludes any  $\gamma c$ -Jak3 association in this case. This mutation however, does not affect high affinity IL2 binding but partly reduce IL2 endocytosis. These data indicate that in certain circumstances, some accessory signaling pathways can partly overcome  $\gamma c$  chain defect to allow T cell differenciation and function although a severe immune deficiency persists.

A familial atypical form of SCID X1 characterized by the presence of a low to normal

Instituto Juan March (Madrid)

### 22

number of poorly functional and oligoclonal T cells of NK cells and by normal serum concentration of the Ig isotypes resulted in an other SCID-X1 patient from a 5 fold reduction  $\gamma c$  expression. High affinity IL2 binding was normal.  $\gamma c$  chain mutation in this patient consists in a substitution of the penultimate base of the first exon causing a splice defect. However alternative splice preserved a functional mRNA in 20% of splice products.

Since expression of a reduced number of  $\gamma c$  chain causes a severe immunodeficiency, effective gene therapy of SCID X1 will probably require high level of  $\gamma c$  expression. First data on  $\gamma c$  retrovirus-infected EBV-B cell line of a SCID X1 patient indicate that selection of transduced B cell clones expressing normal  $\gamma c$  chain occur during in vitro culture and that no evidence for a dominant negative effect of the mutated transcript on the normal  $\gamma c$  expression was found.

Together, these data raise new questions on the role of the  $\gamma c$  chain in T cell development and differenciation and bring preliminary informations on the normal  $\gamma c$  chain expression in transduced SCID X1 patient cells.

X-linked severe combined immunodeficiency (XSCID) is the most common form of SCID, accounting for approximately half of all cases. Noguchi et al. (1993) demonstrated that mutations in the IL-2 receptor y chain results in XSCID in humans. This finding was unexpected since mice and humans deficient in IL-2 have normal numbers of T cells, whereas patients with XSCID have absent or profoundly diminished numbers of T cells. The dilemma was resolved by the striking observations that IL-2Ry is not only a component of the IL-2 receptor, but also is a critical component of the receptors for IL-4, IL-7, IL-9. and IL-15, and hence is also known as the common cytokine receptor y chain, ye. Thus, inactivation of  $\gamma_c$  simultaneously inactivates at least five cytokine systems, explaining the severe immunodeficiency present in XSCID patients. The phenomenon of sharing of cytokine receptor subunits exists for two other sets of cytokines. IL-6, IL-11, oncostatin M, leukemia inhibitory factor, CNTF, and cardiotrophin 1 all share gp130 as a signaling molecule, whereas IL-3, IL-5, and GM-CSF all share the common  $\beta$  chain. These observations collectively have implications related to the understanding of cytokine pleiotropy and redundancy.

In the case of IL-2, it has been established that dimerization of the cytoplasmic domains of IL-2R $\beta$  and  $\gamma_c$  is required for signaling, whereas the role of IL-2Ra is primarily to augment binding affinity and sensitivity of responsiveness to IL-2. A number of laboratories have sought to identify the signaling molecules which can associate with each of these chains. Considerable energy has focused on Janus family tyrosine kinases. IL-2 activates both Jak1 and Jak3, but not Jak2 or Tyk2. Corresponding to this observation, it has been demonstrated that Jak1 associates with IL-2R $\beta$  and Jak3 associates primarily with  $\gamma_c$ , but Jak3 can also associate directly with IL-2R $\beta$ . Interestingly, IL-4, IL-7, and IL-9 also activate Jak1 and Jak3, indicating that the Jak family kinases are not responsible for the differences in signals transduced by these different cytokines. Distinctive proteins, such as IRS-1, which is activated by IL-4, may help to determine specificity. Moreover, the Stat proteins activated by some of these cytokines are different. For example, IL-2 and IL-4 activate different Stat proteins. However, IL-2 and IL-7, which share a number of actions, activate the same Stat proteins in normal peripheral blood lymphocytes. We have clarified the molecular basis for this phenomenon by identifying the tyrosines which when phosphorylated serve as docking sites for the relevant Stat proteins. This serves to provide a molecular basis for some of the signals induced by IL-2 and other cytokines.

Finally, as part of an effort to understand  $\gamma_c$  in an in vivo model, we and others have inactivated the  $\gamma_c$  gene in mice by homologous recombination. The interesting phenotype of these mice contributes further to our understanding of cytokine pleiotropy and redundancy based on both similarities and differences in mice and humans lacking expression of this protein.

### **References:**

- Discovery of γ<sub>c</sub> as the defect in XSCID Noguchi, M. et al., Cell 73, 147-157, 1993.
- Discover of  $\gamma_c$  as a component of the IL-4 and IL-7 receptors. Kondo, M. et al., Science 262, 1874-1877, 1993. Noguchi, M. et al., Science 262, 1877-1880, 1993. Russell, S.M. et al., Science 262, 1880-1883, 1993. Kondo, M. et al., Science 263, 1453-1454, 1994.
- Association of Jak1 and Jak3 with IL-2Rβ and  $\gamma_c$ Boussiotis, et al., Science 266, 1039-1042, 1994. Russell et al., Science 266, 1042-1045, 1994. Miyazaki et al., et al., Science 266, 1045-1047, 1994.

## General reviews

Conley, M.E., Annu. Rev. Immunol. 10, 215-238, 1992. Leonard, W.J. et al., Immunol. Rev. 138, 61-86, 1994

## FIRST SESSION: T-CELL DIFFERENTIATION AND DEFICIENCIES (Continuation)

Chairman: Alain Fischer

### CD40/CD40L interaction and imunodeficiency

#### Raif S. Geha

Interaction between CD40 and CD40L play an important role in T-B cell collaboration. Stimulation of T cells via the TCR/CD3 that recognizes MHC class II peptide complex induces CD40L expression. Engagement of CD40 by CD40L induces expression of B7 molecules on B cells. Interaction of B7 with CD28/CTLA4 costimulates T cells to proliferate and secrete cytokines. Engagement of CD40 on B cells induces isotype switching to the immunoglobulin locus targeted by T cell cytokine e.g. IL-4 primes B cells to switch to IgE.

We have shown that T cell but not anti-CD40 directed isotype switching is deficient in X linked hyperIgM syndrome and that patients with this disease have mutations in the CD40L. We will present limited data on some unusual mutations.

We have created CD40 deficient mice by disruption of the CD40 gene. These mice fail to undergo immunoglobulin isotype switching in response to T cell dependent antigens and fail to develop germinal centers. B cells from these mice fail to elicit proliferation in allogeneic cells. This is reversed by anti-CD28. More importantly, *in vivo* injection of CD40-/- B cells specifically tolerizes the recipient T cells to alloantigens.

Because of its importance in B cell function, we studied the expression of CD40 in B cell ontogeny. CD40 was not detectable on pro-B cells. CD40 was expressed on pre-B cells; however CD40 ligation on pre-B cells did not transduce a signal as determined by CD23 expression and by proliferation in the presence of IL-4. Expression of the bcl-2 transgene permitted pre-B cells to respond to CD40 ligation.

The CD40 signalling complex was investigated. CD40 was associated on the cell surface with a 26kD protein with unique amino acid sequences. p26 association with CD40 was contingent on the presence of the extracellular domain of CD40 and was esssential for CD40 mediated aggregation but not for CD40 mediated lymphotoxin  $\alpha$  gene expression.

The transcriptional regulation of CD40L expression was investigated. There are four consensus NFAT sites in the CD40 regulatory region. These sites bind NFATc and NFATp, but are not transcriptionally active until the NFAT proteins associate with an AP-1 complex containing Fos and Jun proteins as determined by reporter gene luciferase assays in T cells transfected with multimers of the NFAT sites. Wiskott-Aldrich Syndrome

Fred S. Rosen, M.D.

Department of Pediatrics, Harvard Medical School; Center for Blood Research, Boston, MA, USA

The Wiskott-Aldrich syndrome (WAS) is a monogenic defect, which maps to Xp11.23 between OATL1 and GATA 1. The gene that is defective in WAS has been cloned. It encodes a protein of 501 amino acid residues, which has been called the WAS protein. Several mutations have been identified in this protein in WAS patients. It contains several tyrosine residues in its amino-terminal portion that may be phosphorylated and bind to SH2 domains. It also has several stretches of polyproline in its carboxy-terminal portion that bind SH3 domains. It is not related to any protein in the data bank. Its function is at present unknown.

The genetic defect appears to affect blood cell lineages. Obligate female heterozygous carriers exhibit non-random X-inactivation of all leukocyte and megakaryocyte lineages and as well as in these lineages early in commitment. Phenotypically the thrombocytes and T lymphocytes are most profoundly affected and exhibit a pattern of disorganization of their cytoskeleton. As a consequence of this the T lymphocytes are bald and lack microvilli and consistently fail to respond mitogenically to insolubilized anti-CD3. The cell surface sialoglycoproteins are unstable in the T cell membrane.

#### REFERENCES

Derry JMJ, Ochs HD, Francke U. Isolation of a novel gene mutated from Wiskott-Aldrich syndrome. Cell 78:635, 1994.

Kwan S-P, Hagemann TL, Radtke B, Blaese RM, Rosen FS. Identification of mutations in the Wiskott-Aldrich syndrome gene and characterization of a polymorphic dinucleotide repeat DXS6940 adjacent to the disease gene. Proc Natl Acad Sci USA, in press, 1995.

Molina IJ, Kenney DM, Rosen FS. Remold-O'Donnell E. T cell lines characterize events in the pathogenesis of the Wiskott-Aldrich syndrome. J Exp Mcd 176:867, 1992.

Molina IJ, Sancho J, Terhorst C, Rosen FS, Remold-O'Donnell E. T cells of patients with the Wiskott-Aldrich syndrome have a restricted defect in proliferative responses. J Immunol 151:4383, 1993.

Mutations of the WASP Gene Cause Classic Wiskott-Aldrich Gene and X-linked Thrombocytopenia

Hans D. Ochs, Qili Zhu, Min Zhang, Jonathan M. J. Derry, R. Michael Blaese, Anne Junker, Shi-Han Chen, Uta Francke.

The Wiskott-Aldrich syndrome (WAS) is an X-linked recessive disorder characterized by thrombocytopenia, small platelet volume, eczema, recurrent bacterial and viral infections, autoimmune disease, increased risk of malignancies and abnormal B and T cell function (1). In addition to the classic WAS phenotype, patients with congenital thrombocytopenia and small platelets, but without eczema or significant infections, and with normal or only mildly defective immune system, have been described (2). This milder form of WAS has been mapped to the same region of the human X chromosome (3). The gene responsible for the Wiskott-Aldrich syndrome (WASP) has been isolated. cloned and sequenced (4). The gene, composed of 12 exons containing 1823 basepairs, encodes a 502 amino acid proline rich protein that appears to be of central importance for the development of hematopoietic cell lineages. Sequence analysis of genomic DNA, originally carried out in three families with WAS, revealed three independent mutations within exon 2, strongly suggesting that the newly identified gene was associated with WAS (4). To confirm that the WASP gene is responsible for the majority of WAS patients irrespective of clinical phenotype, we selected for mutation analysis the affected members of twelve unrelated WAS families with clinical symptoms and immunologic abnormalities that ranged from mild to severe. To identify the approximate location of mutations within WAS cDNA, we first screened the entire cDNA with the dideoxynucleotide fingerprinting method. This procedure revealed abnormal sequences in all twelve patients studied and allowed us to predict the approximate location of each defect. Based on the screening results, we then amplified the implicated regions of the WAS cDNA for precise analysis of the mutation. Finally, based on the mutations identified by direct sequence analysis of cDNA, we selected and designed primer pairs that allowed amplification of the corresponding regions of genomic DNA.

Point mutations were the most frequent mutations observed, being present in eight of the eleven unique mutations identified. Two unrelated patients with a mild form of WAS were found to have a  $C \rightarrow T$  mutation at position 290, within exon 2, resulting in the substitution of arginine with cysteine at codon 86. Another missense mutation, affecting codon 125 of exon 4, was found in a patient with a severe phenotype. Two patients, both with severe symptoms of WAS, were found to have nonsense mutations resulting in direct stop codons within exon 1 and exon 10, respectively. In all instances, the same point mutations involving the same nucleotides were found in genomic DNA.

Point mutations resulting in deletions or insertions were observed in four familes. These included the skipping of exon 3 in a patient with severe WAS, deletion of exon 11 due to a splice site mutation in a patient with mild WAS, the deletion of 13 nucleotides within exon 9 due to a new splice site in a patient with mild symptoms, and the insertion of 114 nucleotides 5' of intron 9 due to alternative splicing.

Insertions and deletions of genomic DNA, and secondarily of cDNA, were observed in three unrelated WAS patients. These included the insertion of a single nucleotide in exon 1, a deletion and insertion of small DNA fragments, resulting in a new splice site and the addition of 13 nucleotides at the beginning of exon 11, and the deletion of a nucleotide in exon 10, all resulting in premature stop and moderate to severe clinical phenotypes.

Since the clinical phenotype of WAS often varies from family to family, but also within kinships (1), it is almost impossible to correlate the clinical phenotypes with location and type of mutations. However, in two of our patients with mutations affecting nucleotide 290 ( $C \rightarrow T$  transition) the clinical symptoms were very mild, consisting only of thrombocytopenia, small platelets and, in one of the two cases, mild eczema. Both of these patients fulfill the criteria of X-linked congenital thrombocytopenia, suggesting that this variant is part of the spectrum of WAS. We have previously reported two unrelated Spanish families ("classic WAS") with mutations affecting the arginine at codon 86, in one case due to a  $G \rightarrow T$  transversion of nucleotide 291 (Arg86Leu), and in the other case due to a  $G \rightarrow A$  transition of the same nucleotide (Arg86His). These findings suggest that codon 86 is a hot spot and may be mutated in up to 25% of WAS patients.

#### References:

- Sullivan KE, Mullen CA, Blaese RM, Winkelstein JA: A multi-institutional survey of the Wiskott-Aldrich syndrome. J Pediatrics 1994, in press
- Stormorken H, Hellum B, Egeland T, Abrahamsen TG, Hovig T: X-linked thrombocytopenia and thrombocytopathia: attenuated Wiskott-Aldrich syndrome. Thromb Haemost 65:300-305, 1991
- De Saint-Basile G, Schlegel N, Caniglia M, Le Deist F, Kaplan C, Lecompte T, Piller E, Fischer A, Griscelli C: X-linked thrombocytopenia and Wiskott-Aldrich syndrome: similar regional assignment but distinct X-inactivation pattern in carriers. Am Hematol 63:107-110, 1991.
- Derry JMJ, Ochs HD, Francke U: Isolation of a novel gene mutated in Wiskott-Aldrich syndrome. Cell 78 (4) August 26, 1994

#### A NEW EXPERIMENTAL SYSTEM FOR STUDYING THE PATHOGENESIS OF THE WISKOTT-ALDRICH SYNDROME AND THE FUNCTION OF THE WASP GENE.

#### Juan M. Millán, Enrique Aguado, Francisco J. García-Cózar, Manuel Santamaría, José Peña and Ignacio J. Molina\*

Departments of Immunology, University of Cordoba School of Medicine, 14004 Cordoba, and \*University of Granada School of Medicine, 18012 Granada, Spain.

The Wiskott-Aldrich syndrome (WAS) is a primary immunodeficiency whose responsible gene (WASP) has very recently been identified (Derry et al, Cell, 78:635). Nevertheless, the function of WASP remains unclear.

We have been working towards the development of an adequate experimental system to better understand the pathogenesis of WAS. A first step in this direction was the establishment of a panel of allospecific T cell lines from WAS patients (Molina et al, J. Exp. Med. 176:867). These cells have discriminated previously described abnormalities, showing that the CD43 alteration is a secondary defect origininated in the circulation, whereas surface morphology abnormalities are primary defects directly linked to the WAS gene. Furthermore, we showed a specific defect in the CD3 signal transduction pathway of WAS T cell lines that is by all indications directly linked to the WAS gene (Molina et al, J. Immunol. 151: 4383). The CD3-mediated signal is initially transduced but interrupted at an unidentified step. Interestingly, the allospecific stimulation mediated by the  $\alpha/\beta$  TCR chains is normally completed, resulting in vigorous proliferation.

Attempts to improve this experimental system have been successful by infecting the panel of allospecific T cell lines with the Herpesvirus Saimiri 488. This virus is able to immortalize T lymphocytes after interacting with a surface receptor not known yet. The resulting cells are functionally and phenotypically stable, and remain dependent of IL-2 for growth.

We report here the initial characterization of two WAS allospecific T cell lines and one normal counterpart after immortalization following HVS infection. These cell lines have been kept in culture for over 6 months, and during this time have maintained their phenotype and dependence of exogenous IL-2, although they do not need allospecific stimulation. The doubling time varies with every cell line, but generally is 2-4 days.

Upon stimulation with the anti-CD3 mAb OKT3, the immortalized WAS cell lines GOR1-HVS and GOR7-HVS failed to proliferate (left panel), whereas the allospecific RAJI cells trigger a vigorous proliferative response comparable to the immortalized normal counterpart DEE8-HVS, measured in a 72 h <sup>3</sup>H-thymidine uptake assay (right panel). This mirrors the findings of the WAS parental cell lines, and likewise, the early CD3 signal transduction events are intact, including phosphorylation of the CD3- $\zeta$  chain after OKT3 stimulation.

The surface phenotype of WAS-HVS cells does not show any difference respect either the HVS-immortalized normal cell line or the WAS parental cells. Under the scanning electronmicroscope, the examined GOR1-HVS line shows a cell surface completely devoided Instituto Juan March (Madrid) of microvilli. This is a fundamental primary alteration in WAS found in both fresh PBLs and T cell lines. In this case, we have found a very homogeneous and consistent defect.

Finally, the recent cloning of the WASP gene has allowed us to address the expression of the WASP gene. By using a combination of a 5' sense primer with an internal antisense primer, we have detected by Reverse transcriptase-PCR WASP message in normal cells. However, we have not found WASP mRNA levels in GOR1-HVS cells. The DNA mutations contained in the WASP gene of our established cell lines remains to be determined.

Attempts to determine whether a normal WASP gene can rescue our WAS cell lines are currently in progress.

In summary, by immortalizing the allospecific WAS T cell lines, we have improved our experimental system aimed to the understanding of the pathogenic mechanisms of WAS.

## The transcription factor PU.1 is necessary for macrophage proliferation

A. Celada, M.J. Klemsz, C. van Beveren y R.A. Maki

Departamento de Bioquímica y Fisiología (Inmunología), Facultad de Biología, Universidad de Barcelona and La Jolla Cancer Research Foundation, La Jolla, California, USA

PU.1 is an *ets* related transcription factor that is specific for macrophages and B lymphocytes. In serum free media, bone marrow derived macrophages are able to produce colonies. In the presence of M-CSF, proliferation of PU.1 transfected macrophages was enhanced in relation to the controls. A similar effect was observed in the presence of GM-CSF, but not in the presence of IL1, IL2, IL3, IL4, IL6 or G-CSF.

Macrophages transfected with antisense PU.1 showed a decreased response in proliferation to M-CSF in relation to macrophages transfected with the control vector. A decreased proliferation was also observed when antisense oligonucleotides were added to the culture media containing M-CSF.

Mutations by alanines of the serines 41 and 45, that are sites of casein-kinase depending phosphorylation, abrogate the proliferative effect of PU.1.

These experiments demonstrated that the transcription factor PU.1 is necessary for macrophage proliferation and also that phosphorylation of PU.1 is required for the proliferative effect. Instituto Juan March (Madrid)

# A TYROSINE-CONTAINING MOTIF MEDIATES ER RETENTION OF CD3- $\epsilon$ and adopts a Helix / Turn structure

Arrate Mallabiabarrena<sup>1</sup>, M. Angeles Jiménez<sup>2</sup>, Manuel Rico<sup>2</sup> and Balbino Alarcón<sup>1</sup>

(1) Centro de Biología Molecular Severo Ochoa, CSIC-Universidad Autónoma de Madrid, Cantoblanco Madrid 28049, Spain. (2) Instituto de Estructura de la Materia, CSIC, Serrano 119, Madrid 28006, Spain.

The CD3- $\varepsilon$  endoplasmic reticulum (ER) retention motif has been characterized by mutagenesis and nuclear magnetic resonance spectroscopy. Tyrosine 177, leucine 180 and arginine 183 are involved in ER retention. The motif forms an elongated  $\alpha$ -helix in which the tyrosine and leucine residues are closely apposed, followed by a  $\beta$ l' turn that places arginine 183 in the vicinity of leucine 180. The structure formed by tyrosine 177 and the leucine in position +3 is reminiscent of the  $\beta$ -turn structure adopted by tyrosine-containing endocytosis signals. Moreover, substitution of the transferrin receptor (TfR) internalization sequence by the CD3- $\varepsilon$  motif still allowed the rapid internalization of the TfR and, conversely, the chimeric protein resulting from the substitution of the CD3- $\varepsilon$  motif by the endocytosis signal of the low density lipoprotein receptor was ER located. These data support the idea of a functional homology between the two types of signal.

## SECOND SESSION: B CELL DIFFERENTIATION DEFICIENCIES

Chairman: Fred S. Rosen

## B CELL DIFFERENTIATION DEFECTS IN X-LINKED AGAMMAGLOBULINEMIA, IgA DEFICIENCY AND COMMON VARIABLE IMMUNODEFICIENCY

## Max Cooper\*, Linda Billips, Norihiro Nishimoto, Hiromi Kubagawa, Peter Burrows, Harry Schroeder, Stephen Powis, Francesco Cucca, Mauro Congia, Duncan Campbell, Zeng-bian Zhu and John Volanakis

\*University of Alabama at Birmingham and The Howard Hughes Medical Institute, Birmingham, AL

Mutations in the Btk gene have been shown to be responsible for the arrest in B cell differentiation that occurs in boys with X-linked agammaglobulinemia (XLA) and the B lineage cells in female XLA carriers that use the X chromosome containing the defective Btk gene. The cellular abnormality is evident by the late pro-B to pre-B cell stage in differentiation. Our studies suggest that signals generated through the CD19 molecule and the IL-7 receptor arc important in normal pro-B cell differentiation and that these signal pathways are impaired by function-loss mutations in the Btk gene. Selective IgA deficiency (IgA-D) and common variable immunodeficiency (CVID), the two most common primary immunodeficiencies, represent polar ends of a disease spectrum that reflect a common genetic predisposition. One clue to the identity of a susceptibility gene is the frequent occurrence of an extended complex of ancestral MHC genes featuring deletion of the C4A complement gene in family members having either IgA-D or CVID. C4A can bind to antigenantibody complexes and can guide these to receptors on B cells to facilitate activation by antigen. An alternative hypothesis, that a class II gene in the ancestral MHC haplotype predisposes to the immunodeficiency, is rendered less likely by the absence of IgA deficiency in a group of Sardinian individuals who are homozygous for an MHC haplotype featuring an ancestral cross-over between the disease-associated class II genes and the region of the MHC gene complex containing the C4A gene.

#### X-linked agammaglobulinemia: A tyrosine kinase disease

C. I. Edvard SMITH, MD, PhD

Center for BioTechnology, Karolinska Institute, NOVUM, S-141 57 Huddinge Sweden. Fax: +46 8 774 55 38, telephone: +46 8 608 9114.

X-linked agammaglobulinemia (XLA) was the first immunodeficiency identified (Bruton 1952) and is characterized by reduced levels of immunoglobulin and B cells, resulting in a propensity for recurrent bacterial infections (Lederman & Winkelstein 1985, Sideras & Smith 1995). In a collaborative positional cloning project, a cDNA enrichment technique was employed using yeast artificial chromosomes from the implicated region of the X-chromosome (Vetrie et al. 1993, Vorechovsky et al. 1994). The same gene was also identified by searching for novel protein-tyrosine kinases (PTKs) (Tsukade et al. 1993).

The gene involved in XLA encoded a novel cytoplasmic FTK named Bruton's agammaglobulinemia tyrosine kinase (Btk). In another collaborative effort we could demonstrate that the mouse immunodeficiency Xid (X-linked immunodeficiency) is caused by a mutation in the mouse Btk gene (Thomas et al. 1993). Btk belongs to a new family of cytoplasmic PTK's formed by Tec, Itk (also named Tsk and Emt), reviewed in Smith et al. 1994, and Bmx (Tamagnone et al. 1994).

Btk is expressed in most hematopoietic cell lines with the absence of T lymphocytes and plasma cells (Smith et al. 1994). Btk represents the first cytoplasmic PTK in which mutations have been found to cause a hereditary disease. This gives us the unique possibility to make genotype-phenotype correlations in patients to directly delineate functionally important regions in the Btk molecule. In total more than a hundred mutations have been found. An international study group has recently been formed that will provide a mutation database. Each patient will be assigned a unique patient identity number and details on the mutation will be given.

We have recently used molecular modeling techniques to investigate the effect of mutations causing amino acid substitutions. A model of the kinase domain indicated structural clustering of functionally important residues (Vihinen et al. 1994).

#### References.

Bruton OC. Agammaglobulinemia. Pediatrics 9:722-28, 1952.

Lederman HM and Winkelstein JA. X-linked agammaglobulinemia: an analysis of 96 patients. Medicine (Baltimore) 64:145-156, 1985.

Sideras P and Smith CIE. Molecular and cellular aspects of X-linked agammaglobulinemia. Adv Immunol 1995 in press.

Smith CIE, Islam KB, Vorechovsky I, Olerup O, Wallin E, Rabbani H, Baskin B. Hammarström, L. X-linked agammaglobulinemia and other immunoglobulin deficiencies. Immunol Rev 138:159-183, 1994.

Tamagnone L, Lahtinen I, Mustonen T, Virtaneva K, Muscatelli F, Francis F. Alitalo R, Smith CIE, Larsson C, Alitalo K. BMX, a novel cytoplasmic tyrosine knase gene of the BTK/ITK/TEC family located in chromosome Xp22.2. Oncogene  $\mathfrak{S}:3683-3688$ , 1994.

Thomas JD, Sideras P, Smith CIE, Vorechovsky I, Chapman V, Paul W.E. A missense mutation in the X-linked agammaglobulinemia gene colocalizes with the mouse X-linked immunodeficiency gene. **Science** 261:355-358, 1993.

Tsukada S, Saffran DC, Rawlings DJ, Parolini O, Allen RC, Klisak I, Sparkes RS, Kubagawa H, Mohandas T, Quan S, Belmont JW, Cooper MD, Conley ME, Witte ON. Deficient expression of a B cell cytoplasmic tyrosine kinase in human Xlinked agammaglobulinemia. **Cell** 72:279-290, 1993.

Vetrie D, Vorechovsky I, Sideras P, Holland J, Davies A, Flinter F, Hammarström L, Kinnon C, Levinsky R, Bobrow M. Smith CIE, Bentley DR. The gene involved in X-linked agammaglobulinemia is a member of the src family of protein-tyrosine kinases. Nature 361: 226-233, 1993.

Vihinen M, Vetrie D, Maniar HS, Ochs HD, Zhu Q. Vorechovsky I, Webster ADB, Notarangelo LD, Nilsson L, Sowadski JM, Smith CIE. Structural basis for chromosome X-linked agammaglobulinemia: A tyrosine kinase disease. **Proc Natl Acad Sci USA** 91: 12803-12807, 1994.

Vorechovsky I, Vetrie D, Holland J, Bentley DR, Thomas K, Zhou J-N, Notarangelo LD, Plebani A, Fontan G, Ochs HD, Hammarström L, Sideras P, Smith CIE. Isolation of cosmid and cDNA clones in the region surrounding the *BTK* gene at Xq21.3-q22. Genomics 21: 517-524,1994.

#### UNEQUAL CROSSING OVER AS SOURCE OF IGHC LOCUS VARIABILITY

#### Angelo Oscar Carbonara Alfredo Brusco

Dipartimento di Genetica Biologia e Chimica Medica and Centro CNR Immunogenetica ed Oncologia Sperimentale, Università di Torino, Italy

The Immunoglobulin Heavy chain Constant region (IGHC) genes are a multigene family located on chromosome 14 and organised in a 350-kb region as follows: M-D, G3-G1-EP1-A1, GP, G2-G4-E-A2. Their high reciprocal homology leads through meiotic mispairings to deleted and duplicated haplotypes.

The frequencies of these haplotypes in 668 healthy blood donors from 5 different Italian regions were determined by Pulsed Field Gel Electrophoresis and the genes involved in the rearrangements were characterised by Southern blotting with several IGHC probes and by densitometric evaluation of the films. 20 deleted (1.50%) and 60 duplicated (4.50%) haplotypes were identified, all in heterozygous individuals except for two homozygous deletions. The deletions/duplications involved one or more genes. GP-A2, A1-E and G4 duplications, and A1-E and GP-A2 deletions were the most common. In the same survey 5 triplicated haplotypes (two of them spanning from the A1 to E genes) were also found. All the rearranged haplotypes seem to be the result of unequal crossing-over. The difference between the number of duplications and deletions was significant in Sardinia, Lombardy, Puglia and in the total of 668 subjects (p<0.001). The crossing over events should produce an equal number of deleted and duplicated haplotypes. Several mechanisms can be postulated for this imbalance: i) negative selection against deleted haplotypes; ii) positive selection for duplications; iii) genetic drift.

#### Ig A deficiency. Clinical and immunological heterogeneity

Ig A deficiency (Ig A-D) is the most common primary immunodeficiency (PID). These patients form a very heterogeneous group ranging from asymptomatic cases to those presenting allergic manifestations, repeated infections, or autoimmune or malignant diseases. In some cases Ig A-D is associated with Ig G2 deficiency and may benefit from G. Globulin therapy Follow-up is lacking in most published series and very little is known of Ig A -D patients who develop more profound immunodeficiency during their life time.

Common variable immunodeficiency (CVID) is a PID diagnosed in most cases in young adults. Delayed diagnosis is very common and it usually takes some years for correct therapy to be established. The evolution of 1g levels over the years before definitive diagnosis is made is usually not reported and we do not know whether an Ig A-D was previously present.

Autoimmune diseases was diagnosed in 8.3 % of our series of 144 symptomatic patients.

The mothers of two girls have Ig A-D, and SLE was diagnosed in one of them.

Three cases of Ig A-D were diagnosed at 7, 8 and 14 years of age, two of them presented with autoimmune phenomena in two cases (chronic active hepatitis and SLE-like disease) and the third with repeated infections in the third All three were diagnosed of CVID 1, 6 and 12 years later, respectively. G.Globulin has been established with correction of the autoimmune and infectious manifestations (including the negativization of autoantibodies).

The main immunological abnormalities in these three cases are : Ig G2 deficiency, low CD4 number in the second case ( with high ANA and anti-DNA antibodies), normal lymphocyte response to mitogens and anti-CD3.

CD40L expression in stimulated T-cells was also studied in those patients and other CVID and was normal in the majority.

Ig A-D and CVID may represent polar ends of the spectrum reflecting one underlying genetic defect. Genetic and immunological studies of symptomatic Ig A-D cases and closer follow-up will be needed to undorctand the eaucee and immunological evolution of these two common diseases.

T. Español Immunology Unit C.S.Valle Hebron Barcelona

### THIRD SESSION: GENE THERAPY

Chairman: Maxime Seligmann

# T Lymphocyte Gene Therapy for ADA Deficiency (SCID): Results of the initial trial with 4 years observation.

R. Michael Blaese, Kenneth Culver, Charles Carter, Harvey Klein, Melvin Berger, Craig Mullen, Jay Ramsey, Linda Muul, Steven Rosenberg, Thomas Fleisher, Gene Shearer, Mario Clerici, Richard Morgan, A Dusty Miller, and W. French Anderson.

Two girls with severe combined immunodeficiency caused by an autosomal recessive defect in the gene for adenosine deaminase have been treated by retroviral-mediated gene therapy using periodic infusions of gene-corrected autologous T lymphocytes. Neither child had a HLA-matched sibling donor for bone marrow transplantation and both had objective evidence of incomplete immune reconstitution with PEG-ADA enzyme replacement therapy. Autologous peripheral blood T cells were obtained by apheresis, the T cells were induced to proliferate by stimulation with the anti-T cell receptor monoclonal antibody OKT3 and interleukin 2, and the induced proliferating T cells were then transduced by repeated exposures to supernatants containing the LASN retroviral vector. The T cells were expanded 20-100 fold in culture over 9-12 days and were then reinfused into the patients without selection for transgene expressing cells. The period of culture expansion was brief to maintain polyclonality of the T cell repertoire in each cell population. Each child received 11-12 infusions of these cultured, gene-modified cells during the first 23 months of the protocol and have then been under close observation for an additional 2 years to evaluate the overall effects of the treatment. Patient 1 has experienced restoration of total T cell numbers to the normal range. The concentration of ADA enzyme activity in her circulating lymphocytes has risen to roughly half heterozygote levels with an LASN vector DNA signal on Southern analysis equivalent to 0.5 to 1 copy of vector per circulating mononuclear cell. Anergic prior to beginning this treatment protocol, she acquired normal delayed hypersensitivity skin test reactivity, experienced reconstitution of cytokine production and cytolytic T cell activity, and enhanced antibody (isohemagglutinin) production during the first months of this treatment. In addition, she is reported to have experienced fewer episodes of infectious diseases requiring antibiotic treatment and has been able to regularly attend public school. The elevated peripheral blood T cell count and increased level Instituto Juan March (Madrid) of ADA in the cells from patient 1 have persisted for the duration of this observation period demonstrating an unexpectedly long life span for peripheral T cells and showing that silencing of the integrated LASN vector has not been sufficient to eliminate therapeutic effectiveness.

Patient 2 had a more limited and variable immune deficiency before beginning the protocol and evaluation of her response is less definitive. She also experienced normalized total T cell counts and restoration of normal delayed hypersensitivity skin test reactivity. Isohemagglutinins rose from barely detectable to the normal range within 3 months of initiation of therapy. Cytolytic T cell function to influenza improved following the start of gene therapy while responses to allogeneic cells spontaneously improved before this treatment. The efficiency of gene transfer into the T cells from patient 2 was only about 10% of that achieved in patient 1 and her T cells underwent senescence in culture at day 13-16 compared to day 30+ for patient 1. These differences in intrinsic T cell behavior and proliferative potential may explain the presence of only 0.1 to 1% LASN vector containing cells in the circulating blood of this patient with lymphocyte levels of ADA consistent with this limited number of genecorrected cells. The elevated peripheral blood T cells in patient 2 returned to pretreatment levels after about one year. There have been no significant side effects associated with the treatments and no evidence of the appearance of recombinant replication competent retrovirus in either child. We conclude that although much remains to be learned, infusion of culture-expanded autologous peripheral blood T cells modified by retroviral-mediated ADA gene-transfer can be a safe and effective addition to the therapy for this severe primary immunodeficiency disease.

- Blaese RM, Anderson WF, Culver KW. The ADA human gene therapy clinical protocol. Human Gene Therapy 1990;1:327-62.
- Blaese RM, Culver KW. Gene therapy for Primary Immunodeficiency. Immunodeficiency Reviews 1992; 3, 329-349
- Blaese RM. Development of gene therapy for immunodeficiency: Adenosine deaminase deficiency. *Pediatric Research* 1993; 33 (suppl), S49-S55

48

#### Thymic selection

#### Harald von Boehmer Basel Institute for Immunology Grenzacherstrasse 487, CH-4005 Basel, Switzerland

T lymphopoiesis is tightly controlled by  $\alpha$  and  $\beta$  T cell receptor (TCR) genes that rearrange in temporal order and form first the pre-TCR and then the  $\alpha\beta$  TCR. Both receptors associate with signal transducing CD3 molecules. Signaling by both, the pre-TCR as well as the  $\alpha\beta$  TCR is essential for T cell development as evidenced by the fact that defective genes which prevent formation of these receptors or interfere with the signaling cascade lead to an arrest in T cell development. The pre-TCR consists of the TCR  $\beta$  chain in disulfide linkage to the pre-TCR  $\alpha$  (pT $\alpha$ ) chain. Its function is to induce extensive cell proliferation of early thymocytes that have succeeded in the formation of a productive TCR  $\beta$  gene and is accompanied by allelic exclusion of the TCR  $\beta$  locus, CD4/8 coreceptor expression and TCR  $\alpha$  rearrangement as well as other subtile changes that generate shortlived CD4+8+ thymocytes. Signaling by the pre-TCR requires CDe but not CD3  $\delta$  signal transducing proteins and the p56<sup>lck</sup> kinase plays a major role in signal transduction. The pTa gene is expressed in immature but not mature T cells including pro-T cells in fetal blood as well as immature T cells in sites of extrathymic T cell development.

The  $\alpha\beta$  TCR must bind to class I or II MHC molecules to rescue immature T cells from programmed cell death. This binding also decides whether the rescued cells become CD4<sup>+</sup> helper or CD8<sup>+</sup> killer cells. The correlation of class MHC restriction and functional phenotype does not always hold and possible reasons for this will be discussed.

### FOURTH SESSION: OTHER IMMUNODEFICIENCIES

Chairman: Maxime Seligmann

#### Virus infection models affecting CD3 activation

Alfredo Corell, Inmunología Hospital Universitario 12 de Octubre, 28041 MADRID

Secondary immunodeficiencies are a heterogeneous group of disorders in which the immune alteration is not the primary defect but the result of several pathologic conditions such as viral infections, atopy, cancer, etc... Although the molecular mechanisms leading to immunodeficiency are mostly unknown, some of them have been associated with T-cell activation defects though several membrane molecules. Moreover, the complete knowledge of the molecular mechanisms generating the immune defect could led us to a better understanding of the immune normal physiology.

Infections by some viruses have been associated with selective TCR/CD3 activation defects: human immunodeficiency virus (HIV), Epstein-Barr virus (EBV) and cytomegalovirus (CMV). The immunological characteristics of individuals infected by EBV and CMV will be presented.

EBV infection causes T-lymphocyte anergy probably due to a selective and proximal TCR/CD3 signal transduction defect resulting in a low proliferation, IL-2 production and IL-2 receptor expression. It is believed that a viral homologue of IL-10 may be involved in EBV-induced immunosuppression.

CMV infection causes also a selective T-lymphocyte anergy, at least "in vitro". Peripheral mononuclear cells from CMV infected patients showed a low proliferation to IL-2 synthesis in response to bacterial antigens, anti-CD3 monoclonal antibodies and some lectins whereas stimulation through TCR/CD3 independent pathways was normal.

The putative molecular mechanisms involved in these two secondary immunodeficiencies will be discussed.

#### Angel L. Corbí Instituto de Parasitología López-Neyra, CSIC

Cellular adhesion plays an essential role in the onset and regulation of immune and inflammatory processes. Most leukocyte adhesive functions are absolutely dependent on the expression and function of the leukocyte-specific heterodimers LFA-1 (CD11a/CD18), Mac-1 (CD11b/CD18), and p150,95 (CD11c/CD18), which comprise the B2 integrin subfamily (1). Their essential role in leukocyte binding to and migration through the endothelial layer in inflamed tissues is illustrated by the existence of an immunodeficiency disorder termed Leukocyte Adhesion Deficiency (LAD). LAD is an autosomal recessive disease caused by the deficient expression of the three B2 integrins on the cell surface of leukocytes (1,2). LAD patients exhibit delayed umbilical cord separation. impaired pus formation, and recurrent bacterial and fungal infections of soft tissues, as a consequence of the lack of neutrophil migration into sites of inflammation (1-4). Two clinical phenotypes of LAD (Severe and Moderate or Partial), which normally correlate with the severity of the disease, have been defined according to the level of B2 integrin expression on patient leukocytes (3). LAD is originated by heterogeneous mutations within the common B2 (CD18) subunit gene (5,6), which is located on chromosome 21g22 (7). Several types of LAD have been defined (I-V) based on the size and levels of the CD18 subunit precursor, the CD18 messenger RNA, and the resulting phenotype (5). Analysis of Severe and Moderate LAD CD18 alleles has identified aberrant splicing events (8-11), missense mutations (9-15), and a 1 bp deletion (14) within the structural region of the CD18 gene.

We have investigated the molecular basis of the disease in two unrelated Severe patients (HS and ZJO)(16). Both patients share a complete absence of CD18 protein precursor and cell surface expression, but they differ in the level of CD18 mRNA, which is normal in HS and undetectable by Northern blot in ZJO. Determination of the primary structure of the patient HS CD18 mRNA revealed a 10 bp deletion between nucleotides 190-200 (CD18 exon 3), which eliminates residues 41-43 and causes a frameshift into a premature termination codon 17 bp downstream from the deleted region. The 10 bp frameshift deletion maps to a region of the CD18 gene where aberrant mRNA processing has been detected in HS and two other unrelated LAD patients. In the ZJO patient, amplification of lymphoblast CD18 mRNA demonstrated the presence of a non-sense mutation in the third nucleotide of the triplet encoding <sup>534</sup>Cys (TGC-TGA), within exon 12. Both genetic abnormalities were also detected at the genomic level, and affect the restriction pattern of their corresponding genes, thus enabling the detection of the mutant alleles among healthy heterozygous in family studies. The identification of two new LAD CD18 alleles, either carrying a non-sense mutation (ZJO) or a partial gene deletion (HS), expand the repertoire of genetic abnormalities causing LAD and further indicate that the heterogeneity of the molecular basis of LAD is comparable to that of other genetic diseases.

Integrin functional activity is absolutely dependent on the state of cellular activation and differentiation (17). To be completely effective, the ligand-binding activity of integrins must be turned "on" and "off" in a very precise manner and during a brief period of time (e.g. during bleeding)(17). Such a rapid regulation of the integrin functional activity takes place on the plasma membrane, and cellular agonists rapidly modulate the integrin ligand binding properties, in a process that has been termed "inside-out" signalling (17). The regulation of affinity for ligands has now been demonstrated for numerous integrins and can be accomplished *in vitro* by cell stimulation with phorbol esters, by cross-linking certain cell surface receptors, or by treatment with activating monoclonal antibodies (17). To study the influence of the cellular environment on the leukocyte integrins functional activity and analyze their involvement in hematopoitic cell differentiation, we have developped stable stable transfectants of the three 62 integrins in cultured cell lines whose differentiation can be induced *in vitro*. Like on circulating leukocytes, the integrins expressed on U937 or K562 cells were expressed in a constitutively inactive state, as demonstrated by the lack of adhesion to their cellular counterreceptors or soluble ligands, the absence of CD18-dependent

intercellular aggregation and their inability to mediate adhesion to protein-coated plates. However, while B2 integrin adhesive functions in U937 cells could be induced upon cellular activation, their function in K562 cells was unaltered by cellular agonists and could only be upregulated with activating monoclonal antibodies. Unlike the B2 subfamily, B1 integrins expressed on K562 cells acquired the high affinity ligand-binding state during cellular activation, threfore suggesting that K562 cells are defective in a putative B2-specific component of the inside-out signalling parthway. This suggestion raises also the possibility of the existence of alternative LAD patients whose molecular basis would map to the B2-specific component of the cellular "inside-out" signalling pathway. The acquisition of CD11b/CD18-dependent adhesive functions in *c-fes*-transfected K562 cells (18) would indicate the involvement of a tyrosine kinase in this putative B2-specific inside-out signalling pathway.

#### REFERENCES

- 1.- Kishimoto, T.K., Larson, R.S., Corbi, A.L., Dustin, M.L., Staunton, D.E., Springer, T.A. 1989. Adv. Immunol. 46:149-182.
- 2.- Arnaout, M.A. 1990. Immunol. Rev. 114:145-180.
- Anderson, D.C., F.C. Schmalstieg, M.J. Finegold, B.J. Hughes, R. Rothlein, L.J. Miller, S. Kohl, M.F. Tosi, R.L. Jacobs, T.C. Waldrop, A.S. Goldman, W.T. Shearer, and T.A. Springer. 1985. J. Infect. Dis. 152:668-689.
- Fischer, A., B. Lisowska-Grospierre, D.C. Anderson, and T.A. Springer. 1988. Immunodef. Rev. 1:39-54.
- 5.- Kishimoto, T.K., N. Hollander, T.M. Roberts, D.C. Anderson, and T.A. Springer. 1987. Cell. 50:193-202.
- 6.- Dimanche, M.T., A. Guyot, G. De Saint-Basile, A. Fischer, C. Griscelli, and B. Lisowska-Grospierre. 1988. Eur. J. Immunol. 18:1575-1579.
- Corbí, A.L., R.S. Larson, T.K. Kishimoto, T.A. Springer, and C.C. Morton. 1988. J. Exp. Med. 167:1597-1607.
- 8.- Kishimoto, T.K., K. O'Connor, and T.A. Springer. 1989. J. Biol. Chem. 264:3588-3595.
- 9.- Nelson, C., H. Rabb, and M.A. Arnaout. 1992. J. Biol. Chem. 267:3351-3357.
- 10.- Back, A.L., W.W. Kwok, and D.D. Hickstein. 1992. J. Biol. Chem. 267:5482-5487.
- Matsuura, S., F. Kishi, M. Tsukahara, H. Nunoi, I. Matsuda, K. Kobayashi, and T. Kajii. 1992. Biochem. Biophys. Res. Commun. 184:1460-1467.
- 12.- Arnaout, M.A., N. Dana, S.K. Gupta, D.G. Tenen, and D.F. Fathallah. 1990. J. Clin. Invest. 85:977-981.
- 13.- Wardlaw, A.J., M.L. Hibbs, S.A. Stacker, and T.A. Springer. 1990. J. Exp. Med. 172:335-345.
- 14.- Sligh, J.E., M.Y. Hurwitz, C. Zhu, D.C. Anderson, and A.L. Beaudet. 1992. J. Biol. Chem. 267:714-718.
- 15.- Corbí, A.L., A. Vara, A. Ursa, M.C. García-Rodriguez, G. Fontan, and F. Sánchez-Madrid. 1992. Eur. J. Immunol. 22:1877-1881.
- 16.- López-Rodriguez, C., Nueda A., Grospierre, B., Sánchez-Madrid, F., Fischer, A., Springer, T.A., and Corbí, A.L. 1993. *Eur. J. Immunol.*, 23:2792-2798.
- 17.- Hynes, R.O. Cell 1992. 69:11-25.
- 18.- Yu, G., Smithgall, T.E., and Glazer, R.I. J. Biol. Chem. 1989. 264:10276-10281.

#### ATAXIA-TELANGIECTASIA: PATHOGENESIS AND POSITIONAL CLONING. RA Gatti.

UCLA School of Medicine, Department of Pathology, Los Angeles, CA, USA.

Several lines of evidence are converging to produce new insights into the cancer susceptibility of ataxia-telangiectasia (A-T) patients and heterozygotes. Defective or damaged DNA is tumorigenic and can result from inherited or somatic changes. Inherited defects in mismatch repair genes that maintain the integrity of DNA have recently been identified in colon cancer families (1-3). Cancer-associated mutations of p53, Rb and p16, all part of a pathway leading to G1 cell cycle arrest and the time needed to repair DNA damage from metabolic processes, have also been identified. Ataxia-telangiectasia (A-T) cells do not respond normally to ionizing radiation, showing a poor p53 response, a foreshortened G1/S delay, and an accumulation of cells in G2/M (4-6). These cell cycle transition defects are believed to result in the replication of poorly edited DNA. This new understanding may also explain the 38% incidence of cancer in A-T homozygotes and the 5-fold increased risk of breast cancer in female A-T heterozygotes (7).

In addition to our studies characterizing the cell cycle traverse in A-T cells (8-11), we are attempting to isolate the major A-T gene by positional cloning (12-15). Linkage analysis and genotyping data on 176 families in the A-T International Consortium database localize the major A-T gene to a "500 kb region, between the markers A4(S1819) and A2(S1818). They localize the Group A A-T gene (ATA) to a region that contains the "500 kb region. The region of the Group C gene also contains the "500 kb region (6). The data are compatible with linking Groups D and E as well as ATFresno to 11q22-23, but not NBS-V1 or NBS-V2 (6,16), suggesting the presence of other loci, not at chromosome 11q22-23, for the latter two rare clinical variants. Four non-linking but classical A-T families contain only single affecteds, leaving open the possibility that these may be spontaneous mutations rather than evidence for another classical A-T gene outside the 11q22-23 region. On the other hand, the 11q22-23 region may contain more than a single A-T gene within it.

Concannon et al. have recently identified a 42 kb cosmid from the candidate region which corrects the radioresistant DNA synthesis of A-T Group A fibroblasts, but not fibroblasts from patients in Groups C or D (17). In collaboration with this laboratory, we are isolating cDNAs from the region so as to sequence them and screen patient mRNAs and DNAs for mutations, insertions or deletions. We have already characterized one gene of approximately 18 kb; we have sequenced the entire cDNA (4.3 kb) but find no mutations in over 80 patients. We have also translated its protein, using cDNA templates from 13 patients, and find no new or truncated proteins. We have defined the genomic boundary of that gene and are subcloning the remaining 24 kb of the cosmid to establish a better genomic map. Meanwhile, others in the lab are isolating cDNAs and GC-rich genomic clones from the 24 kb region, which we are screening for mutations. If these efforts fail to identify an A-T gene, we will return to our analyses of the larger "500 kb candidate region.

- 1. Fishel et al. Cell 75: 1027-1038, 1993.
- 2. Nicolaides et al. Nature 371: 75-80, 1994.
- 3. Nystrom-Lahti et al. Am J Hum Genet 55: 659-665, 1994.
- Kastan et al. Cell 71: 587-597, 1992.
- 5. Lavin et al. Intl J Radiat Biol. November 1994.
- 6. Gatti et al. Cancer Res. 54:6007-6010, 1994.
- Swift et al. New Engl. J. Med. 325: 1831-6, 1991.
- 8. Huo et al. Cancer Res 54: 2544-2547, 1994.

HANN NA POLICIAN

- 9. Naelm et al. Mod Path 7: 587-592, 1994.
- 10. Hong et al. Radiat Res 140: 17, 1994.
- 11. Skog et al. Cancer Detect Prevent. 1995. 12. Gatti et al. Nature 336, 577-580, 1988.
- 13. Sanal et al, Am J Hum Genet 47: 860-66, 1990.
- 14. Foroud et al. Amer J Hum Genet. 49: 1263-1279, 1991.
- 15. Gatti et al. Intl J Radiat Biol. November 1994.
- Stumm et al. Eur Soc Hum Genet. Berlin 1995. Abstract.
   Concannon et al. V(D)J Recombination, DNA Repair, & Hypermutation. Birmingham, 1994. Abstract.

TWO EARLY PHASES OF LYMPHOHEMATOPOIESIS IN THE MOUSE EMBRYO. M.A.R. Marcos and S.G. Copin, Centro de Biologia Molecular S.O., CSIC-UAM, Madrid, Spain; I. Godin and F. Dieterlen-L., Institut d'Embryologie, Nogent-sur-Marne, France; S. Morales and M.L. Gaspar, Instituto Carlos III, Madrid, Spain.

Various inducing microenvironments support hematopoiesis throughout mouse lifespan, from the earliest embryo sites to the adult BM.

Multipotent hematopoietic stem cells (HSCs) are detected in paraaortic splanchnopleura SPL/AGM, yolk sac (YS), blood and liver in early (8,5-12 days) mouse embryos. The most immature progenitors intrinsically differ from adult BM HSCs in that they are bigger in cell size, Sca-1 (Ly6A), and AA4.1+c-Kit+. On the basis of timing of appearance, kinetics of Ag expression and decrease in cell size, the following ontogenical sequence of cell populations has been characterized: A) c-Kit+AA4.1-Mac-1- --> B) c-Kit+AA4.1lowMac-1- -->C) c-Kit+AA4.1lowMac--- D) c-Kit+AA4.1highMac-1high, In organs of the same low developmental age, the most mature phenotype is present in SPL/AGM, followed by blood/YS, and being the liver the most delayed site in terms of hematopoietic cell differentiation. Studies of quantitative RT-PCR reveal a rather ubiquituous expression of c-Kit and CD34 genes in the 9-12 days embryo. Recombinase-encoding RAG-1 gene is firstly expressed in SPL/AGM between days 9.5-11, being delayed 0.5 days its expression in the YS. In the liver, RAG-1 gene expression is detected for the first time at day 12 of gestation, as well as in simultaneous blood and omentum. Pre-B cell specific  $\lambda 5$  gene transcription only shows up in day 13-liver (sensitivity levels of the assay: 1-10 cells). Genomic IgH DJ rearrangements are, however, detected from day 9.5 on, in the mouse embryo. Two clear-cut patterns of DJH usage are observed: 1) Predominant usage of J4 in SPL/AGM, YS, blood, omentum and pre-day 12,5 liver, and 2) Randomized utilization of the four Js in post-day 13 liver.

We conclude that emergent embryonic hematopoiesis proceeds through site-specific, ontogenical pathways and speeds of maturation. Early B-lymphoid generation do not pass through a  $\lambda 5$ -encoded pre-B cell receptor-expressing stage.

### POSTERS

### Structure/function analysis of the tyrosine kinase pp56<sup>1ck</sup> involved in the establishment of immunocompetence.

### Ana C. Carrera<sup>1,2</sup>, Thomas M. Roberts<sup>2</sup> and Carlos Martinez-A<sup>1</sup>.

 Centro Nacional de Biotecnología, C.S.I.C., Universidad Autonoma Campus de Cantoblanco, Cantoblanco, 28059-Madrid, Spain. 2. Dep. of Cellular and Molecular Biology, Dana Farber Cancer Institute, Harvard Medical School, Boston, MA-02115, USA.

pp56<sup>lck</sup> is a tyrosine kinase mainly expressed in T lymphocytes. pp56<sup>lck</sup> has been shown to be required for several specific T cell processes. Interestingly, mice overexpressing pp56<sup>lck</sup> display a similar immunodeficient phenotype than mice lacking pp56<sup>lck</sup> expression. We have analyzed which subdomains of pp56<sup>lck</sup> are important to yield its enzymatic activity and its biological function. To investigate the functions of these subdomains we constructed and analyzed a collection of chimeras and deletion mutants of pp56<sup>lck</sup>. The conclusions from our studies and their interest for gene therapy will be discussed.

#### CD40L EXPRESSION IN HYPER IGM SYNDROME AND CVID

M. Hernandez, I. Caragol, X. de Gracia, T. Marco, JM. Bertran and T. Español. Immunology Unit, Pediatric Hospital and Pneumology, C.S.Valle Hebron, Barcelona and Pediatric Hospital, Sabadell. Spain.

Absence of the ligand of CD40 molecules (CD40L) in activated T cells has been demonstrated to be the cause of abnormal T-B cell collaboration resulting in Hyper Ig M syndrome (HIGM). The incidence of this genetic abnormality in other humoral Immunodeficiencies is not well known.

We are interested in the role of this molecule in different forms of Hypogammaglobulinemias and non X-linked HIGM. We have therefore studied 4 boys and 2 girls with HIGM syndrome, 12 cases of Common Variable Immunodeficiency (CVID) and normal controls. Three of the boys with HIGM have a classical family history of an X-linked disease and one of the girls, 7 years old, is a congenital rubella syndrome. CVID patients, 10 males and 2 females, were diagnosed at young adult age except 4 boys diagnosed between 2 and 8 years of age. The two female cases were diagnosed of Ig A deficiency some years earlier and controlled because of autoimmune phenomena or repeated infections.

PBL cells were stimulated with PMA and ionomycin, 6-8 h. and labeled with a MoAb anti-CD40L (TRAP-1, provided by Dr. L. Notarangelo) and FITC-goat antimouse (indirect IF) The results were measured by a FACScan cytometer (B-D). CD69 was used as an activation marker control

Absence of CD40L molecules in activated T-cells was detected in the three cases with X-linked forms, but the expression was normal in the fourth boy and the girls. In CVID a few cases had very low expression but the majority had normal or near normal values. Correlation with the number of B cells in peripheral blood, Ig levels and severity of the clinical manifestations will be presented.

Knowledge of the genetic defects will aid understanding these heterogeneous ID and may have prognostic and therapeutic implications.

BamHI AND Pstl POLYMORPHISM OF THE HUMAN CD50 (ICAM-3) GENE. Manel Juan, Agustí Miralles, Jordi Esparza, Maria Cinta Cid, Ramon Vilella, Antoni Gayà, Jordi Vives and Jordi Yagüe.

Servei d'Immunologia. Hospital Clínic, 08036 Barcelona, SPAIN.

The differentiation antigen, CD50 (or ICAM-3), is an adhesion molecule defined as a 120 kD highly-expressed and hematopoetically-restricted leucocyte surface glycoprotein. CD50 seems to be preferentially used by lymphocytes in the LFA-1-mediated binding during the initial interactions of immune recognition and lymphocyte activation, when the other two LFA-1 counter-receptors, CD54 (ICAM-1) and CD102 (ICAM-2) molecules, are not yet up-regulated.

Using a Restriction Fragment Lenght Polymorphism (RFLP) assay, we demostrated two polimorphism in the CD50 genomic map. We have observed a Pstl polymorphism characterized by two bands (4.2 and 4.4 kb). In addition a BamHI polymorphism, defined by two other bands (4.2 and 4.5 kb), was also observed. Both polymorphism are quite informative in the population analysis: 4.2 kb Pstl allele is present in 19% of the population and 4.4 kb Pstl allele is present in 81% of the population, while 4.5 kb BamHI allele is present in 78% of the population and 4.9 kb BamHI allele is present in 22% of the population.

These data open a easy way to analyze both CD50 alleles in heterozygous people (more than 50%, if we use any of both polymorphisms), being possible by this method to study some genetic disorders, as chromosome deletions, where CD50 gen could be involved.

63

# THE HUMAN AUTOSOMAL SCID AND OMENN'S SYNDROME: AN INCREASED RADIOSENSITIVITY WITHOUT DNA REPAIR DEFECT.

Nicolas, N., Cavazzana-Calvo, M., Le Deist, F., De Saint Basile, G., \*Papadopoulo, D., Fischer, A., and De Villartay, J.P. INSERM U132, Hôpital Necker-Enfants Malades, Paris, and \*Institut Curie, Section Biologie, Paris.

About 20% of patients with Severe Combined Immunodeficiency (SCID) present a phenotype characterised by the absence of T and B lymphocytes, together with the presence of NK cells. This SCID phenotype is inherited as autosomal recessive and resembles the seid mutation in mice by several criteria. We analysed the radiosensitivity of marrow GM-CFU and fibroblasts from several patients with T(-) B(-) phenotype, patients with the Omenn immunodeficiency syndrome, patients with X-linked T(-) B(+) SCID, and healthy controls. GM-CFU and fibroblasts from T(-) B(-) SCID and Omenn's showed increased radiosensitivity as measured by survival curves following  $\gamma$ -ray irradiation. This increased radiosensitivity was comparable to that observed for the murine scid.

In order to better understand the qualitative and quantitative aspects of the human SCID defect we went on to analyse the kinetics of DNA repair following double-strand breaks. This was performed using Pulse-Field Gel Electrophoresis (PFGE) on C14-Thymidine labelled fibroblast-DNA and quantification with the PhosphorImager. The CHO mutant xrs6 cell line together with the K1 parental line were used as controls. Only a 40% repair was achieved following 50Gray irradiation of the xrs6 cell line. The defect persisted after 24h and was also demonstrable using 30Gray and 40Gray doses. In contrast, the K1 parental cell line achieved a 90% DNArepair after 2h. Murine scid skin fibroblasts repaired at a much lower rate (50% after 5h) after a 50Gray irradiation; The reparation was however completed after 24h. Finally, neither cells from the T(-) B(-) SCID patients nor from Omenn's showed any defect in the capacity to repair DNA-double strand breaks (90% repair after 3h).

In summary, although the human autosomal SCID patients have an increased radiosensitivity, this defect is clearly different from the DNA-repair abnormality seen in the xrs6 mutant and may also differ from the murine scid mutation.

#### CLINICAL AND IMMUNOLOGICAL STUDY FROM A HYPER-IgM IMMUNODEFICIENCY SYNDROME.

L. Tricas, J. Fetro, A. Suárcz, M<sup>A</sup>, A. Diéguez, Inmunologia, L. Molinos, Neumologia, Hospital Central de Asturias, Oviedo, (Asturias, Spain), J. Rubio and Molins, Medicina Interna, Hospital Jove, Gijón (Asturias, Spain)

The present study emphasizes the clinical and immunological heterogeneity in Hyper-IgM immunodeficiency syndrome, something referred to some years ago. Our patient was first seen at the Hospital General de Asturias in 1978 and a diagnosis of Dysgammaglobulinenia with severe deficits of IgG and IgA and marked elevation of IgM was made on the basis of serum immunoglobulin levels, and recurrent infections of bacterial origin. Histological examination of the lymph nodes was performed but unfortunately no germinal centers could be evaluated.

*Clinical Features:* The patient, a 31 year old man, with Hyper-IgM serum (1250 mg/dl), low IgA (<7 mg/dl) and low IgG (<33 mg/dl), who represents a sporadic case with no similarly affected relatives, either male or female. The clinical history showed the following infections: pneumonia, otitis, sinusitis occurring at an early age, meningitis at age 15 but with no etiological agent being demonstrated, and meningitis at age 30 caused by Neisseria meningitidis. No hematological disturbances (anemia, thrombocitopenia and neutropenia) were present. The patient received no replacement Ig-globulin at the time of inununological study.

Immunological study: Phenotypic analysis of peripheral blood mononuclear cells was performed using monoclonal antibodies from B-D and FITC goat anti human IgM, IgD,(F(ab)'<sub>2</sub> fragment) purchased from Caltag, and Tago respectively. Positive cells: CD3 = 86%, CD4 - 31%, CD8 = 52%, CD19 = 8%, CD20 = 9%. CD20 IgM-bearing cells = 4% and CD20 IgD bearing cells = 7%. Peripheral blood mononuclear cells (PBMC) proliferation assay was induced by mitogens, anti-CD3 monoclonal antibody and alloantigens. PBMC when stimulated with pokewced mitogen *in vitro* secreted IgM exclusively.

Analysis of CD40 ligand expression: CD40 ligand surface expression by activated T cells was made following the activation and staining protocol described in Cell 72, 291-300 (1993). CD401g and an unrelated protein Leu81g were donated by D. Hollenbaugh and A. Aruffo (Bristol-Myers Squibb Pharmaceutical Research Institute, Seattle, USA). Activated T cells from this HIM patient show surface expression of CD40 ligand (30%, MFI = 66) The histogram image is quite different from the normal control

	Normal Control		HIM patient	
	%	MFI	%	MFI
CD3	87	138	76	106
CD4	54	54	40	46
CD40Ig goat anti-human IgG 1/400	50	46	30	66
Lcu81g goat anti-human JgG 1/400	100	6	100	8
CD69	100	118	100	108
Isotype control	100	4	100	7

probably because the percentage of lymphocyte T CD4+ is smaller for the patient.

Preliminary molecular study on CD40 ligand: Total RNA from stimulated lymphocytes was extracted by the guanidium thiocyanate phenol-chloroform method according to Chomczynski et al and subjected to reverse transcriptase PCR (BRL RT) as described in Cell 72, 291-300 (1993). (see poster number 128) Direct sequence of PCR-amplified CD40 ligand cDNA is now being performed. No mutations were found from residue 169 to 247, a sequence where previous studies (Immunological Reviews, 1994, N°138) have placed 30% of the CD40-ligand gene mutants.

#### Conclusions:

We have identified a sporadic presentation of a Hyper-IgM immunodeficiency syndrome in a male patient whose activated T cells show CD40 ligand expression and it does not look like he is not an X-linked form of Hyper-IgM. Sequence of his CD40-ligand is being performed. Other functional studies are being considered.

The Authors thank Dr. D. Hollenbaugh and Dr. A. Aruffo for the kindly donating the CD40Ig and Leu8Ig reagents.

#### A RAPID PCR-SSCP MUTATION ANALYSIS OF THE BRUTON'S TYROSINE KINASE GENE IN X-LINKED AGAMMAGLOBULINAEMIA I. Vořechovský<sup>1</sup>, M. Vihinen<sup>2</sup>, G. de Saint Basile<sup>3</sup>, L. Hammarström<sup>1</sup>, L. Nilsson<sup>2</sup>, A. Fischer<sup>3</sup> and C. I. E. Smith<sup>1</sup> Karolinska Institute at NOVUM, <sup>1</sup>Center for BioTechnology and <sup>2</sup>Center for Structural Biochemistry, Huddinge, Sweden, <sup>3</sup>INSERM U132, Hopital Necker-Enfants-Malades, Paris, France.

The identification of the BTK (Bruton's tyrosine kinase) gene defective in human immunoglobulin deficiency X-linked agammaglobulinaemia (XLA) and characterisation of BTK exon-intron boundaries has now allowed the analysis of mutations and polymorphisms at the level of genomic DNA. Using Southern blot analysis and the polymerase chain reaction single strand conformation polymorphism (PCR-SSCP) assay, amplifying all 19 exons and the putative promoter region with a single annealling temperature, mutations have been identified in 19 out of 24 unrelated patients diagnosed as having XLA. Apart from a large deletion involving exon 19, nine missense (F25S, R288W, I370M, M509V, R525P, N526K, R562W, A582V and G594R), two nonsense (E277X and R525X), five frameshift and two splice site mutations have been found affecting most coding exons and all major enzyme domains. No mutations or polymorphisms were detected in the putative promoter region. A single nucleotide deletion located in the last exon, resulting in a truncation of the eight carboxyterminal residues of Btk and a typical XLA phenotype, indicates structural and/or functional importance of Btk helix I in the catalytic domain. The structural consequences of Btk defects due to deduced amino acid substitutions were analysed with computer-aided molecular modelling. A rapid and sensitive individual mutation analysis of the BTK gene can now be achieved in a one-day procedure.

#### PILOT LINKAGE MAPPING OF SELECTIVE IMMUNOGLOBULIN A DEFICIENCY AND COMMON VARIABLE IMMUNE DEFICIENCY: LACK OF EVIDENCE FOR AN INVOLVEMENT OF THE MHC REGION IN FAMILIES WITH AUTOSOMAL DOMINANT SEGREGATION I. Vořechovský<sup>1</sup>, H. Zetterquist<sup>1,5</sup>, C. I. E. Smith<sup>1,5</sup>, R. Paganelli<sup>2</sup>, S. Koskinen<sup>3</sup>, A.D.B. Webster<sup>4</sup> and L. Hammarström<sup>1,5</sup>

<sup>1</sup>Karolinska Institute at NOVUM, Center for BioTechnology, Huddinge, Sweden; <sup>2</sup>Universita La Sapienza, Rome, Italy; <sup>3</sup>Blood Transfusion Service, Helsinki, Finland; <sup>4</sup>Royal Free Hospital School of Medicine, London, UK; <sup>5</sup>Department of Clinical Immunology, Huddinge Hospital, Huddinge, Sweden.

Screening of close relatives of Swedish patients with selective immunoglobulin A deficiency (IgAD) and common variable immune deficiency (CVID) for serum immunoglobulin levels has identified 32 multiplex families, in 25 of which the disorders segregated as an autosomal dominant trait. A high relative risk for sibs. low phenocopy rate and a permanent phenotype make the two disorders, which have been proposed to represent an allelic condition, amenable to genetic linkage analysis. In a pilot multicentre linkage study, involving the 16 most informative multiplex families with dominant inheritance of IgAD/CVID and comprising a total of 49 affected and 43 unaffected individuals, we have attempted to confirm previously reported genetic linkage of the disease to the major histocompatibility complex (MHC) region. Surprisingly, using both parametric and nonparametric linkage analysis with a series of microsatellite markers at and flanking the MHC region, no evidence for genetic linkage was found (Z = -5.48, -6.39, -3.48, -3.19, -10.57 and -9.50 at zero recombination fractions at D6S291, D6S273. D6S306, D6S464, D6S299 and D6S285, respectively; 0.30 at all lociin an extended sib-pair analysis, ESPA). Negative lod scores were found for 15 out of 16 families using 5 tightly linked polymorphic loci mapped to the MHC region, including PCR-typed polymorphism at codon 57 of DOB1 chain (Z= -3.64 at zero recombination fraction; p=0.63,  $\chi^2=0.11$  in ESPA). In accordance with these results, no evidence for linkage to the MHC region was obtained by computing lod scores in retrospect, based on the previously published segregation data at the MHC region in multiplex families with IgAD/CVID in more than one generation. Assuming a major locus under the dominant model, we conclude that, in contrast to type I diabetes mellitus, the previously reported allelic associations of largely sporadic IgAD/CVID to this region should be explained by reasons other than a presence of closely linked, deficient gene(s) at the MHC region cosegregating with the disease in our families.

### APO-1/Fas (CD95) mediated T cell receptor induced suicide

Jens Dhein<sup>\*</sup>, Henning Walczak<sup>\*</sup>, Caroline Bäumler, Klaus-Michael Debatin, Peter H. Krammer. Tumor Immunology Program, German Cancer Research Centre, Heidelberg, Germany

The APO-1/Fas (CD95) cell surface receptor is a member of the nerve growth factor (NGF)/tumor necrosis factor (TNF) receptor superfamily and mediates apoptosis. Peripheral activated T cells (ATC) from lymphoproliferation (lpr/lpr) mutant mice that express a reduced number of APO-1 have a defect in T cell receptor (TCR)-induced apoptosis. This suggests that TCR-induced apoptosis involves APO-1. We tested this hypothesis in various human T cells: (i) malignant Jurkat cells, (ii) an alloreactive T cell clone (S13), and (iii) peripheral ATC. TCR triggering via immobilized anti-CD3 antibodies or Staphylococcus Enterotoxin B (SEB) superantigen induced expression of the APO-1 ligand and apoptosis in these cells. Anti-CD3-induced apoptosis of Jurkat cells was demonstrated even in single cell cultures. In all cases apoptosis was substantially inhibited by blocking anti-APO-1 antibody fragments [F(ab')2 anti-APO-1] and soluble APO-1 receptor decoys [immunoglobulin fusion proteins (APO-1-Fc)]. The APO-1 ligand was found in the supernatant of activated Jurkat cells as a soluble cytokine. Based on these findings we propose that TCR-induced apoptosis in ATC can occur via an APO-1 ligand mediated autocrine suicide. These results provide a mechanism for suppression of the immune response and for peripheral tolerance by T cell deletion.

· Both authors contributed equally to this work.

### PERSPECTIVES

### A. Fischer and A. Arnaiz-Villena

The general feeling is that the meeting was successful in discussing and gathering the most interesting question on inherited immunodeficiencies.

There were established the genetic defects underlying many immunodeficiencies; the pre- $\alpha$  TCR chain in pre-T cells was identified in humans; several "knock out" mice and their immune function were described and gene therapy for ADA deficiency was show to be effective.

It is inferred that the following questions should be addressed in the near future:

- 1.- To test which deficiencies are susceptible for gene therapy.
- 2.- To do population studies both to calculate the precise carrier numbers and to establish if certain genetic anomalies accompanying deficiencies are not polymorphisms.
- 3.- Population studies are also necessary to explain why some deficiencies (i.e.: IgA) are very common and the possible advantage of the defective genes could also be studied.
- 4.- Many other deficiencies should be studied with the available molecular and genetic methodology.
- 5.- The common but still unexplained relationship between immunodeficiency and autoimmunity should be investigated.
- 6.- A more comprehensive effort to classify inherited (and other) immunodeficiencies should be tried; severe combined immunodeficiency should only be regarded as severe symptomatology.

## List of Invited Speakers

Frederick W. Alt	The Howard Hughes Medical Institute, Children's Hospital, Center for Blood Research and Department of Genetics, Harvard Univ. Medical School, Boston, MA. 02115 (USA). Tel.: 1 617 735 72 90 Fax : 1 617 730 04 32
Antonio Arnaiz-Villena	Inmunología, Hospital Universitario 12 de Octubre, Universidad Complutense, Ctra. Andalucía Km. 5,4, 28041 Madrid (Spain). Tel.: 34 1 390 83 15 34 1 460 58 78 Fax : 34 1 390 83 99
R. Michael Blaese	Clinical Gene Therapy Branch, NIH/NCHGR, Building 49, Room 2A03, 49 Convent Drive, Bethesda, MD. 20892 (USA). Tel.: 1 301 402 18 33 Fax : 1 301 496 71 84
Harald von Boehmer	Basel Institute for Immunology, Grenzacherstrasse 487, CH-4005 Basel (Switzerland). Tel.: 41 61 605 13 27 Fax : 41 61 605 13 72
Angelo Oscar Carbonara	Dipartimento di Genetica Biologia e Chimica Medica and Centro CNR Immunogenetica ed Oncologia Sperimentale, Università di Torino, 10100 Torino (Italy). Tel.: 39 11 633 65 77 Fax : 39 11 696 01 47
Max D. Cooper	University of Alabama at Birmingham and The Howard Hughes Medical Institute, Birmingham, AL. 35294 (USA). Tel.: 1 205 934 33 70 Fax : 1 205 975 72 18
Angel Luis Corbí	Instituto de Parasitología López-Neyra, C.S.I.C. Ventanilla 11, 18001 Granada (Spain). Tel.: 34 58 20 38 02 Fax : 34 58 20 33 23
Alfredo Corell	Inmunología, Hospital Universitario "12 de Octubre," 28041 Madrid (Spain). Tel.: 34 1 390 83 15 Fax : 34 1 390 83 99
Alain Fischer	INSERM U 429 Hôpital Necker-Enfants Malades, 149 Rue de Sèvres, Paris, (France). Tel.: 33 1 444 94 822 Fax : 33 1 444 95 070

R.A. Gatti	UCLA School of Medicine, Department of Pathology, 10883 Le Coute Avenue, Los Angeles, CA. (USA). Tel.: 1 310 825 76 18 Fax : 1 310 206 51 78
Raif S. Geha	Division of Immunology, Children's Hospital, 300 Longwood Avenue, Boston, MA.02115 (USA). Tel.: 1 617 735 76 02 Fax : 1 617 735 82 05
Warren J. Leonard	Laboratory of Molecular Immunology, National Heart, Lung and Blood Institute, NIH, Bethesda, MD. 20817 (USA). Tel.: 1 301 496 00 98 Fax : 1 301 402 09 71
Bernard Mach	University of Geneva Medical School, 9 Av. de Champel, CH- 1211 Geneva (Switzerland). Tel.: 41 22 702 56 61 Fax : 41 22 346 72 37
Tak W. Mak	Ontario Cancer Institute, 500 Sherbourne Street, Toronto, Ontario, M4X 1K9 (Canada). Tel.: 1 416 924 06 71 Fax : 1 416 926 65 88
Hans D. Ochs	Division of Infectious Diseases, Immunology/Rheumatology, Department of Pediatrics, University of Washington School of Medicine, Seattle, WA. 98195 (USA). Tel.: 1 206 543 32 07 Fax : 1 206 543 31 84
José R. Regueiro	Departamento de Inmunología, Facultad de Medicina, Universidad Complutense, 28040 Madrid (Spain). Tel.: 34 1 394 16 42 Fax : 34 1 394 16 41
Fred S. Rosen	Department of Pediatrics, Harvard Medical School, Center for Blood Research, 800 Huntington Avenue, Boston, MA. 02115 (USA). Tel.: 1 617 731 64 70 Fax: 1 617 278 34 93
Geneviève de Saint Basile	Hôpital Necker-Enfants Malades, INSERM U429, 149 Rue de Sèvres, Paris (France). Tel.: 33 1 44 49 50 71 Fax : 33 1 42 73 06 40
C. I. Edvard Smith	Center for Biotechnology, Karolinska Institute, NOVUM,S- 14157 Huddinge (Sweden). Tel.: 46 8 608 91 14 Fax : 46 8 774 55 38
	Instituto Juan March (Madrid)

### List of Participants

Balbino Alarcón	Centro de Biología Molecular Severo Ochoa, CSIC-Universidad Autónoma de Madrid, Cantoblanco, Madrid 28049 (Spain). Tel.: 34 1 397 50 70 Fax : 34 1 397 47 99
Ana C. Carrera	Centro Nacional de Biotecnología, CSIC, Universidad Autónoma Campus de Cantoblanco, 28059 Madrid (Spain) and Dep. of Cellular and Molecular Biology, Dana Farber Cancer Inst., Harvard Medical School, Boston, MA-02115.(U.S.A.) Tel.: 34 1 585 45 30 Fax: 34 1 372 04 93
Antonio Celada	Departamento de Bioquímica y Fisiología (Inmunología), Facultad de Biología, Universidad de Barcelona, Diagonal, 645, 08028 Barcelona (Spain). Tel.: 34 3 402 15 55 Fax : 34 3 411 03 58
Teresa Español	Unidad Inmunología, Institut Català de la Salut, Ciutat Sanitària i Universitària Vall d'Hebron, 08071 Barcelona (Spain). Fax : 34 3 428 04 43
Manuel-Nicolás Fernández	Servicio de Hematología y Hemoterapia, Clínica Puerta de Hierro, Facultad de Medicina, Universidad Autónoma, c/San Martín de Porres 4, 28035 Madrid (Spain). Tel.: 34 1 316 22 40 Fax: 34 1 373 05 35
Richard B. Gallagher	Science International, Thomas House, 14 George IV Street, Cambridge, CB2 1HH (U.K.). Tel.: 44 1223 30 20 67 Fax : 44 1223 30 20 68
Juan Martín García-Sancho	Unidad de Inmunología, Hospital La Paz, Paseo de la Castellana 261, 28046 Madrid (Spain). Fax : 34 1 729 22 80
Abel Gayo Lana	Servicio de Inmunología, Hospital Central de Asturias, c/Celestino Villamil s/n°, 33006 Oviedo (Spain). Tel.: 34 8 510 61 30 Fax : 34 8 525 45 09
Alf Grandien	Lab. 115, Centro Nacional de Biotecnología, CSIC, Campus de Cantoblanco, Ctra. de Colmenar Viejo, Km. 15,500, 28049 Madrid (Spain). Tel.: 34 1 585 45 30 or 585 45 62 Fax : 34 1 585 45 06

Manuel Hernández	Immunology Unit, Pediatric Hospital and Pneumology, C.S. Valle Hebron, 08071 Barcelona and Pediatric Hospital, Sabadell. (Spain). Fax : 34 3 428 04 43
Manel Juan	Servei d'Immunologia, Hospital Clínic, 08036 Barcelona (Spain). Tel.: 34 3 454 49 20 Fax : 34 3 451 80 38
Miguel López-Botet	Sección de Inmunología, Hospital de la Princesa, Universidad Autónoma, Diego de León, 62, 28006 Madrid (Spain). Tel.: 34 1 402 33 47 Fax : 34 1 309 24 96
Miguel A.R. Marcos	Centro de Biología Molecular "Severo Ochoa", CSIC, Facultad de Ciencias, Universidad Autónoma, Cantoblanco, 28049 Madrid (Spain). Tel.: 34 1 397 80 53 Fax : 34 1 397 83 44
Ignacio J. Molina	University of Granada School of Medicine, 18012 Granada (Spain). Tel.: 34 57 21 75 32 Fax : 34 58 24 35 18
Lourdes Mozo	Servicio de Inmunología, Hospital Central de Asturias, c/Julián Clavería s/nº, 33080 Oviedo (Spain). Tel.: 34 8 510 61 30 Fax : 34 8 525 45 09
Nathalie Nicolas	INSERM U132, Hôpital Necker-Enfants Malades, 149 rue de Sèvres, 75 015 Paris (France). Tel.: 33 1 44 49 50 78 Fax : 33 1 42 73 06 40
Manuel Ortíz de Landázuri	Sección de Inmunología, Hospital de la Princesa, Universidad Autónoma, Diego de León, 62, 28006 Madrid (Spain). Tel.: 34 1 402 33 47 Fax : 34 1 309 24 96
Mercedes Pérez Blas	Departamento de Medicina, Universidad de Alcalá, Campus Universitario, Ctra. Madrid-Barcelona Km. 33,600, 28871 Alcalá de Henares, Madrid (Spain). Tel.: 34 1 885 45 26 Fax: 34 1 885 45 26

José Carlos Rodríguez Gallego	Inmunología, Hospital Universitario "12 de Octubre", Universidad Complutense, Ctra. Andalucía Km. 5,4, 28041 Madrid (Spain). Tel.: 34 1 390 83 15 Fax : 34 1 390 83 99
Francisco Sánchez-Madrid	Sección de Inmunología, Hospital de la Princesa, Universidad Autónoma, Diego de León, 62, 28006 Madrid (Spain). Tel.: 34 1 402 33 47 Fax : 34 1 309 24 96
Julia Sequí	Inmunología, Centro de Investigaciones Clínicas, Instituto de Salud Carlos III, Sinesio Delgado 10, 28029 Madrid (Spain). Tel.: 34 1 314 10 10 Fax : 34 1 733 66 14
Maxime Seligmann	Hôpital Saint-Louis, Policlinique d'Hematologie et d'Immunologie, 1 avenue Claude Vellefaux, 75475 Paris (France). Tel.: 33 1 42 49 96 93 Fax : 34 1 42 49 40 40
José Luis Subiza	Immunología, Hospital Universitario "12 de Octubre", Universidad Complutense, Ctra. Andalucía Km. 5,4, 28041 Madrid (Spain). Tel.: 34 1 390 83 15 y 460 58 78 Fax : 34 1 390 83 99
María Luisa Toribio	Centro de Biología Molecular "Severo Ochoa", CSIC, Facultad de Ciencias, Universidad Autónoma, Cantoblanco, 28049 Madrid (Spain). Tel.: 34 1 397 80 76 Fax: 34 1 397 83 44
Lourdes Tricas	Inmunología, Hospital Central de Asturias, c/Julián Clavería s/nº, 33006 Oviedo (Spain). Tel.: 34 8 510 61 30 Ext. 36394 Fax : 34 8 525 45 09
Igor Vořechovský	Karolinska Institute at NOVUM, Center for BioTechnology, 14157 Huddinge (Sweden). Tel.: 46 8 608 91 06 Fax : 46 8 774 55 38
Henning Walczak	Tumor Immunology Program, German Cancer Research Centre, Im Neuenheimer Feld 280, 69120 Heidelberg (Germany). Tel.: 49 6221 423 735 Fax : 49 6221 411 715

### Texts published in the SERIE UNIVERSITARIA by the FUNDACIÓN JUAN MARCH concerning workshops and courses organized within the Plan for International Meetings on Biology (1989-1991)

\*: Out of stock.

- \*246 Workshop on Tolerance: Mechanisms and Implications. Organizers: P. Marrack and C. Martínez-A.
- \*247 Workshop on Pathogenesis-related Proteins in Plants. Organizers: V. Conejero and L. C. Van Loon.
- \*248 Course on DNA Protein Interaction. M. Beato.
- \*249 Workshop on Molecular Diagnosis of Cancer. Organizers: M. Perucho and P. García Barreno.
- \*251 Lecture Course on Approaches to Plant Development. Organizers: P. Puigdomènech and T.

Nelson.

- \*252 Curso Experimental de Electroforesis Bidimensional de Alta Resolución. Organizer: Juan F. Santarén.
- 253 Workshop on Genome Expression and Pathogenesis of Plant RNA Viruses. Organizers: F. García-Arenal and P. Palukaitis.
- 254 Advanced Course on Biochemistry and Genetics of Yeast. Organizers: C. Gancedo, J. M. Gancedo, M. A. Delgado and I. L. Calderón.
- \*255 Workshop on the Reference Points in Evolution. Organizers: P. Alberch and G. A. Dover.

\*256 Workshop on Chromatin Structure and Gene Expression. Organizers: F. Azorín, M. Beato and A. A. Travers.

- 257 Lecture Course on Polyamines as Modulators of Plant Development. Organizers: A. W. Galston and A. F. Tiburcio.
- \*258 Workshop on Flower Development. Organizers: H. Saedler, J. P. Beltrán and J. Paz-Ares.
- \*259 Workshop on Transcription and Replication of Negative Strand RNA Viruses. Organizers: D. Kolakofsky and J. Ortín.
- \*260 Lecture Course on Molecular Biology of the Rhizobium-Legume Symbiosis. Organizer: T. Ruiz-Argüeso.
- 261 Workshop on Regulation of Translation in Animal Virus-Infected Cells. Organizers: N. Sonenberg and L. Carrasco.
- \*263 Lecture Course on the Polymerase Chain Reaction. Organizers: M. Perucho and E. Martínez-Salas.
- \*264 Workshop on Yeast Transport and Energetics. Organizers: A. Rodríguez-Navarro and R. Lagunas.
- 265 Workshop on Adhesion Receptors in the Immune System. Organizers: T. A. Springer and F. Sánchez-Madrid.
- \*266 Workshop on Innovations in Proteases and Their Inhibitors: Fundamental and Applied Aspects. Organizer: F. X. Avilés.

- 267 Workshop on Role of Glycosyl-Phosphatidylinositol in Cell Signalling. Organizers: J. M. Mato and J. Larner.
- 268 Workshop on Salt Tolerance in Microorganisms and Plants: Physiological and Molecular Aspects.

Organizers: R. Serrano and J. A. Pintor-Toro.

269 Workshop on Neural Control of Movement in Vertebrates. Organizers: R. Baker and J. M. Delgado-García.

#### Texts published by the CENTRE FOR INTERNATIONAL MEETINGS ON BIOLOGY

- 1 Workshop on What do Nociceptors Tell the Brain? Organizers: C. Belmonte and F. Cerveró.
- \*2 Workshop on DNA Structure and Protein Recognition. Organizers: A. Klug and J. A. Subirana.
- \*3 Lecture Course on Palaeobiology: Preparing for the Twenty-First Century. Organizers: F. Álvarez and S. Conway Morris.
- \*4 Workshop on the Past and the Future of Zea Mays. Organizers: B. Burr, L. Herrera-Estrella and P. Puigdomènech.
- \*5 Workshop on Structure of the Major Histocompatibility Complex. Organizers: A. Arnaiz-Villena and P. Parham.
- \*6 Workshop on Behavioural Mechanisms in Evolutionary Perspective. Organizers: P. Bateson and M. Gomendio.
- \*7 Workshop on Transcription Initiation in Prokaryotes Organizers: M. Salas and L. B. Rothman-Denes.
- \*8 Workshop on the Diversity of the Immunoglobulin Superfamily. Organizers: A. N. Barclay and J. Vives.
- 9 Workshop on Control of Gene Expression in Yeast. Organizers: C. Gancedo and J. M. Gancedo.

- \*10 Workshop on Engineering Plants Against Pests and Pathogens. Organizers: G. Bruening, F. García-Olmedo and F. Ponz.
- 11 Lecture Course on Conservation and Use of Genetic Resources. Organizers: N. Jouve and M. Pérez de la Vega.
- 12 Workshop on Reverse Genetics of Negative Stranded RNA Viruses. Organizers: G. W. Wertz and J. A. Melero.
- \*13 Workshop on Approaches to Plant Hormone Action Organizers: J. Carbonell and R. L. Jones.
- \*14 Workshop on Frontiers of Alzheimer Disease. Organizers: B. Frangione and J. Ávila.
- \*15 Workshop on Signal Transduction by Growth Factor Receptors with Tyrosine Kinase Activity. Organizers: J. M. Mato and A. Ullrich.
- 16 Workshop on Intra- and Extra-Cellular Signalling in Hematopoiesis. Organizers: E. Donnall Thomas and A. Grañena.
- \*17 Workshop on Cell Recognition During Neuronal Development. Organizers: C. S. Goodman and F. Jiménez.

- 18 Workshop on Molecular Mechanisms of Macrophage Activation. Organizers: C. Nathan and A. Celada.
- Workshop on Viral Evasion of Host Defense Mechanisms.
   Organizers: M. B. Mathews and M. Esteban.
- \*20 Workshop on Genomic Fingerprinting. Organizers: M. McClelland and X. Estivill.
- 21 Workshop on DNA-Drug Interactions. Organizers: K. R. Fox and J. Portugal.
- \*22 Workshop on Molecular Bases of Ion Channel Function. Organizers: R. W. Aldrich and J. López-Barneo.
- \*23 Workshop on Molecular Biology and Ecology of Gene Transfer and Propagation Promoted by Plasmids. Organizers: C. M. Thomas, E. M. H. Wellington, M. Espinosa and R. Díaz Orejas.
- \*24 Workshop on Deterioration, Stability and Regeneration of the Brain During Normal Aging. Organizers: P. D. Coleman, F. Mora and M. Nieto-Sampedro.
- 25 Workshop on Genetic Recombination and Defective Interfering Particles in RNA Viruses. Organizers: J. J. Bujarski, S. Schlesinger and J. Bomero.
- 26 Workshop on Cellular Interactions in the Early Development of the Nervous System of Drosophila. Organizers: J. Modolell and P. Simpson.
- 27 Workshop on Ras, Differentiation and Development. Organizers: J. Downward, E. Santos and D. Martín-Zanca.
- 28 Human and Experimental Skin Carcinogenesis Organizers: A. J. P. Klein-Szanto and M. Quintanilla.
- 29 Workshop on the Biochemistry and Regulation of Programmed Cell Death. Organizers: J. A. Cidlowski, R. H. Horvitz, A. López-Rivas and C. Martínez-A.

- 30 Workshop on Resistance to Viral Infection. Organizers: L. Enjuanes and M. M. C. Lai.
- 31 Workshop on Roles of Growth and Cell Survival Factors in Vertebrate Development Organizers: M. C. Raff and F. de Pablo.
- 32 Workshop on Chromatin Structure and Gene Expression Organizers: F. Azorín, M. Beato and A. P. Wolffe.
- 33 Workshop on Molecular Mechanisms of Synaptic Function Organizers: J. Lerma and P. H. Seeburg.
- 34 Workshop on Computational Approaches in the Analysis and Engineering of Proteins Organizers: F. X. Avilés, M. Billeter and E. Querol.
- 35 Workshop on Signal Transduction Pathways Essential for Yeast Morphogenesis and Cell Integrity. Organizers: M. Snyder and C. Nombela.
- 36 Workshop on Flower Development. Organizers: E. Coen, Zs. Schwarz-Sommer and J. P. Beltrán.
- 37 Workshop on Cellular and Molecular Mechanisms in Behaviour. Organizers: M. Heisenberg and A. Ferrús.

<sup>\*:</sup> Out of stock.

The Centre for International Meetings on Biology was created within the Instituto Juan March de Estudios e Investigaciones, a private foundation specialized in scientific activities which complements the cultural work of the Fundación Juan March.

The Centre endeavours to actively and sistematically promote cooperation among Spanish and foreign scientists working in the field of Biology, through the organization of Workshops, Lecture and Experimental Courses, Seminars, Symposia and the Juan March Lectures on Biology.

> In the last six years, a total of 70 meetings and 6 Juan March Lecture Cycles, all dealing with a wide range of subjects of biological interest, were organized within the scope of the Centre.



Instituto Juan March de Estudios e Investigaciones Castelló, 77 • Teléfono 34 - 1 - 435 42 40 • Fax 576 34 20 28006 Madrid (España)

The lectures summarized in this publication were presented by their authors at a workshop held on the 6th and 7th of March, 1995, at the Instituto Juan March.

All published articles are exact reproduction of author's text.

There is a limited edition of 400 copies of this volume, available free of charge.